

DIFERENCIACIÓN CELULAR

ASPECTOS GENERALES

MECANISMOS CELULARES Y MOLECULARES



Dr. Mario Galindo
Programa de Biología Celular y Molecular
ICBM, Facultad de Medicina
Universidad de Chile

**EL PROCESO DE FORMACION DE ORGANOS Y TEJIDOS
IMPLICA TRES PROCESOS BASICOS
CONTROLADOS POR ESTIMULOS EXTRACELULARES**

PROLIFERACIÓN CELULAR

DIFERENCIACIÓN CELULAR

MUERTE CELULAR

DIFERENCIACIÓN CELULAR

DEFINICIÓN CONCEPTUAL:

**PROCESO MEDIANTE EL CUAL UNA CELULA ADQUIERE
CARACTERISTICAS MORFOLÓGICAS Y FUNCIONALES PARTICULARES**

DEFINICIÓN OPERACIONAL:

COMO PODEMOS DEFINIR UNA CELULA DIFERENCIADA?

**PODEMOS DEFINIR UNA CELULA DIFERENCIADA
EN FORMA OPERACIONAL**

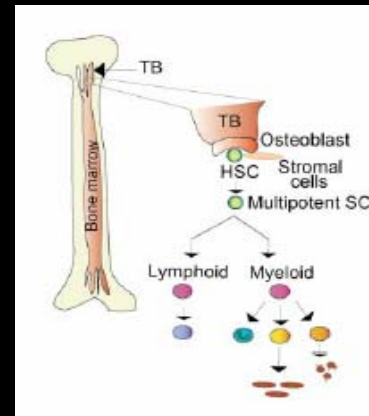
- 1.- CARACTERISTICAS CELULARES O MORFOLÓGICAS**
- 2.- CARACTERISTICAS MOLECULARES
(EXPRESIÓN DIFERENCIAL DE GENES)**

Diferenciación celular

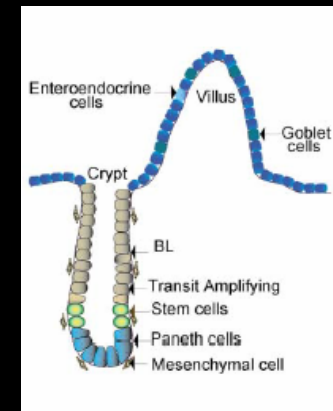
- Desarrollo embrionario y postnatal



- Renovación de tejidos homeostasis

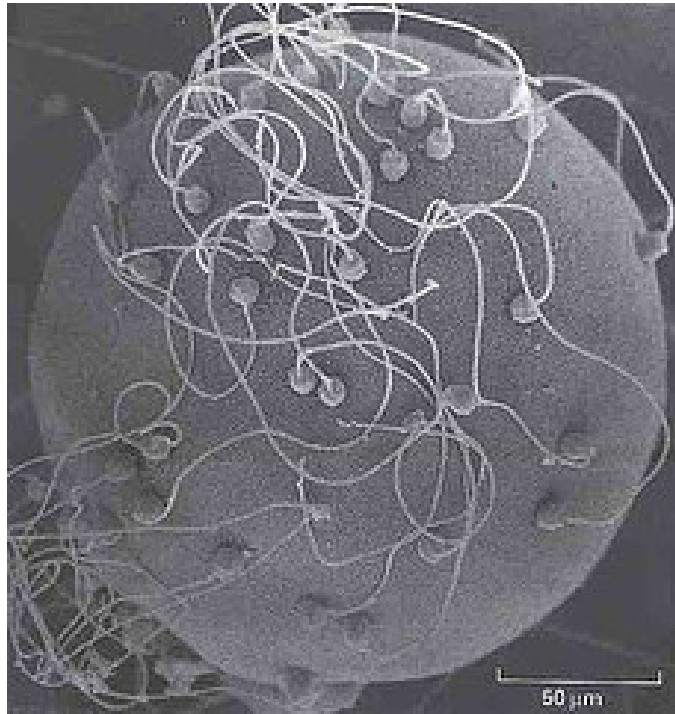


- Reparación de tejidos daño

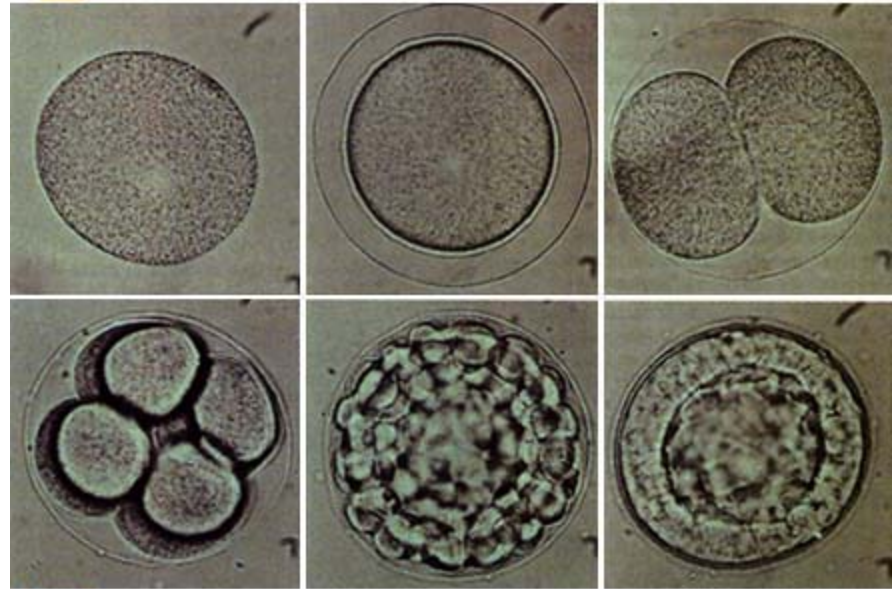


- Ingeniería de tejidos



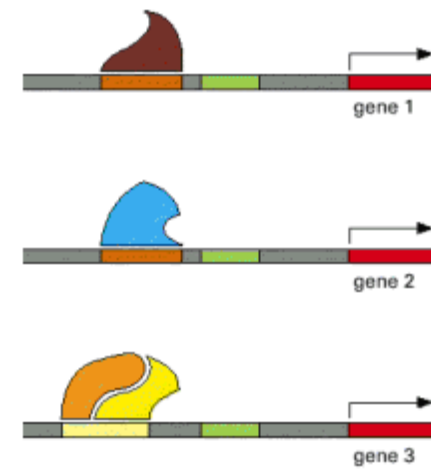
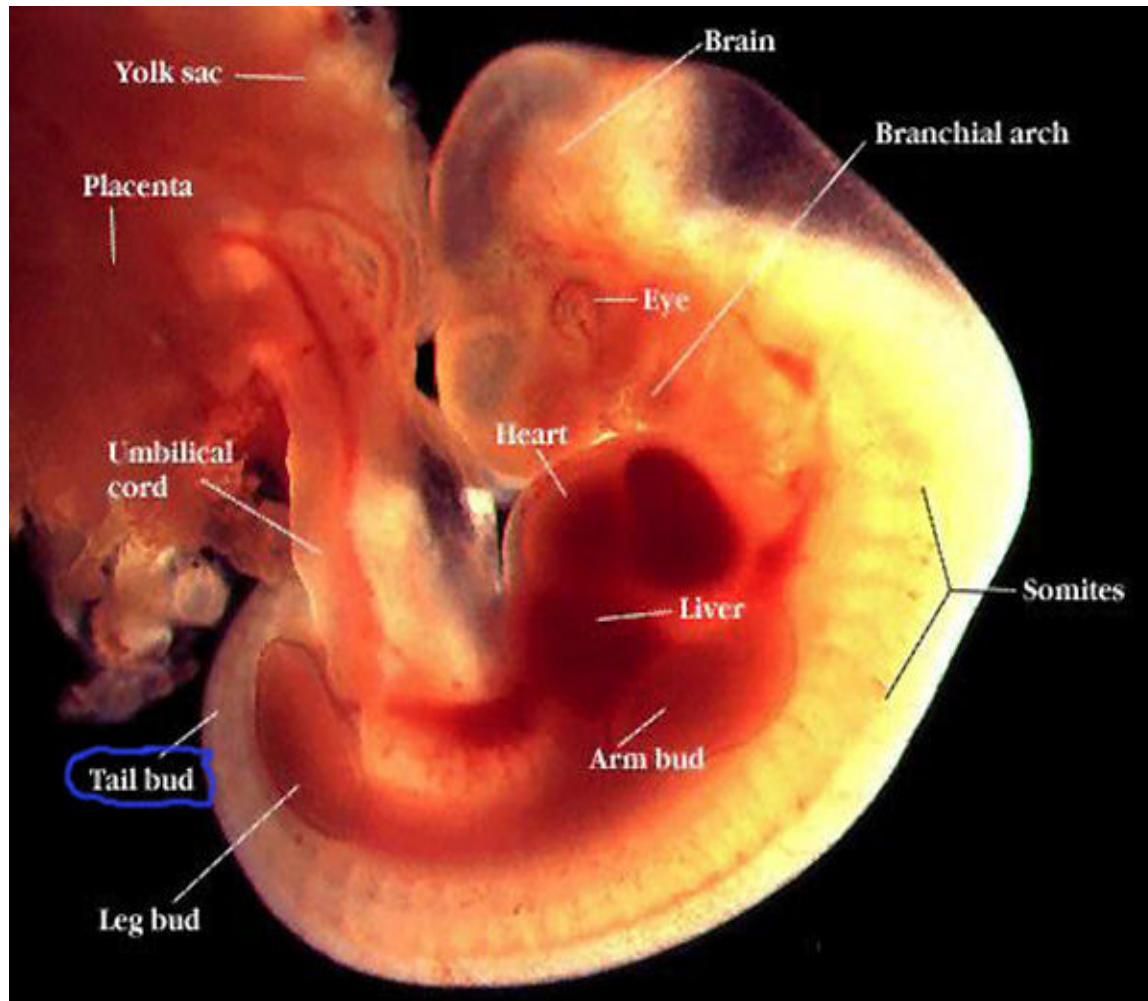


A

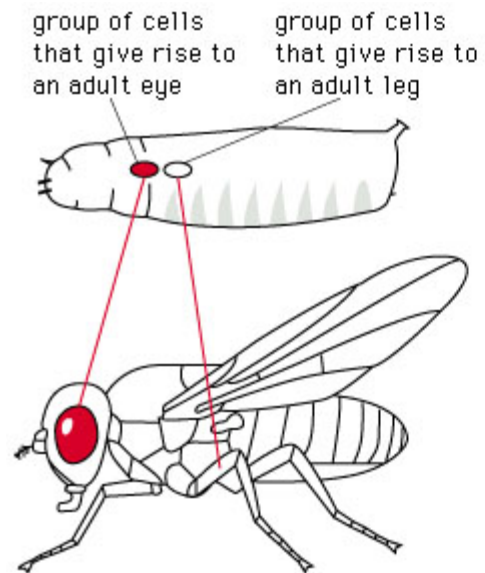


B





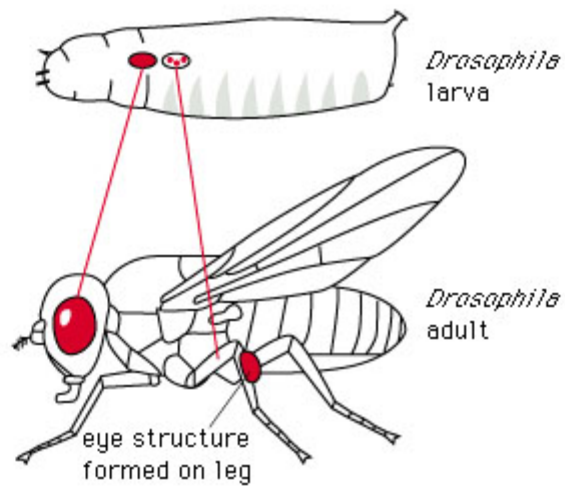
ey



normal fly

(A)

(red shows cells expressing *ey* gene)



fly with *ey* gene artificially expressed in leg precursor cells



(B)



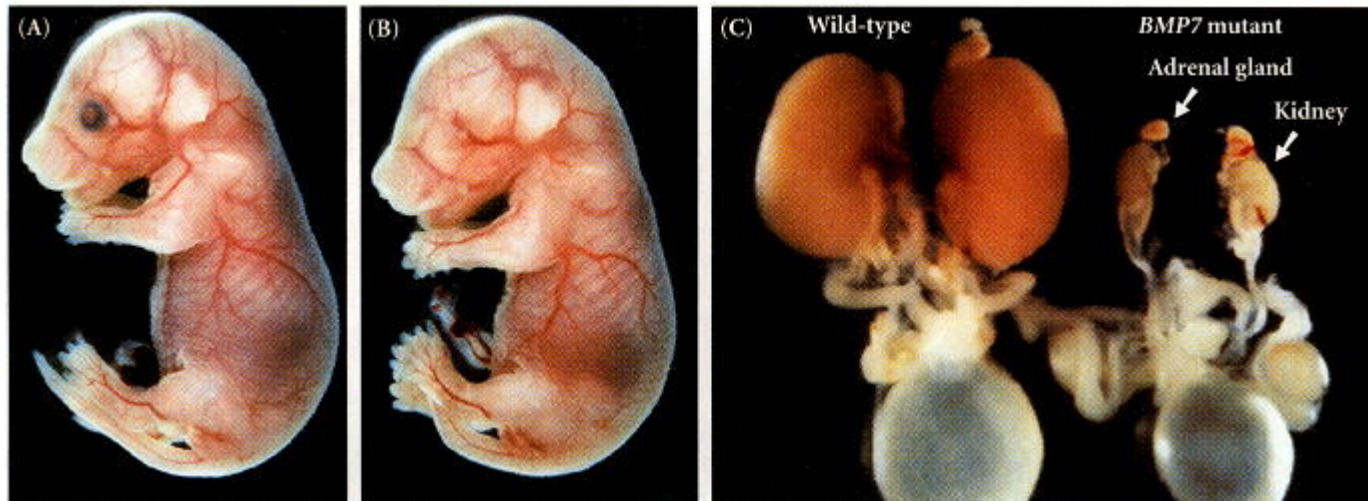
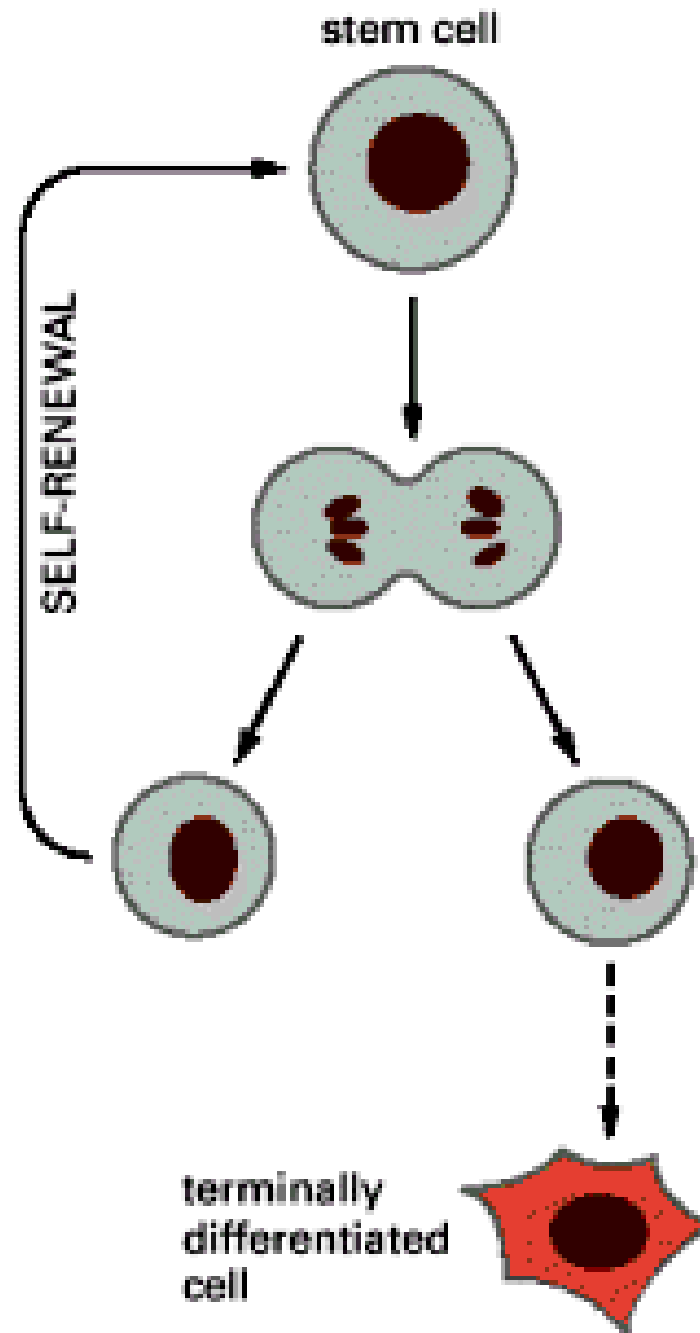


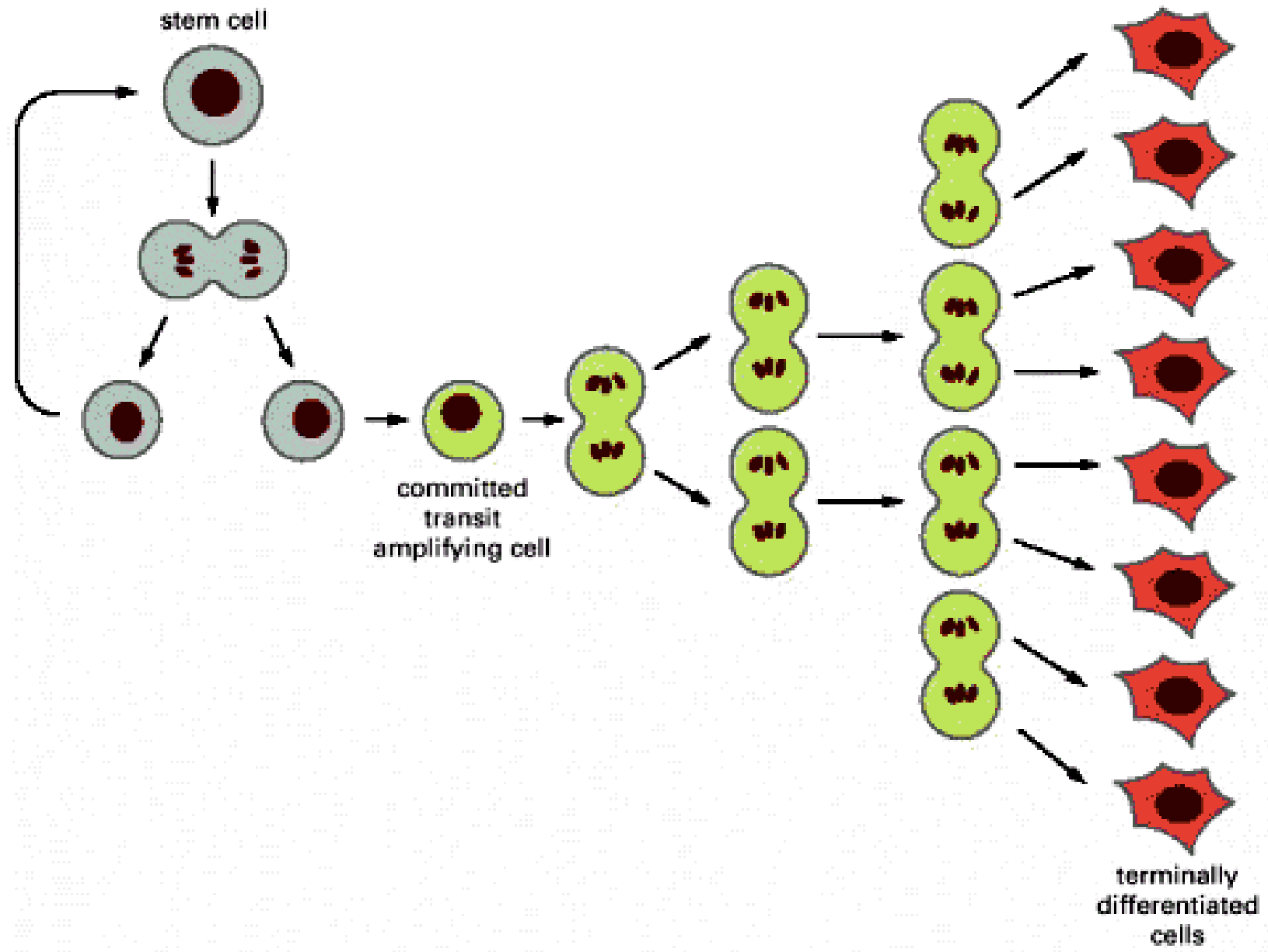
Figure 4.21. Morphological analysis of *BMP7* knockout mice. A wild-type (A) and a homozygous *BMP7*-deficient mouse (B) at day 17 of their 21-day gestation. The *BMP7*-deficient mouse lacks eyes. The kidneys of these mice at day 19 of gestation are shown in (C). The kidney of the *BMP7*-deficient mouse is severely atrophied. Microscopic sections revealed the death of the cells that would otherwise have formed the nephrons. (From [Dudley et al. 1995](#); photographs courtesy of E. Robertson.)

TEJIDOS EN RENOVACIÓN

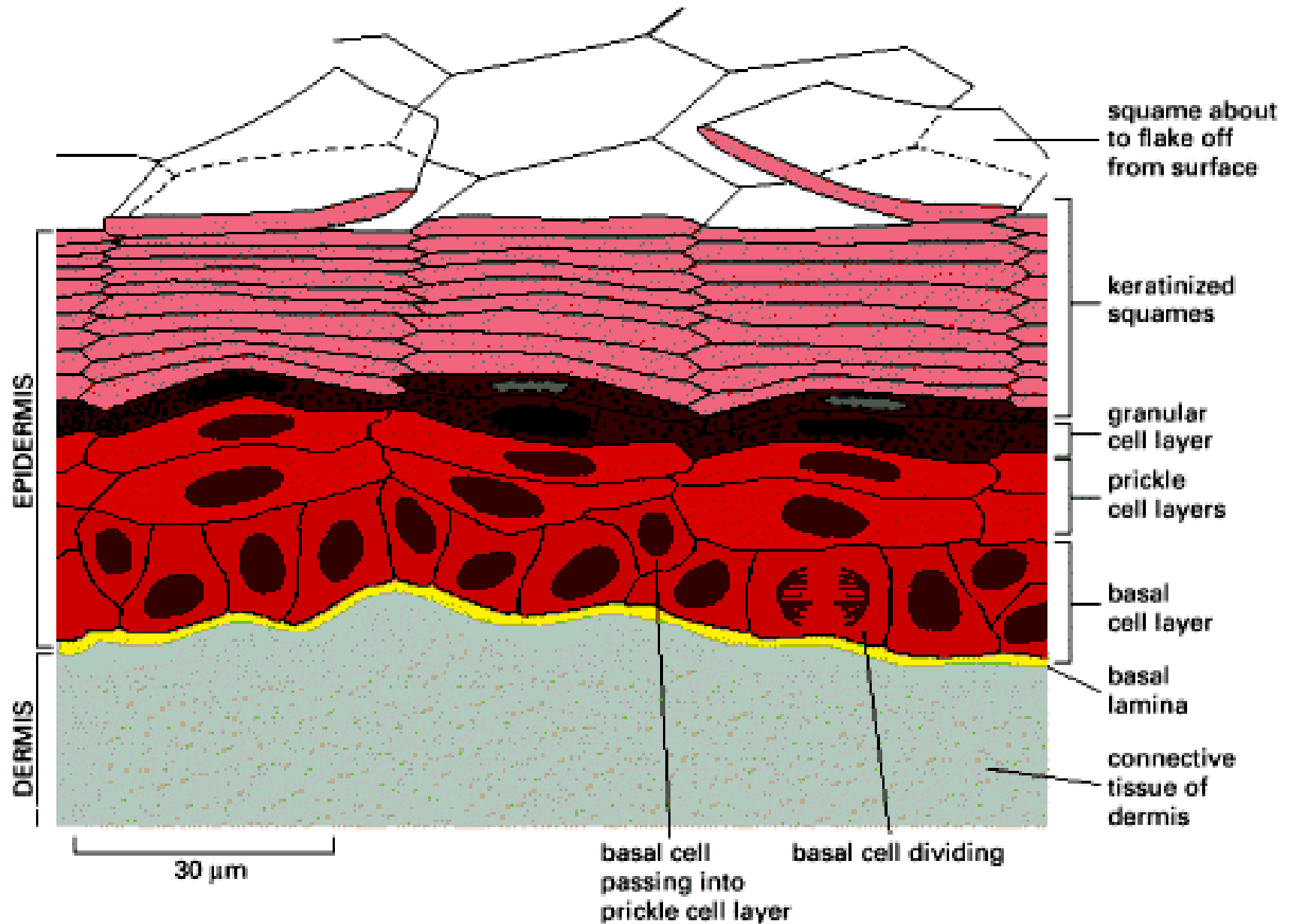
STEM CELL - CÉLULAS TRONCALES

MONOPOTENCIALES





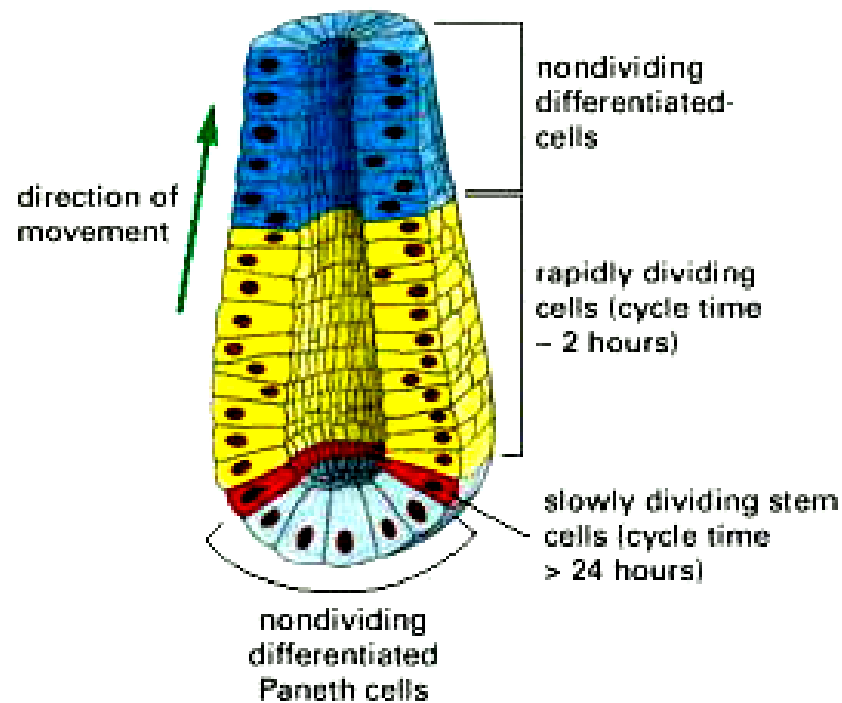
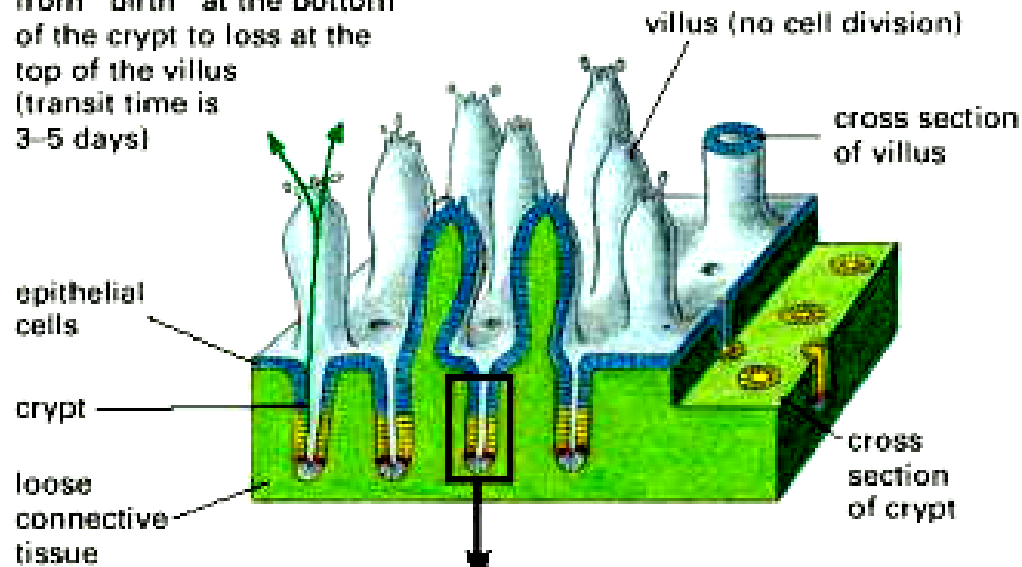
PIEL



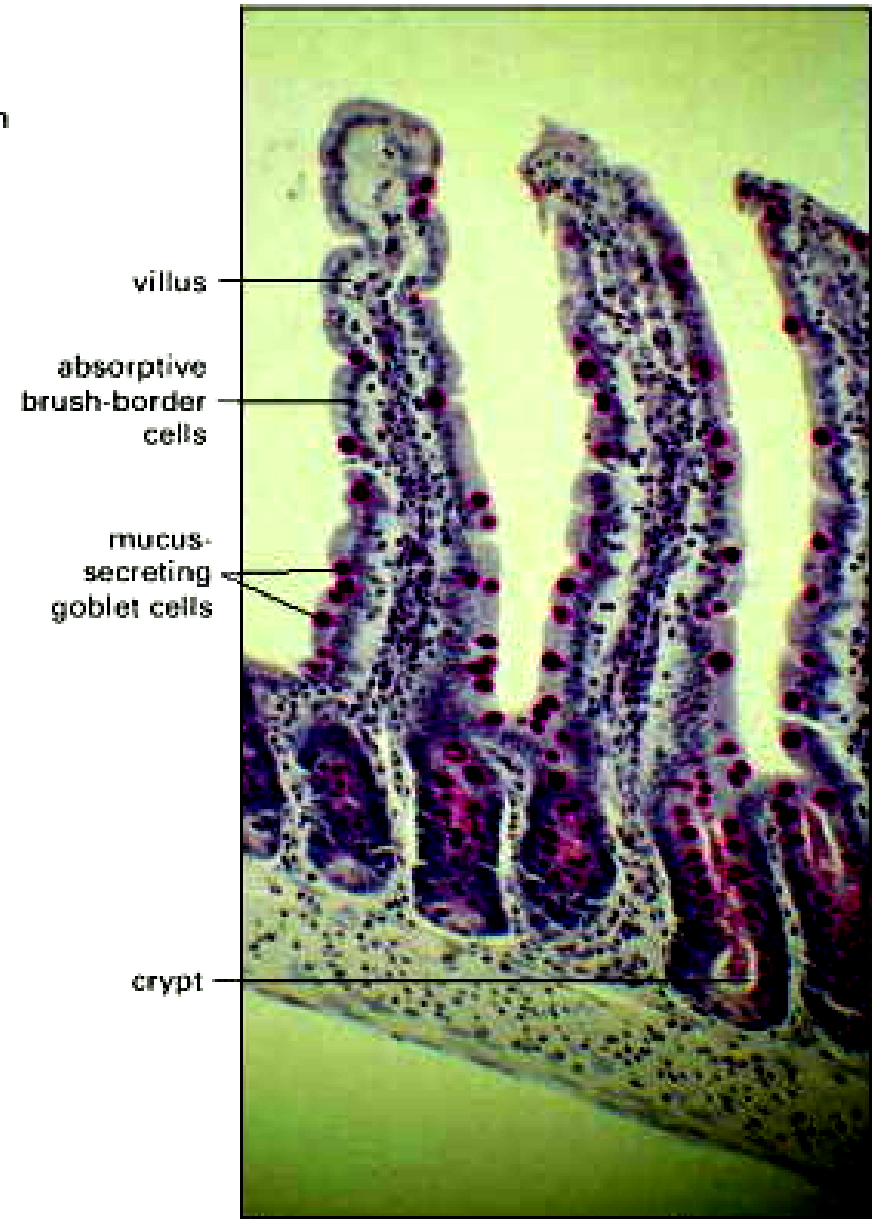
LUMEN OF GUT

EPITELIO INTESTINAL

epithelial cell migration from "birth" at the bottom of the crypt to loss at the top of the villus (transit time is 3-5 days)



(A)



(B)

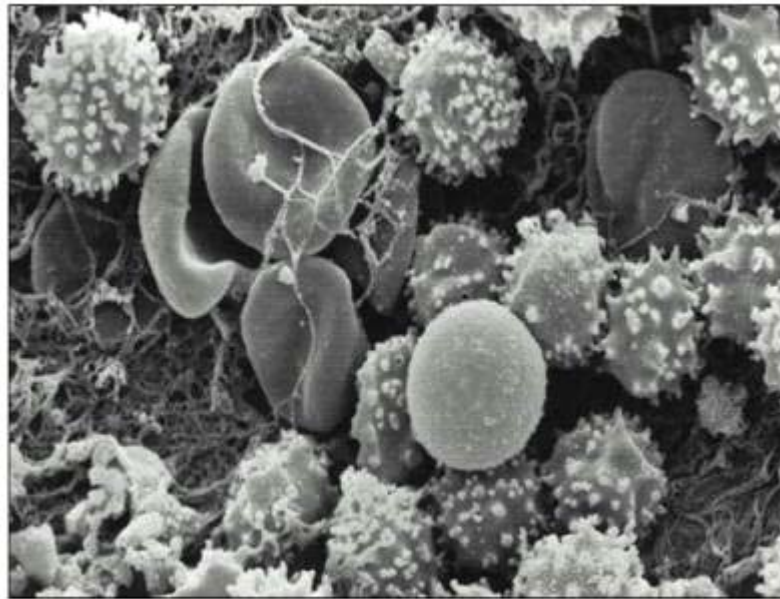
100 μm

TEJIDOS EN RENOVACIÓN

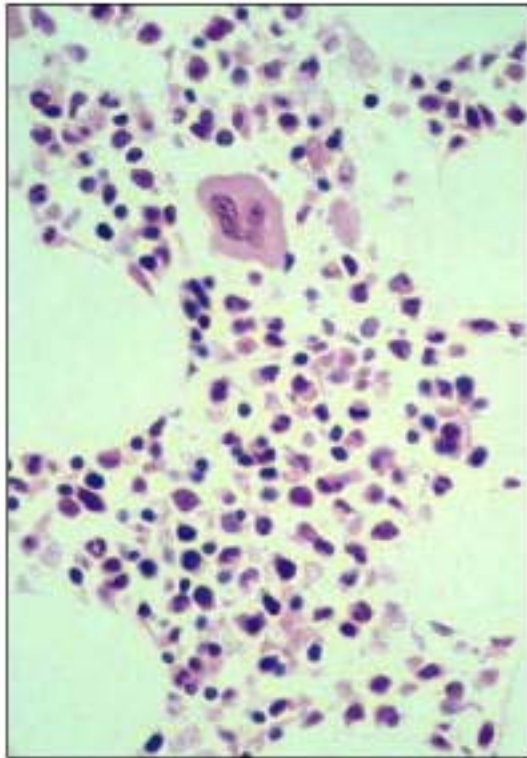
STEM CELL - CÉLULAS TRONCALES

PLURIPOTENCIALES

CÉLULAS DE LA SANGRE

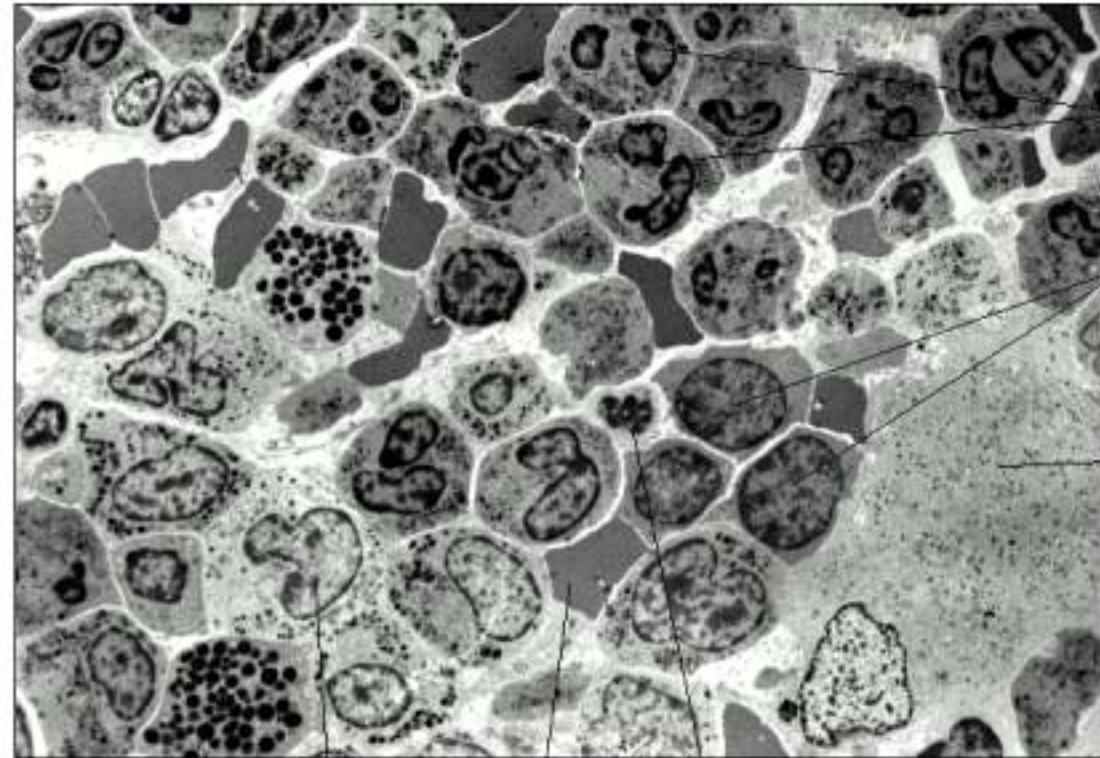


5 μm



(A)

50 μm



(B)

immature
eosinophil

immature
monocyte

erythrocyte

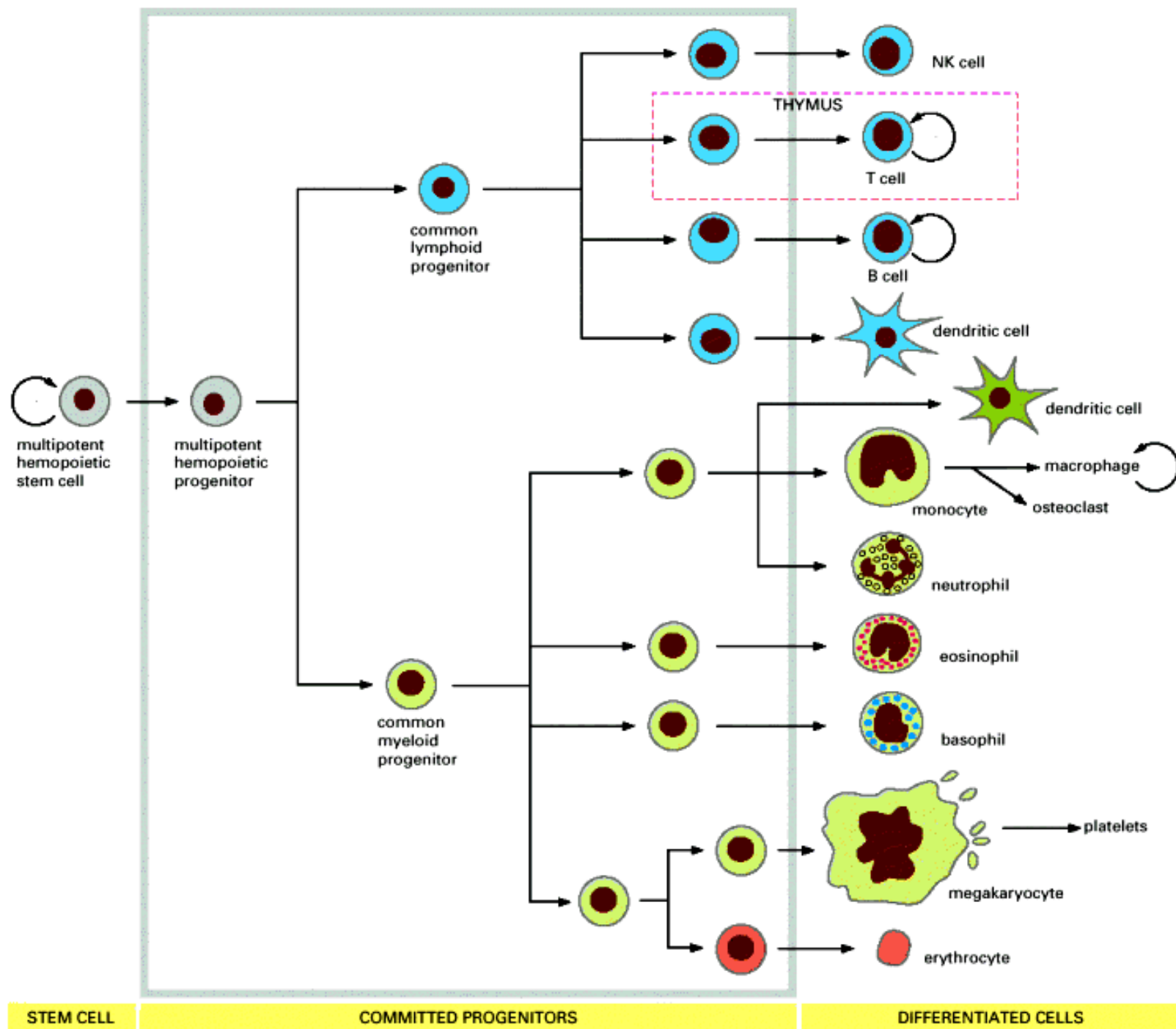
immature
lymphocyte

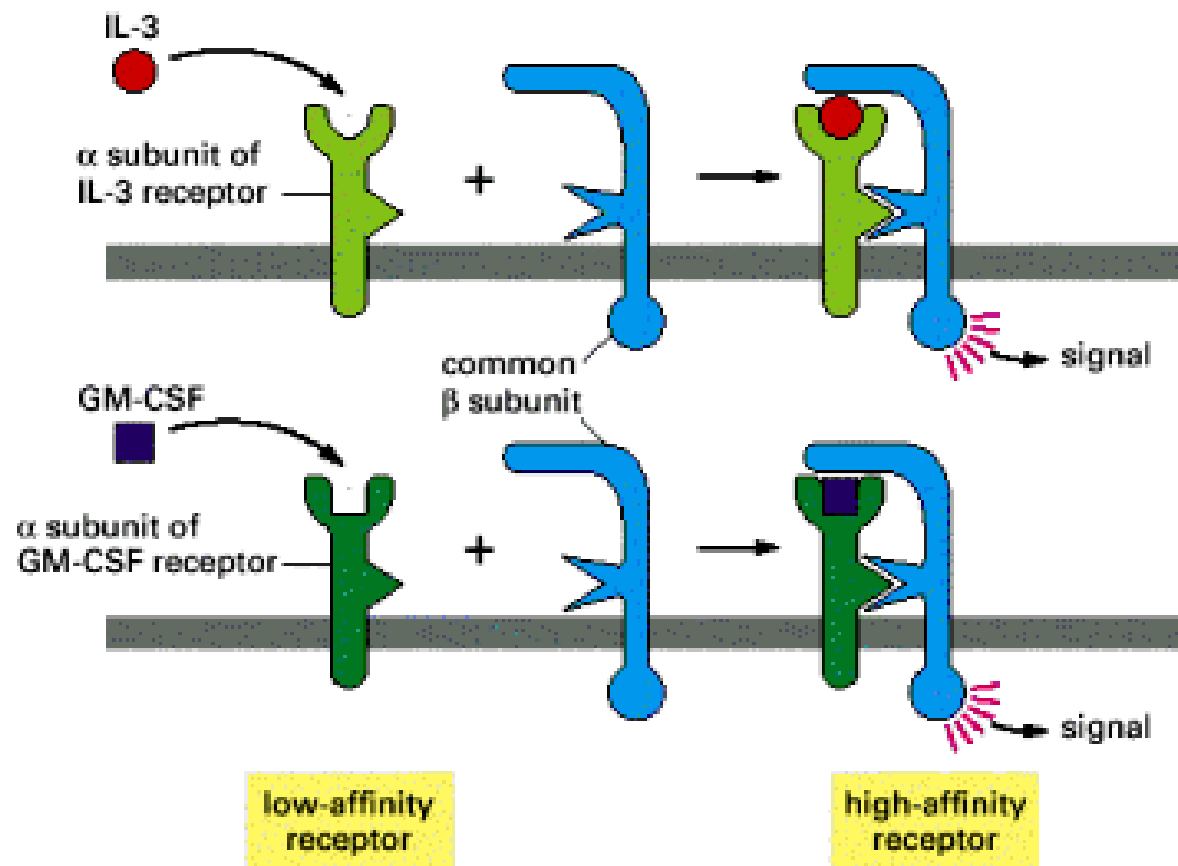
immature
neutrophils

erythrocyte
precursors

immature
megakaryocyte

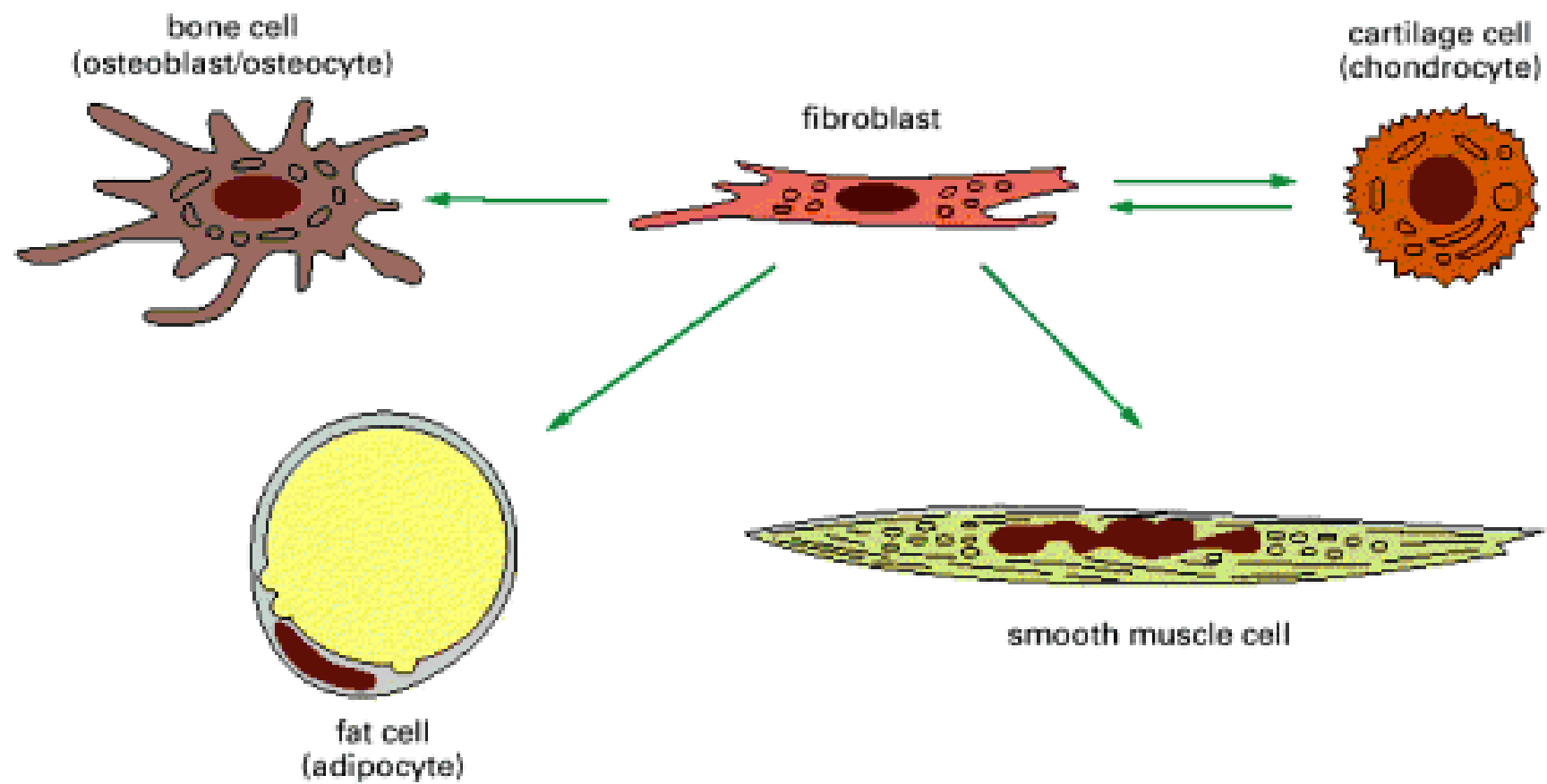
10 μm



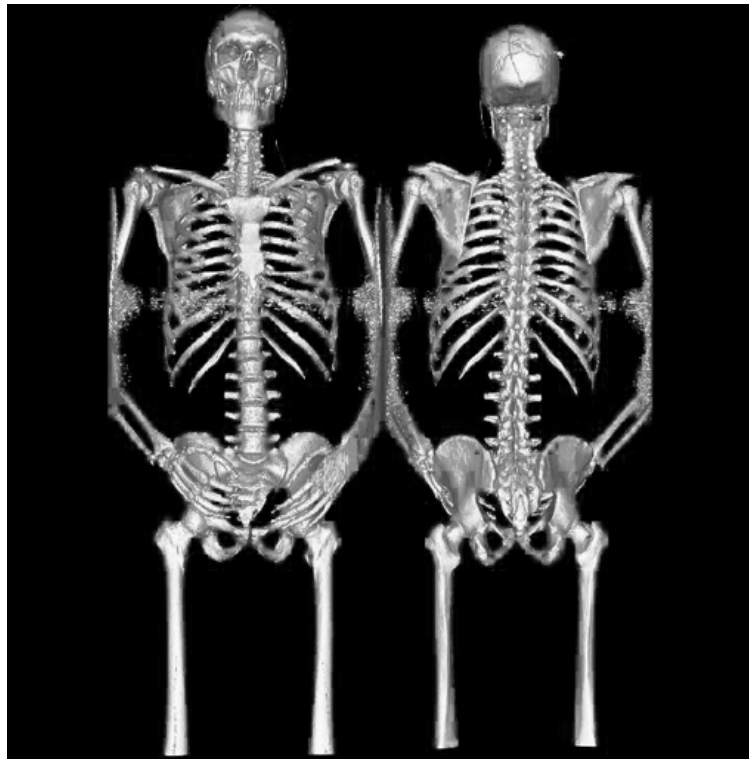


FACTOR	TARGET CELLS	PRODUCING CELLS	RECEPTORS
Erythropoietin	CFC-E	kidney cells	cytokine family
Interleukin 3 (IL-3)	multipotent stem cell, most progenitor cells, many terminally differentiated cells	T lymphocytes, epidermal cells	cytokine family
Granulocyte/ macrophage CSF (GM-CSF)	GM progenitor cells	T lymphocytes, endothelial cells, fibroblasts	cytokine family
Granulocyte CSF (G-CSF)	GM progenitor cells and neutrophils	macrophages, fibroblasts	cytokine family
Macrophage CSF (M-CSF)	GM progenitor cells and macrophages	fibroblasts, macrophages, endothelial cells	receptor tyrosine kinase family
Steel factor (stem cell factor)	hemopoietic stem cells	stromal cells in bone marrow and many other cells	receptor tyrosine kinase family

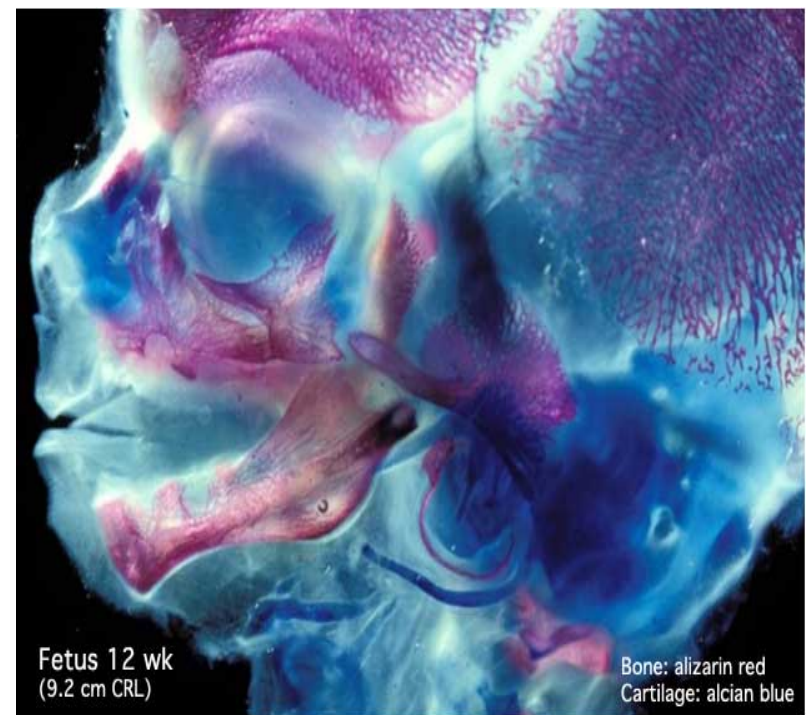
**TEJIDO CONJUNTIVO – CÉLULAS MESENQUIMÁTICAS
FIBROBLASTOS INMADUROS**



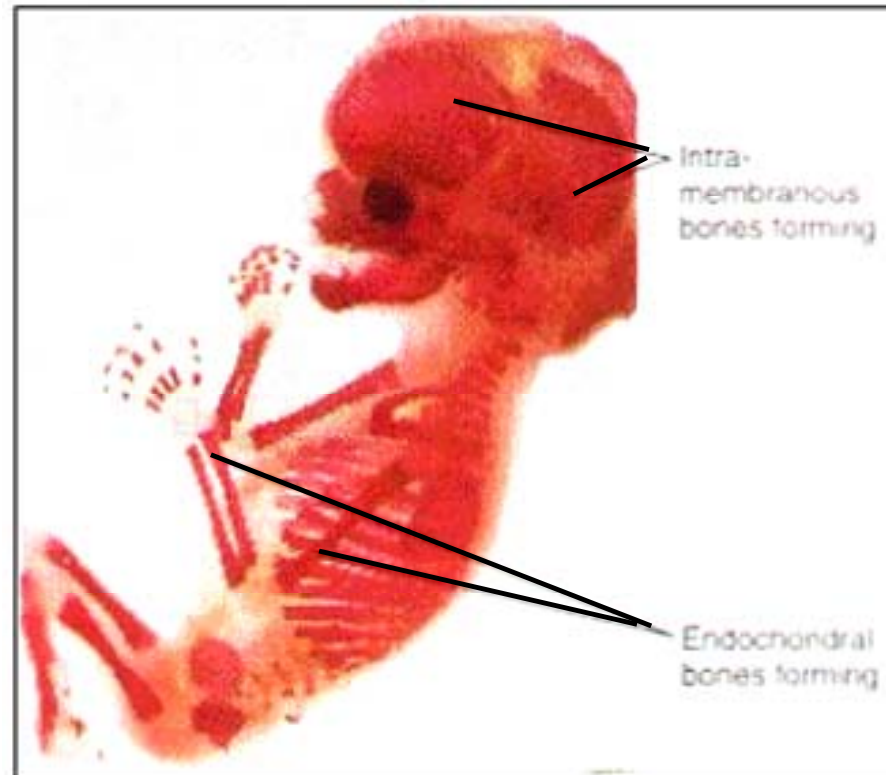
EL SISTEMA OSEO COMO MODELO PARA COMPRENDER EL PROCESO DE DIFERENCIACIÓN CELULAR

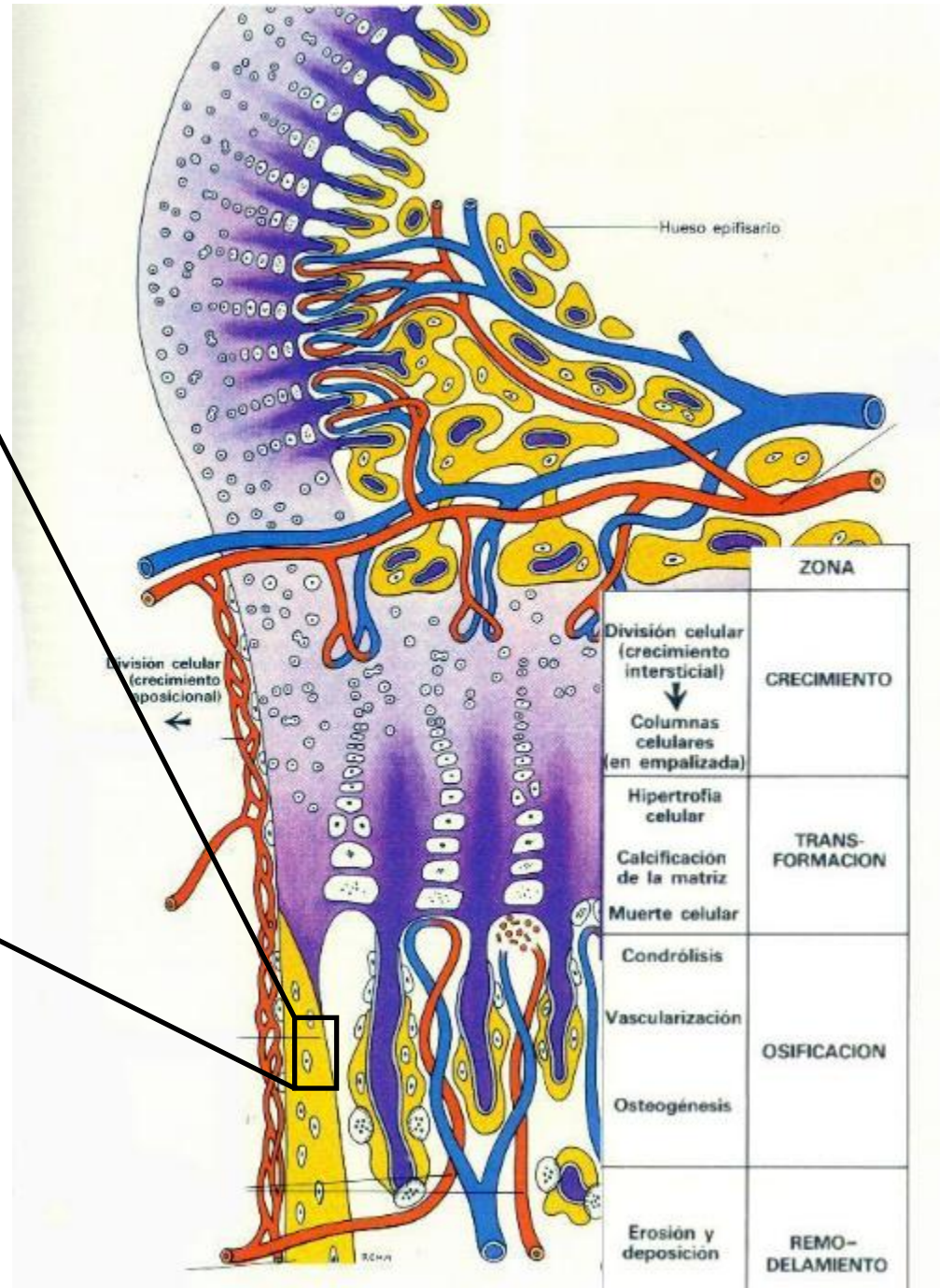
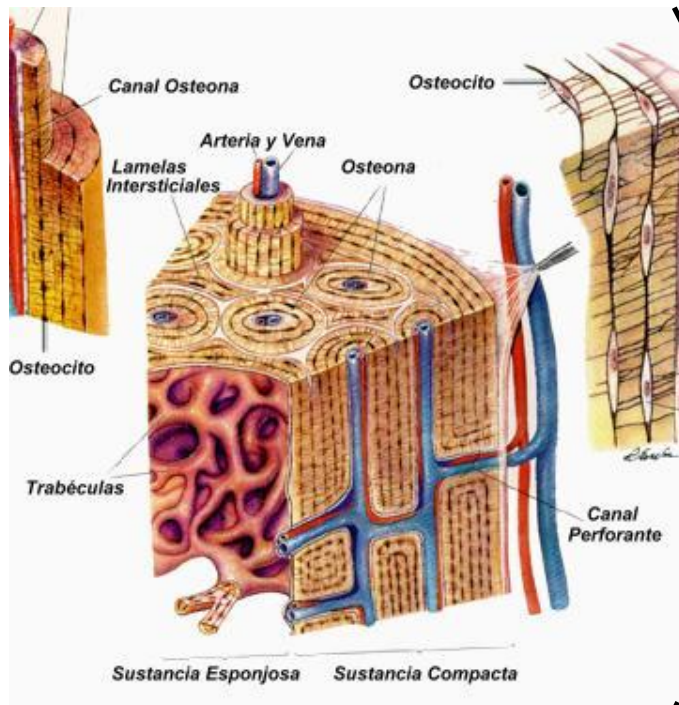


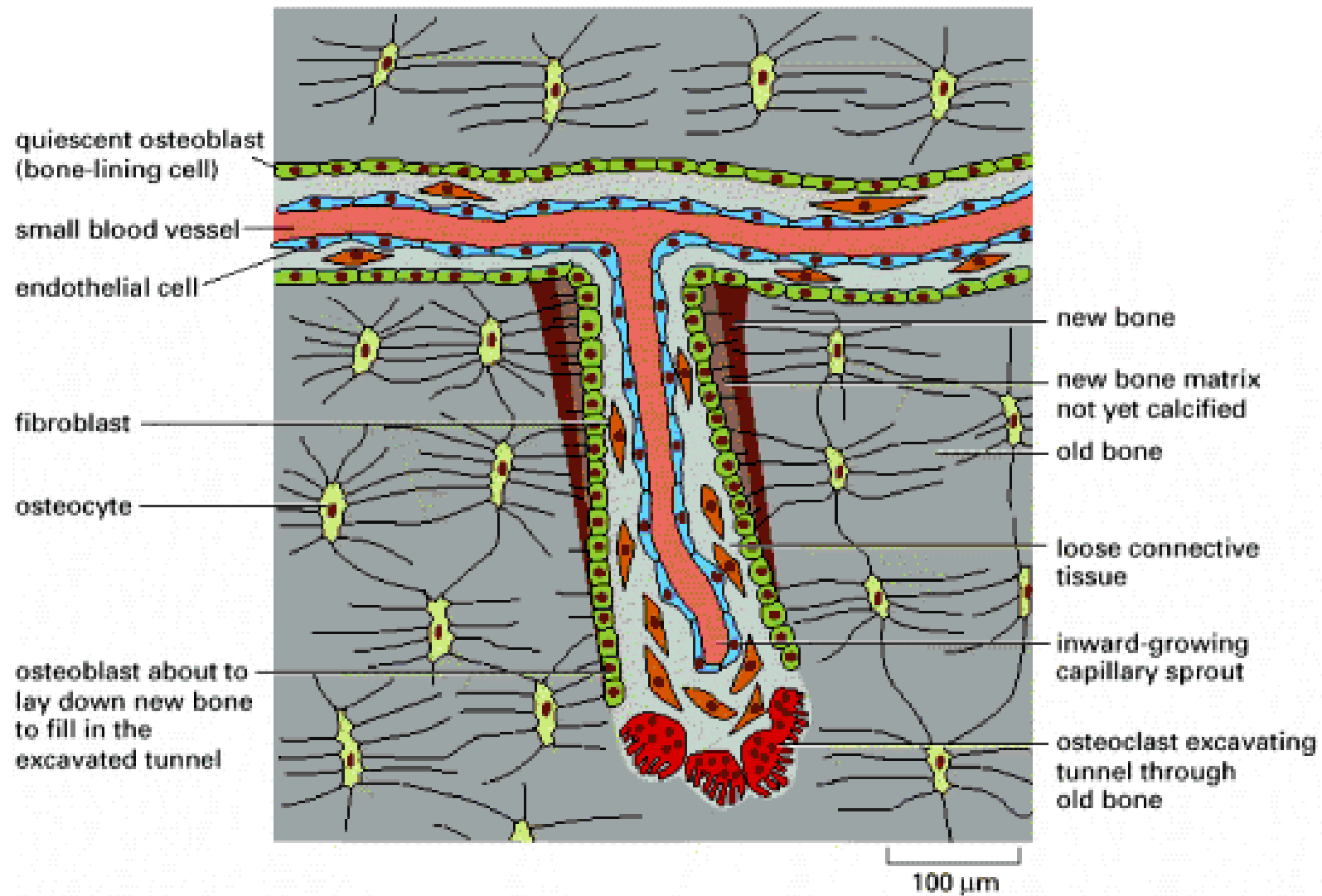
Formación Ósea



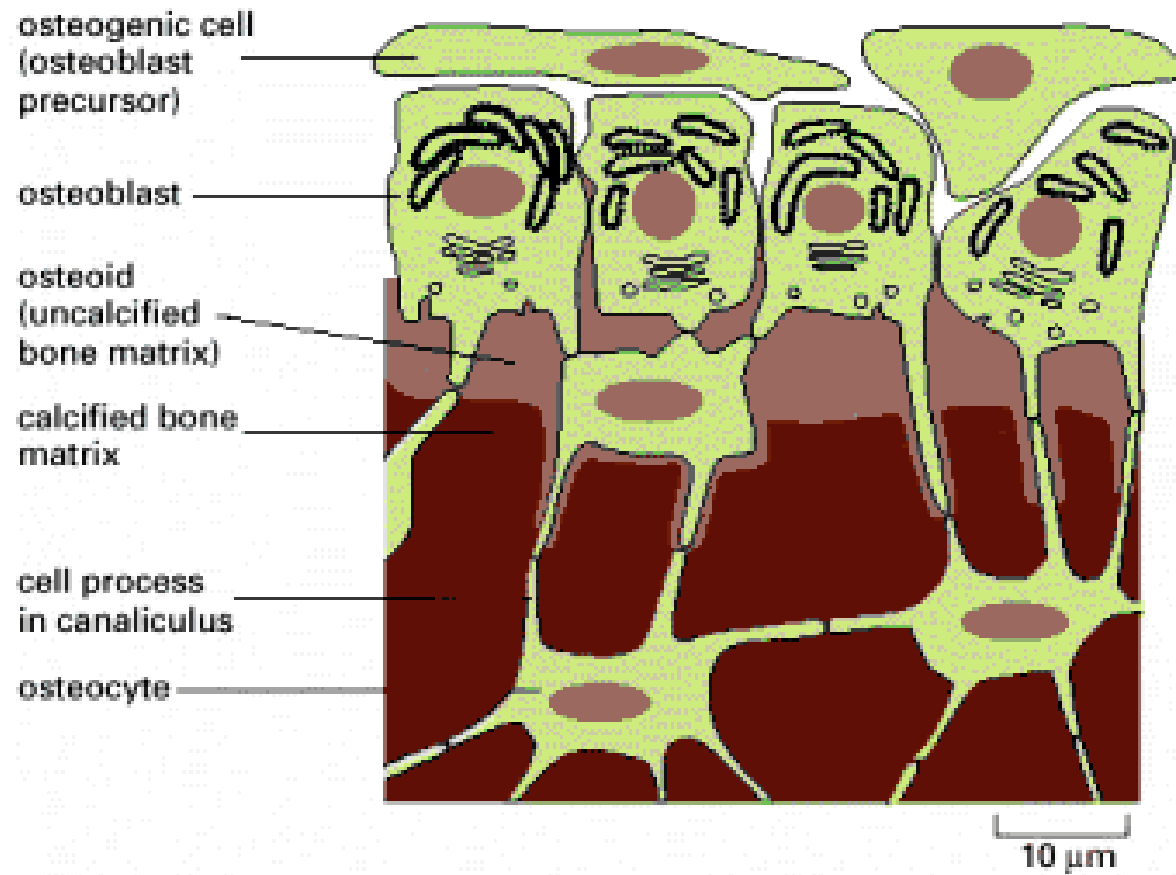
FORMACION OSEA



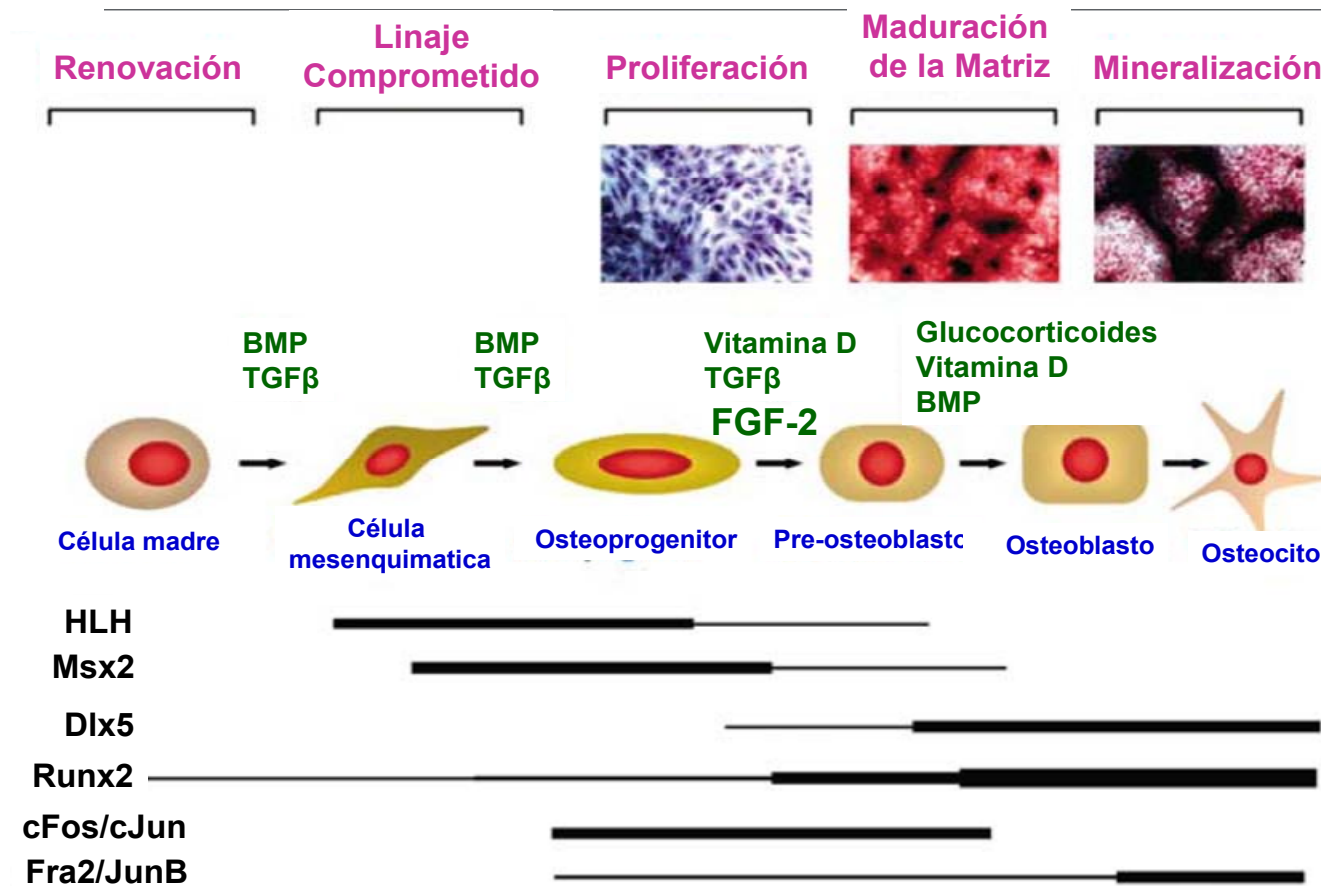


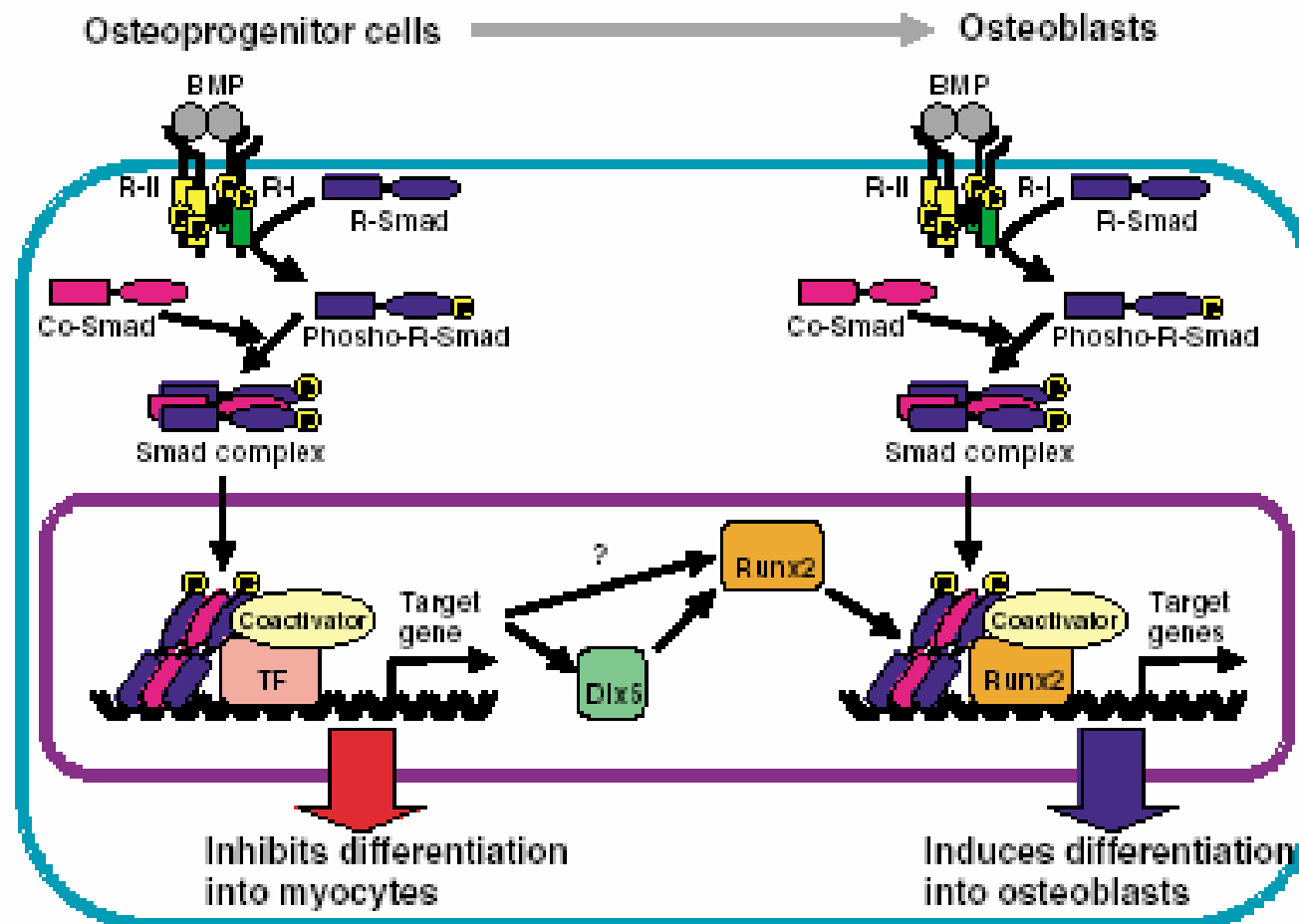


OSTEOBLASTOS



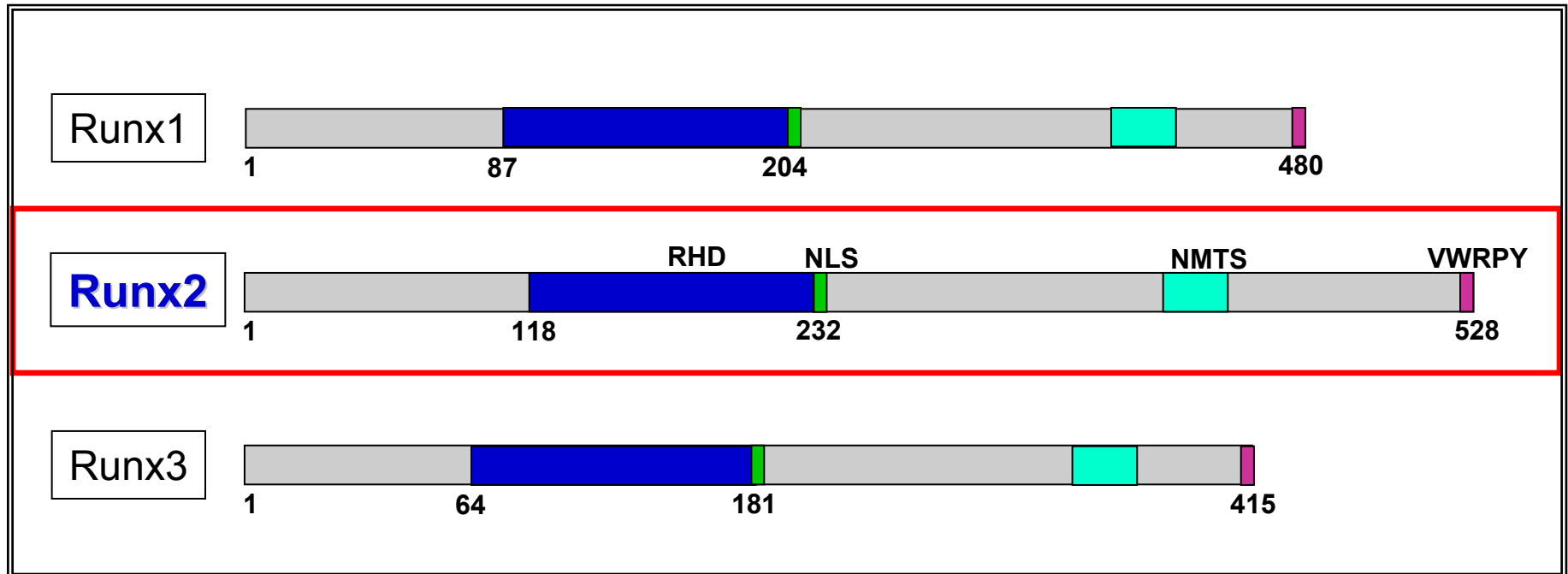
PROLIFERACION Y DIFERENCIACION DEL LINAJE CELULAR OSTEOBLASTICO





EL FACTOR DE TRANSCRIPCIÓN RUNX2

DOMINIOS FUNCIONALES DE LAS PROTEINAS RUNX



RHD

Runt Homology Domain (DNA binding, Protein-Protein interaction)

NSL

Nuclear Signaling Localization

NMTS

Nuclear Matriz Targeting Signal (PST-Rich Domain)

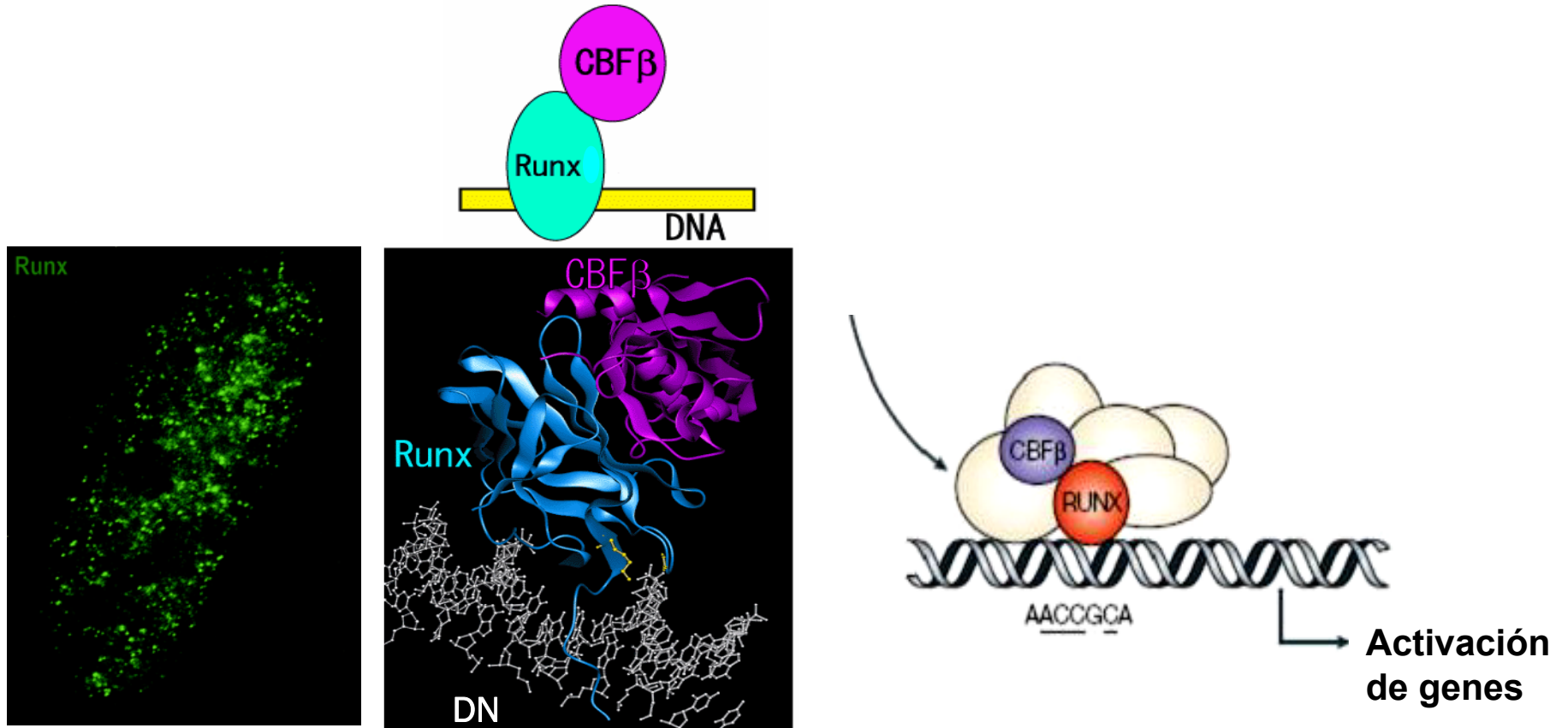
VWRPY

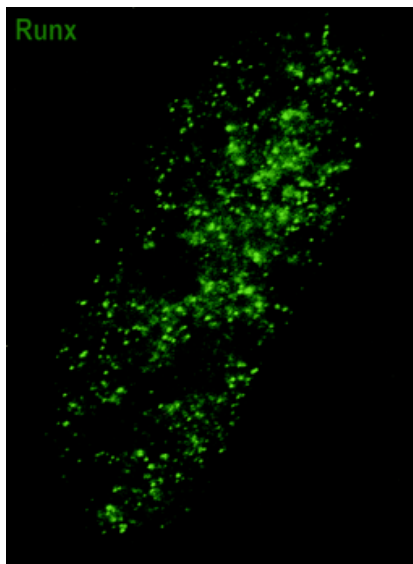
Secuencia conservada de aminoácidos (Groucho/TLE)

RUNX2

FACTOR DE TRANSCRIPCION

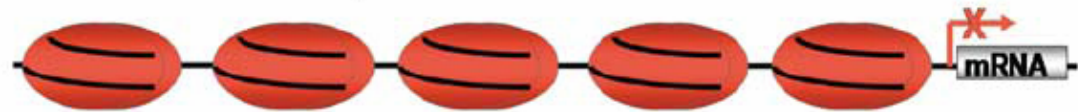
RUNX2 SE LOCALIZA EN EL NUCLEO DE LA CÉLULA





a

Inactive Osteocalcin Gene



b

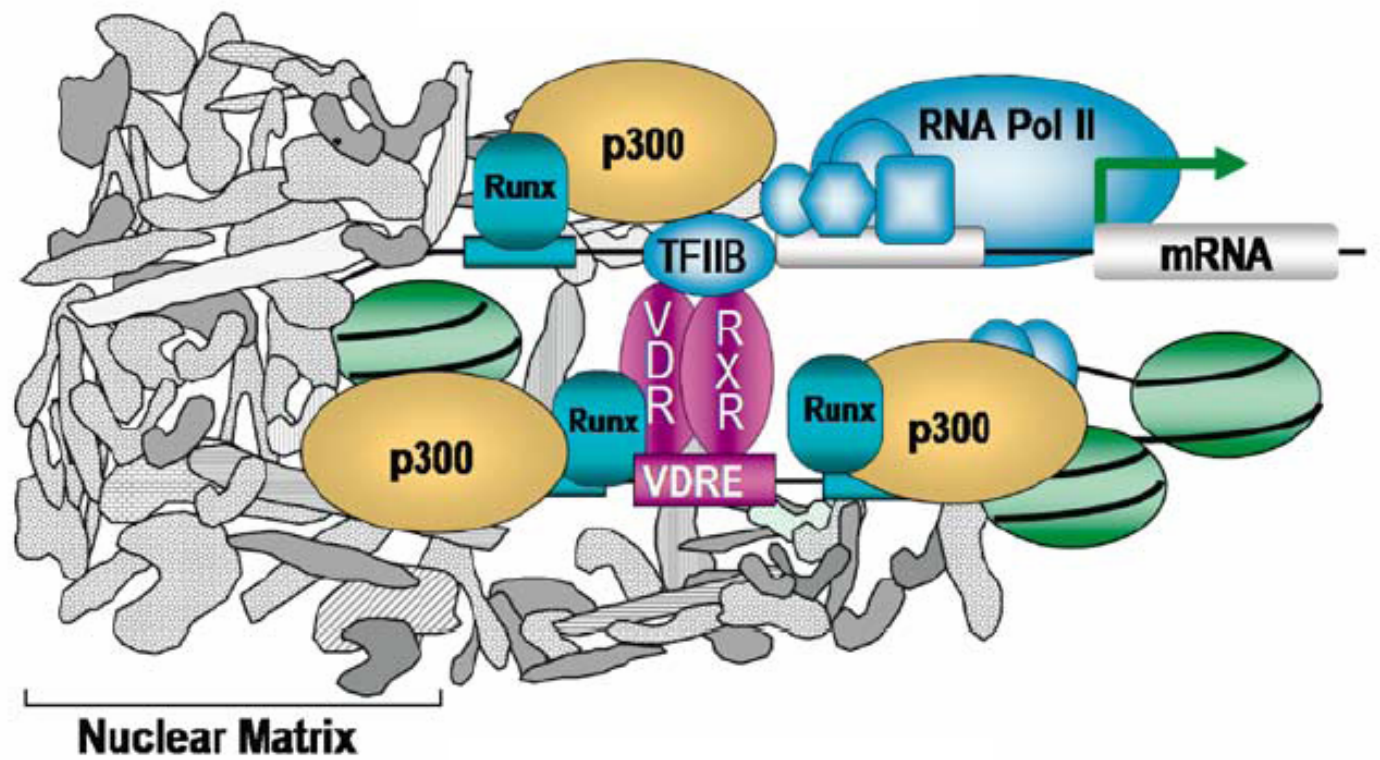
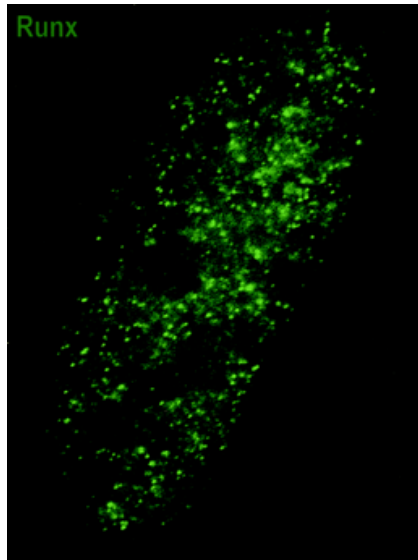
Basal Transcription of OC Gene

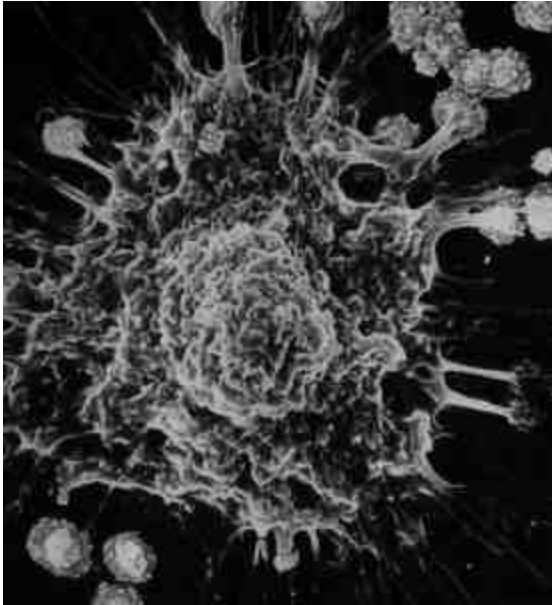


c

Vitamin D Induced Transcription of OC Gene

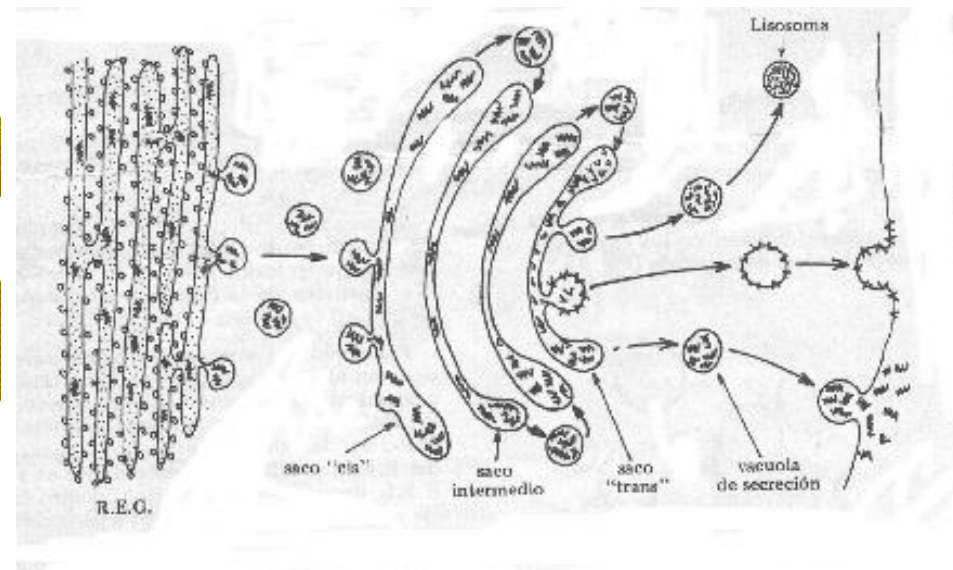
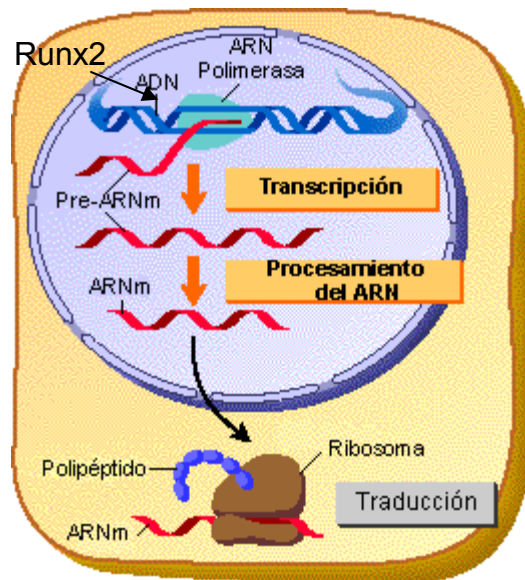




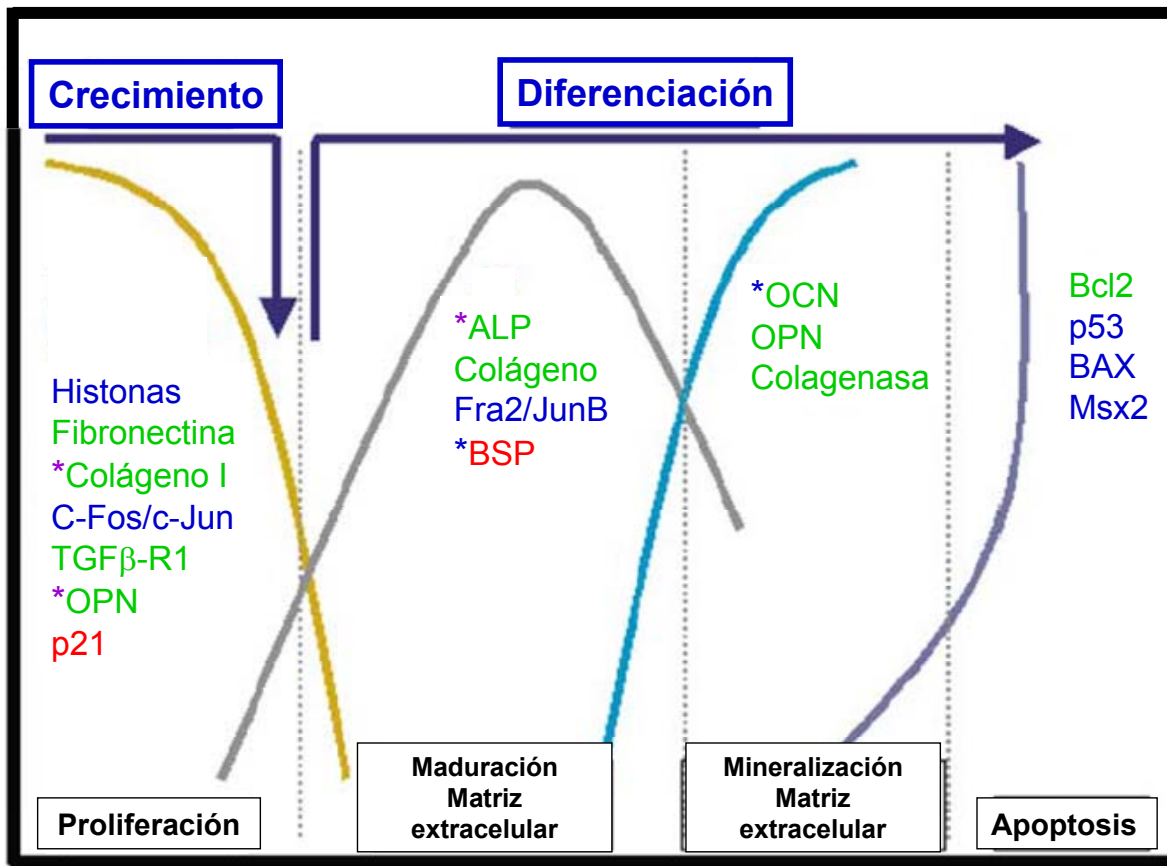


Osteoblasto

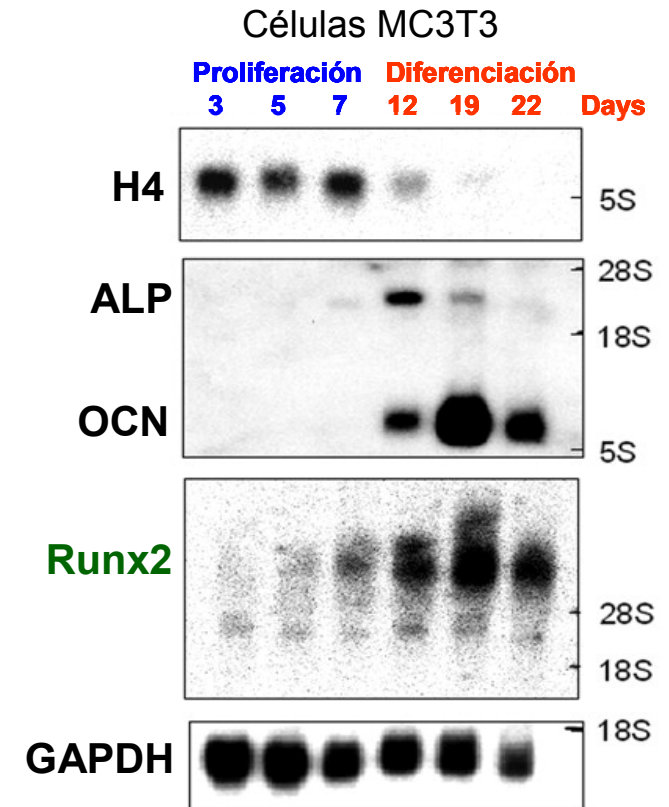
Osteoblasto es una célula especializada en la secreción de proteína que estructuran una matriz extracelular mineralizada.



PROGRAMA DE EXPRESION GENICA ASOCIADA A PROLIFERACION Y DIFERENCIACION OSTEOLASTICA



(En verde genes regulados positivamente por Runx2)



Niveles relativos de proteína Runx2

GENES REGULADOS POR RUNX2

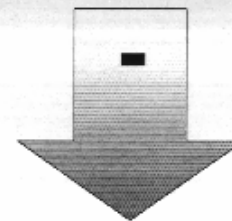
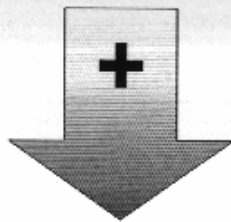
<u>Bone Related</u>	
- Runx2.....	[Drissi et al., 2000b]
- Collagen.....	[Enomoto et al., 2000], [Kern et al., 2001]
- TGF β Receptor.....	[Ji et al., 1998]
- Estrogen Receptor α	[McCarthy et al., 2003; Sasaki-Iwaoka et al., 1999; Tou et al., 2001]
- Vitamin D receptor.....	[Sasaki-Iwaoka et al., 1999]
- Alkaline phosphatase.....	[Harada et al., 1999]
- Collagenase.....	[Hess et al., 2001; Jimenez et al., 1999; Selvamurugan et al., 1998; Selvamurugan et al., 2000]
- Type X collagen.....	[Zheng et al., 2003]
- Bone sialo protein.....	[Javed et al., 1999]
- Dentin sialo protein.....	[Chen et al., 2002b]
- Osteoprotegerin (RANKL).....	[Thirunavukkarasu et al., 2000]
- Osteopontin.....	[Sato et al., 1998]
- Osteocalcin.....	[Ducy et al., 1997; Merriman et al., 1995]
- Galectin-3.....	[Stock et al., 2003]
- C/EBP δ	[McCarthy et al., 2000]

SENALES EXTRACELULARES ACTIVAN LA TRANSCRIPCIÓN DEL GEN RUNX2

A

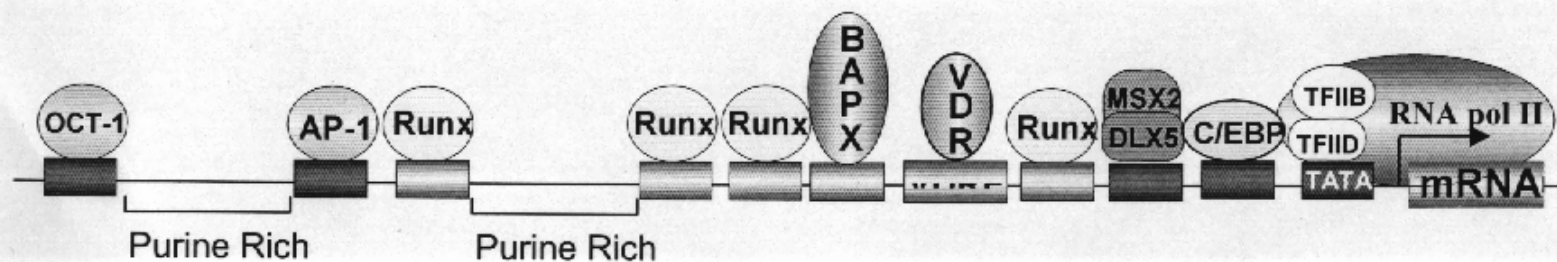
- BMP/TGF β (Lee et al., 1999;)
- MAPK (Xiao et al., 2000))
- PKA (Selvamurugan et al., 2000)
- FGF (Zhou et al., 2000, Xiao et al., 2002)
- Hedgehog (Takamoto et al., 2003)

- Glucocorticoids (Chang et al., 1998)
- Vitamin D (Ducy et al., 1997; Drissi et al., 2002)
- PPAR γ 2 (Lecka-Czernik et al., 1999)
- cAMP (Tintut et al., 1999)
- Src (Marzia et al., 2000)
- TNF α (Gilbert et al., 2002)
- BAPX1 (Lengner et al 2003)

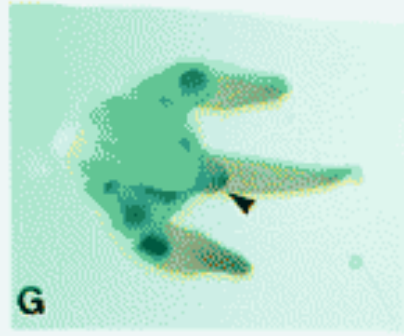
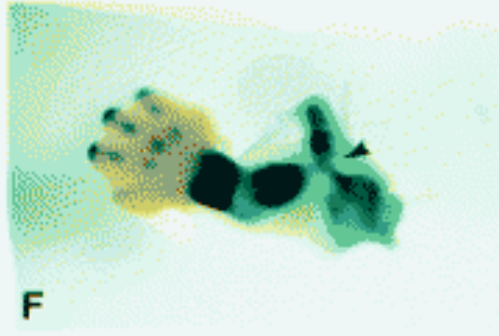
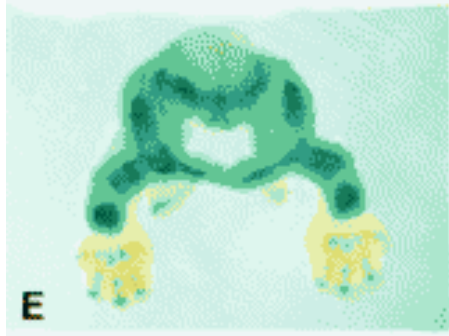
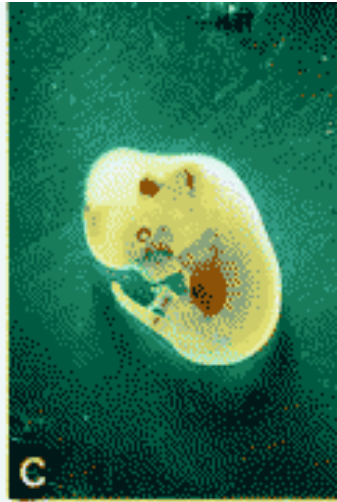
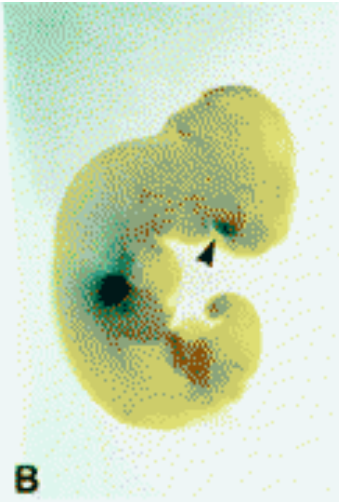


Runx2

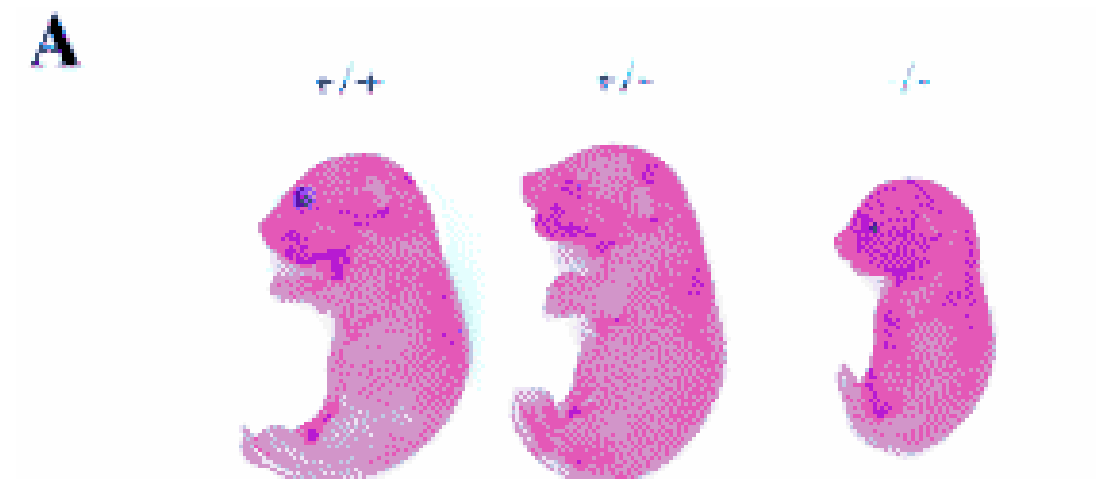
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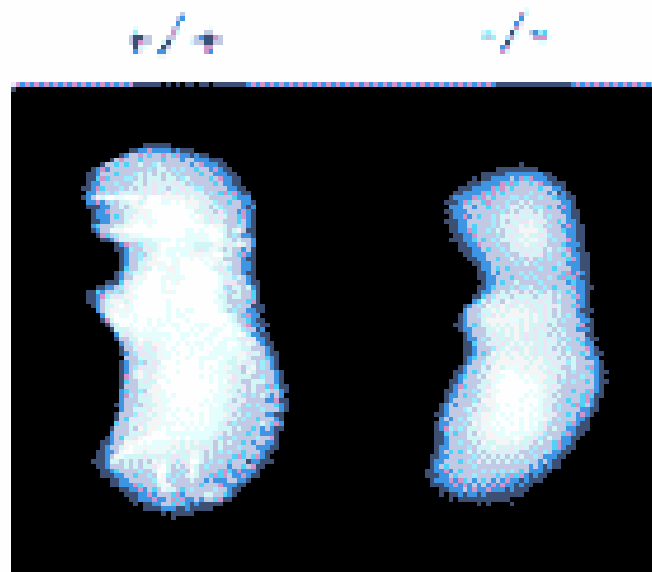
- Functionally validated sites
- Putative sites

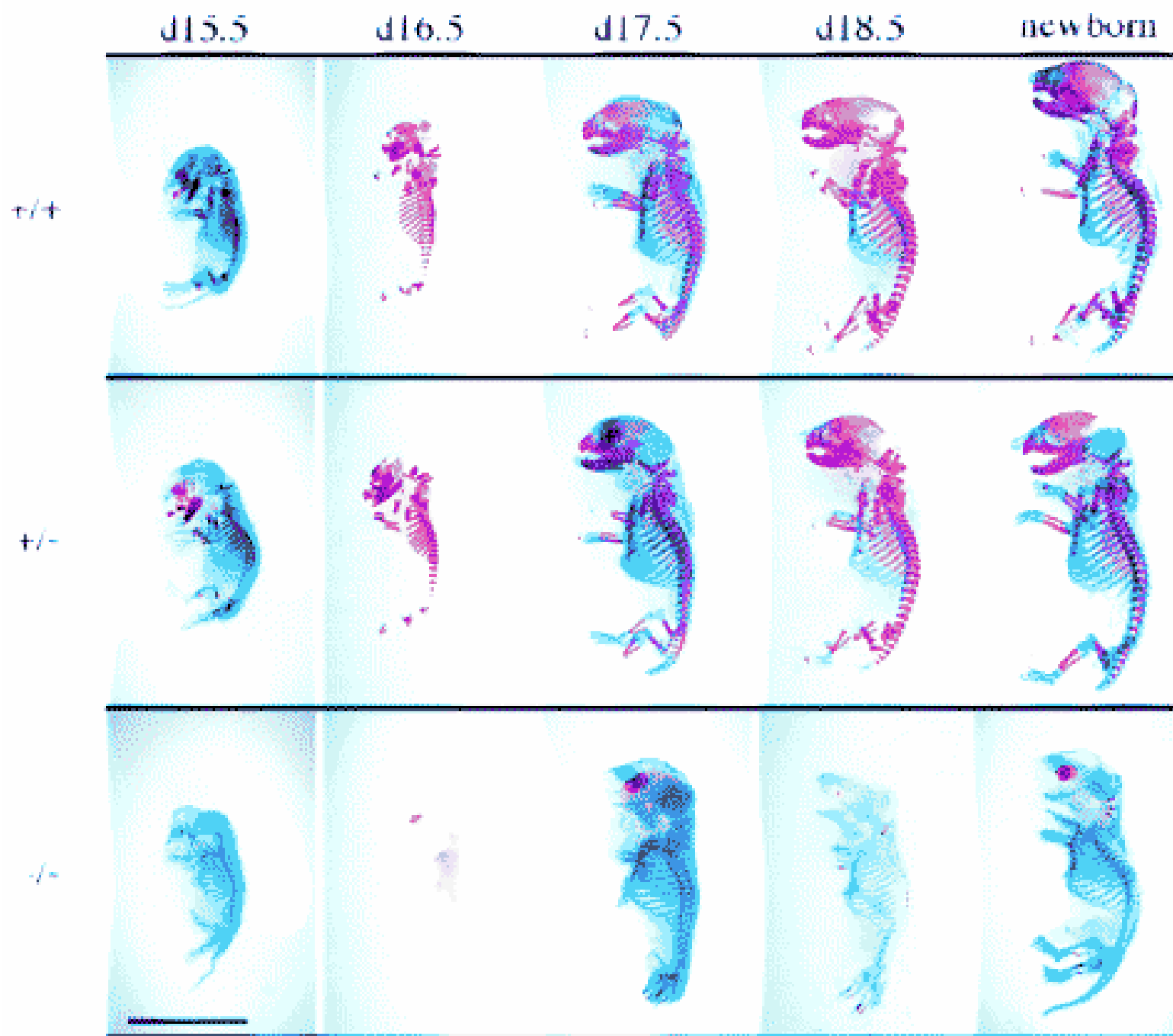


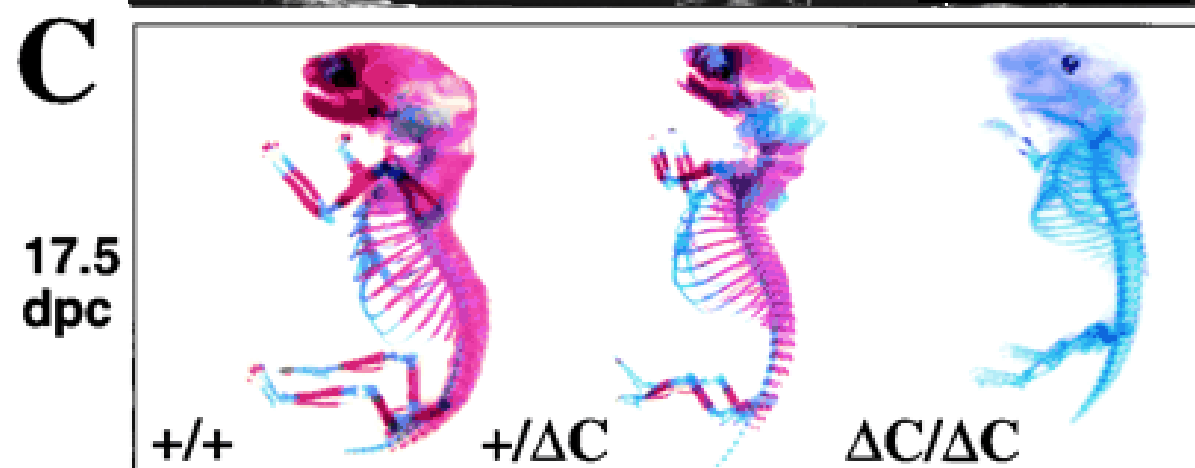
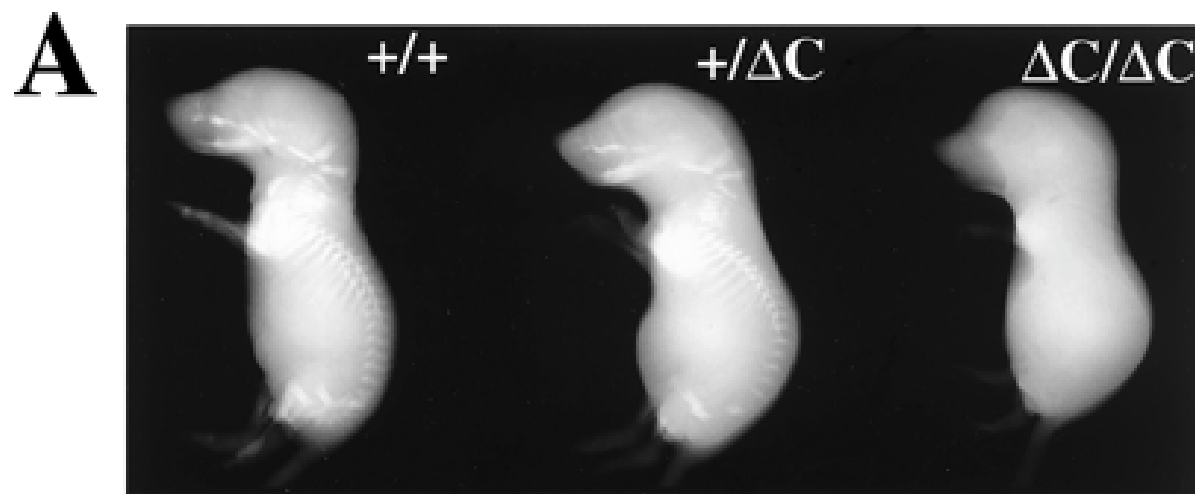
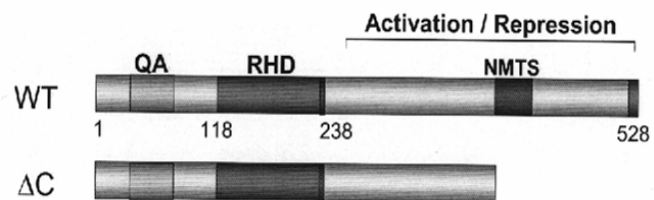
A



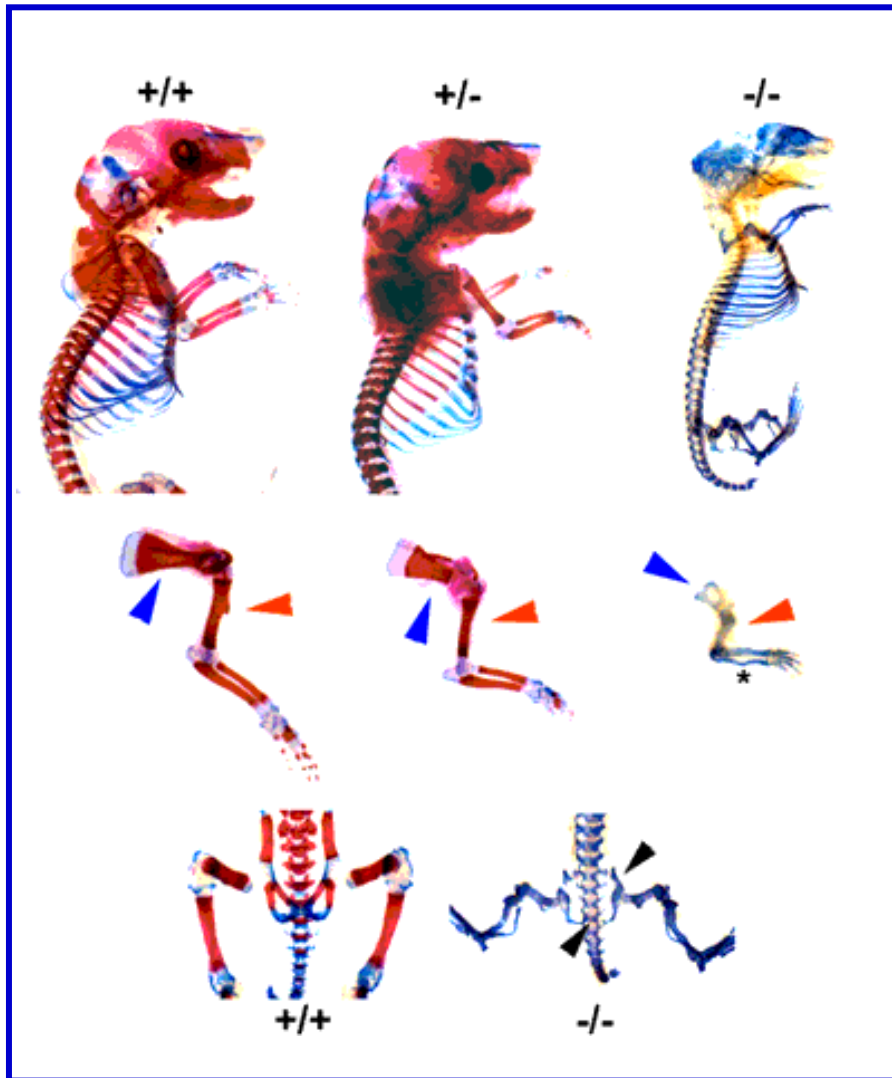
B







IMPORTANCIA DE RUNX2 EN LA DIFERENCIACION OSEA



Fenotipo esquelético en ratones

- wt (Runx2 $+/+$)
- heterocigoto (Runx2 $+/-$)
- homocigoto (Runx2 $-/-$) *knock out*

Nótese la ausencia completa de tinción roja (hueso) en el ratón Runx $-/-$.

Komori y cols., 1987.

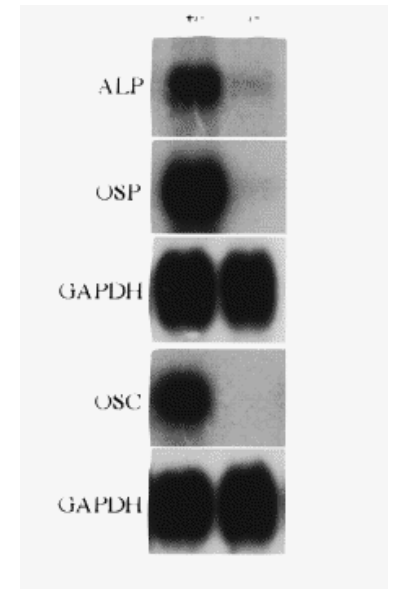
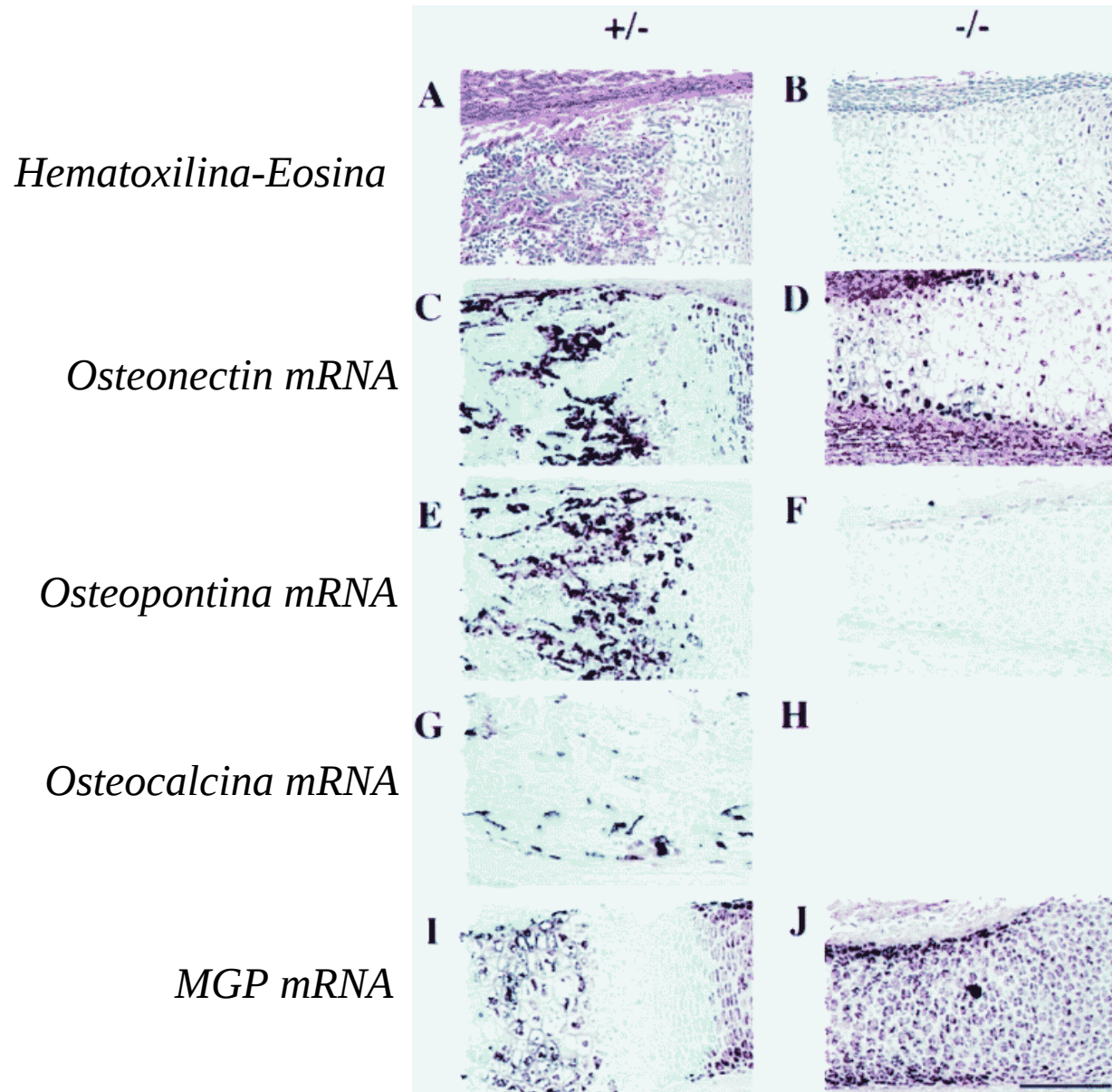


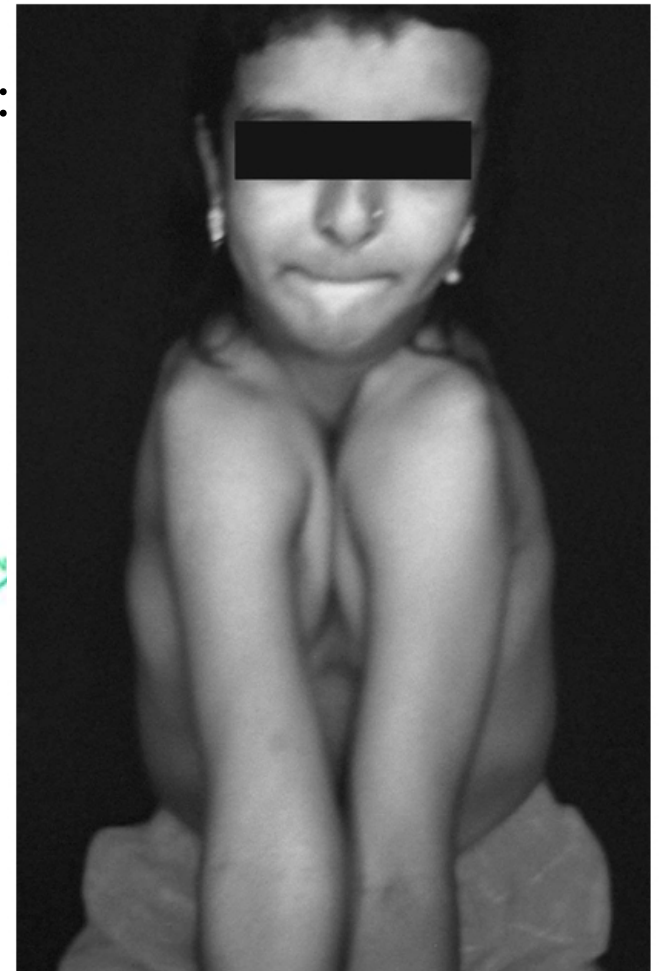
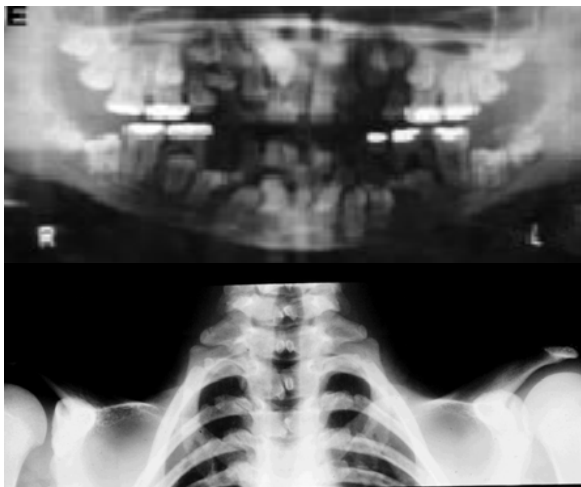
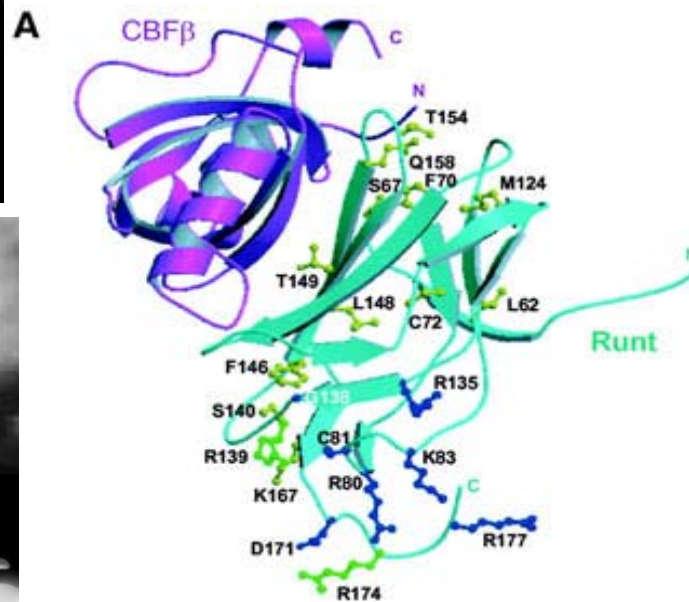
Figure 5. Distribution of *Osteonectin*, *Osteopontin*, *Osteocalcin*, and *MGP* mRNA (A, C, E, G, and I) Radius from d18.5 control (+/-) embryo. (B, D, F, H, and J) Radius from d18.5 mutant (-/-) embryo. (A and B) Staining with hematoxylin and eosin. (C and D) In situ hybridization by *Osteonectin* antisense probe. (E and F) *Osteopontin* antisense probe. (G and H) *Osteocalcin* antisense probe. (I and J) *MGP* antisense probe. Bar = 0.15 mm.

Rol de Runx2 en el proceso de diferenciación ósea



Patología humana asociada:
Displasia cleidocraneal

Cr. 6p21



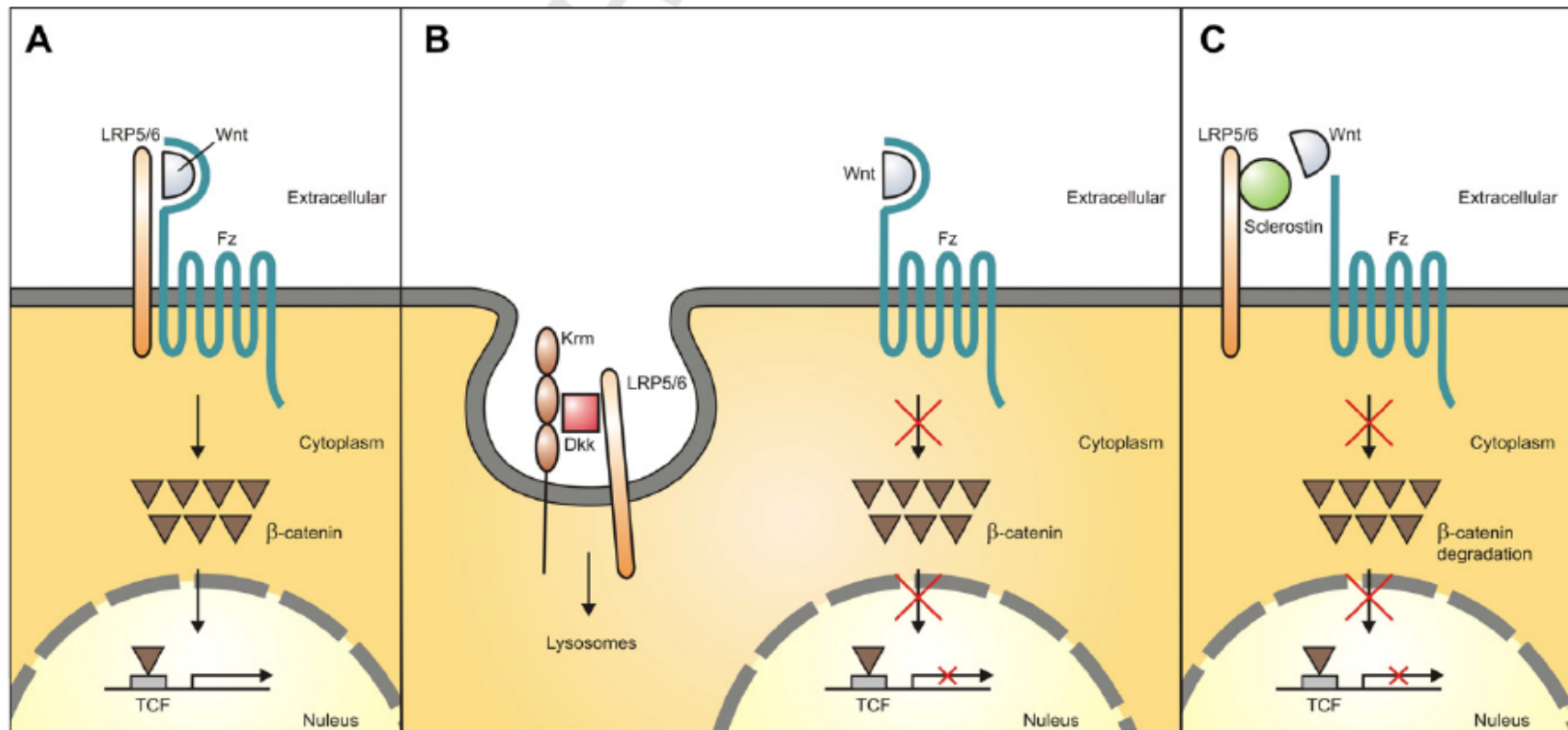


Fig. 2. The canonical wnt/β-catenin signaling pathway and its extracellular regulation. (A) Extracellular binding of wnt to the Fz–LRP5/6 receptor complex causes intracellular accumulation of β-catenin that can induce the expression of target genes after translocation to the nucleus. (B) In the presence of Dkk and Krm a tertiary protein complex can be made with LRP5/6 for internalization thus inhibiting wnt signaling as β-catenin will no longer be stabilized but will be phosphorylated and subsequently degraded. (C) The extracellular sclerostin prevents by binding to LRP5/6–Fz further signaling.

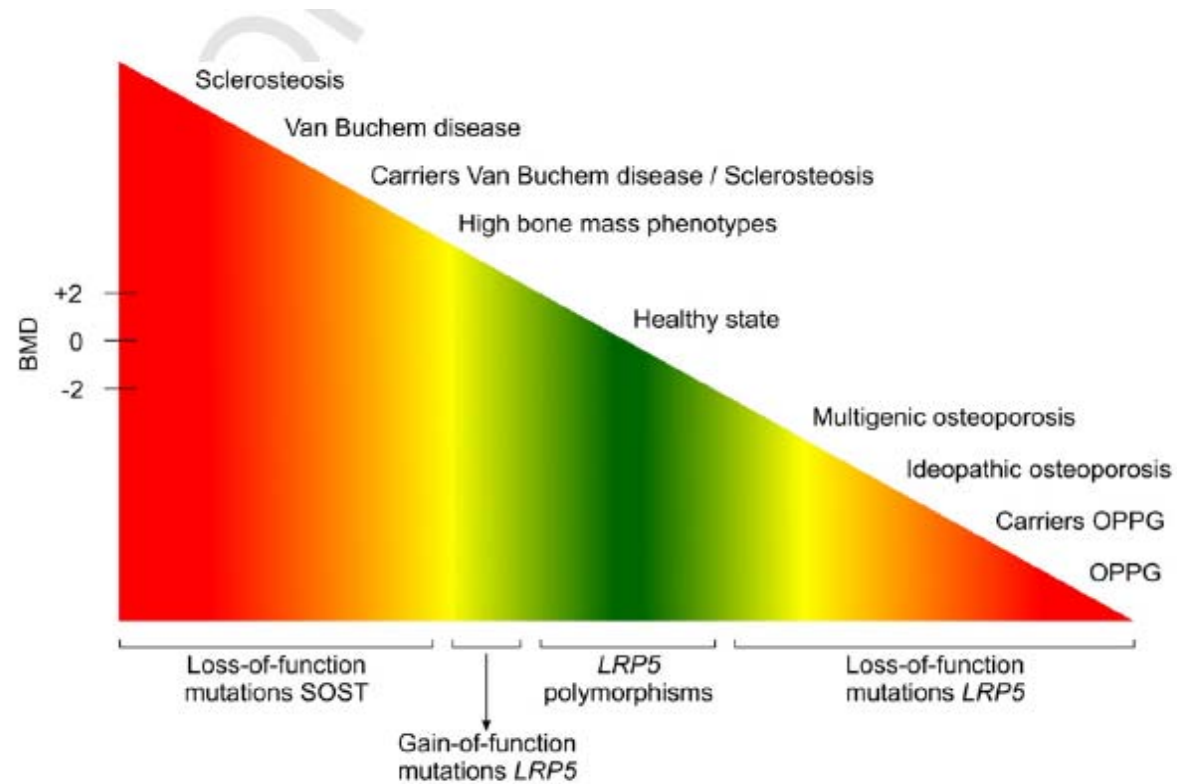


Fig. 3. Mutations or genetic variation in two components of the wnt-signaling pathway (*SOST* and *LRP5*) result in a broad spectrum of bone mineral densities.

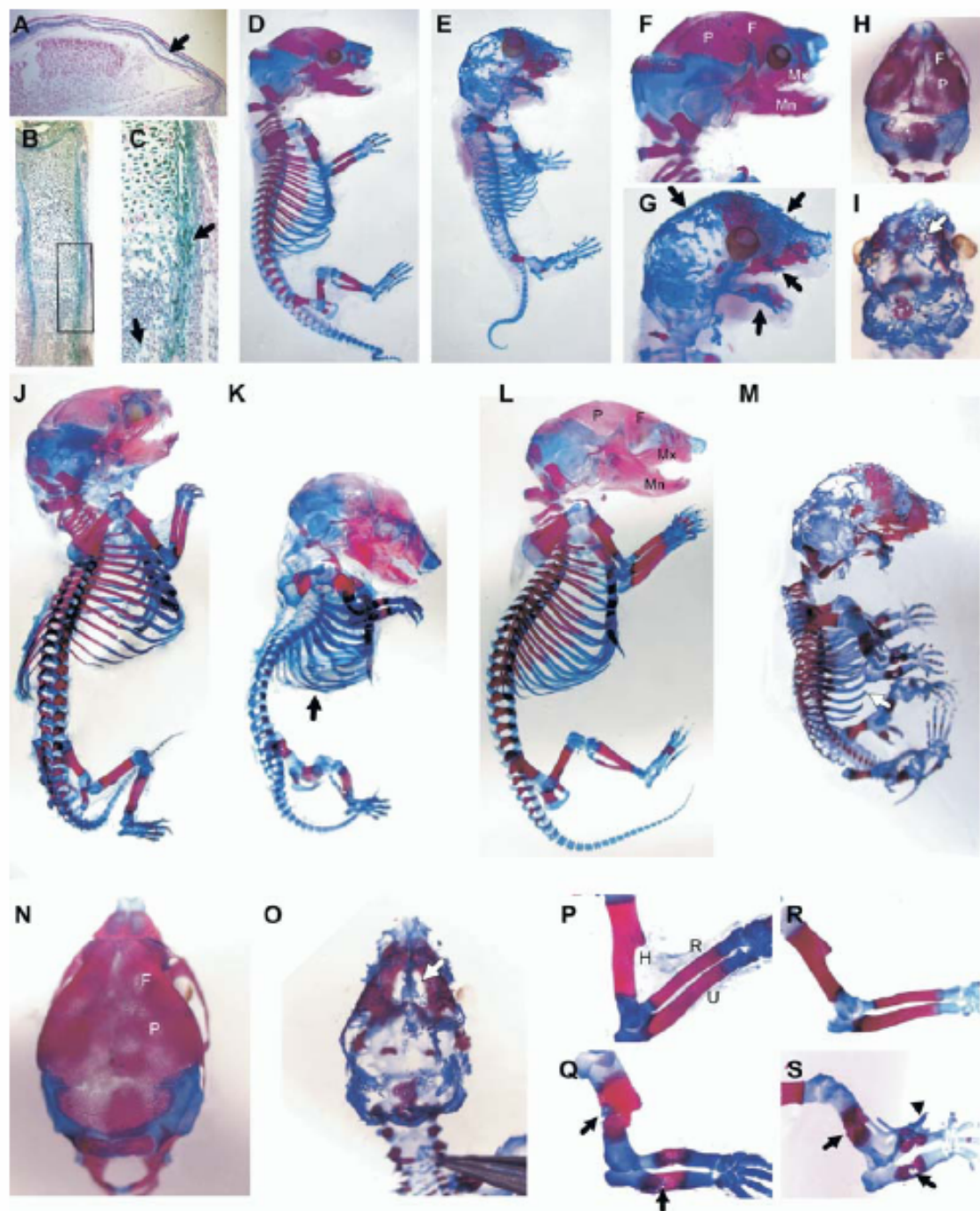


Figure 3. Inactivation of β -catenin during Both Intramembranous and Endochondral Ossification Led to Ectopic Cartilage Formation at the Expense of Bone Formation

(A) LacZ expression in the developing frontal bone (arrow) at 14.5 dpc in the *Dermo1-Cre;R26R* mouse.

(B) LacZ expression in both chondrocytes and perichondrium of a tibia in the *Col2a1-Cre;R26R* mouse embryo at 15.5 dpc.

(C) The boxed region in (B) is enlarged, showing LacZ-expressing cells (arrows) in both periosteum and primary spongiosa.

(D–S) Skeletal preparations of mouse embryos. Cartilage was stained by Alcian blue whereas bone was stained by Alizarin red. Loss of bone formation and ectopic cartilage formation are indicated by arrows. F, frontal bone; P, parietal bone; Mx, maxilla; Mn, mandible.

(D) *Catnb^{cl/+};Dermo1-Cre* embryo at 15.5 dpc.

(E) *Catnb^{cl/c};Dermo1-Cre* embryo (littermate of [D]).

(F) Enlarged head region of (D).

(G) Enlarged head region of (E).

(H) Enlarged head region of (D), dorsal view.

(I) Enlarged head region of (E), dorsal view.

(J) *Catnb^{cl/+};Col2a1-Cre* embryo at 17.5 dpc. A forelimb is shown in (P).

(K) *Catnb^{cl/+};Col2a1-Cre* embryo (littermate of [J]). A forelimb is shown in (Q).

(L) *Catnb^{cl/+};Dermo1-Cre* mouse embryo at 17.5 dpc. The dorsal view of the head is shown in (N) and a forelimb is shown in (R).

(M) *Catnb^{cl/c};Dermo1-Cre* embryo (littermate of [L]). The dorsal view of the head is shown in (O) and a forelimb is shown in (S).

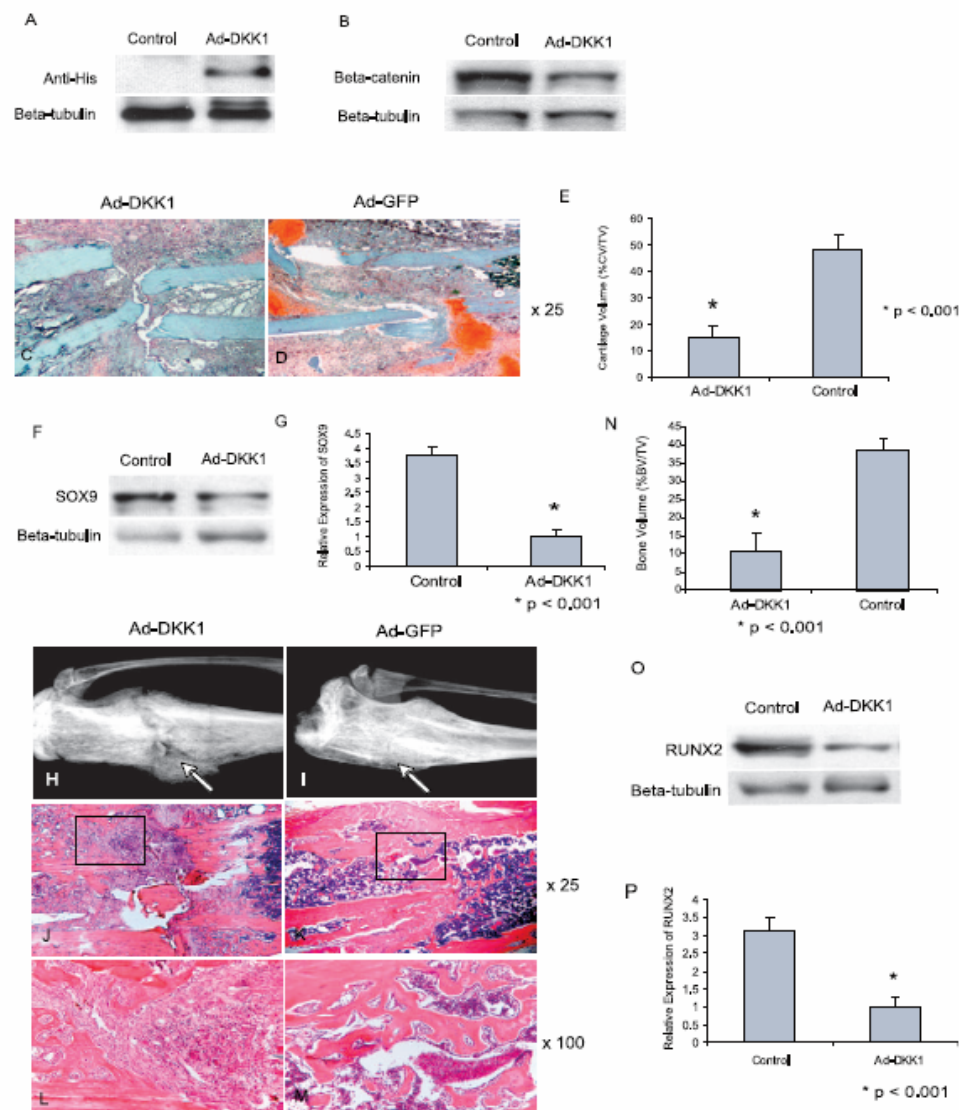


Figure 3. WNT Ligands Regulate β-Catenin during Fracture Healing

(A) Western blot analysis shows expression of DKK1 as detected by an antibody to the His tag. (B) β-catenin was substantially down-regulated in fractures treated with DKK1. (C and D) SO staining of fracture samples from DKK1-treated mice (C) and control mice (D) at 1 wk after fracture show that DKK1 treatment down-regulates chondrogenic differentiation. Images are magnified 25x. (E) Histomorphometric analysis shows a down-regulation of cartilage volume (CV) at 1 wk after fracture, as a percentage of total callus tissue volume (TV), in DKK1-treated mice. (F and G) SOX9 was also down-regulated after treatment with Ad-DKK1 at 1 wk following fracture. The expression data are consistent with our histological findings. (H and I) Radiographic analysis from DKK1-treated mice (H) and from control mice (I), at 3 wk after fracture, shows a lack of healing in the DKK1-treated animals. Arrows indicate fracture site. (J-M) HE staining from DKK1-treated mice (J and L) and from control mice (K and M) undifferentiated mesenchymal-like cells at the fracture site in DKK1-treated animals at 3 wk after fracture. Images (J and K) are magnified 25x; (L and M) show 100x magnifications of the area shown in the box in the lower-magnification images. (N) Assay of bone volume (BV) as a percentage of total callus tissue volume (TV) shows a significant decrease in bone regeneration after DKK1 treatment. (O and P) RUNX2 was also significantly down-regulated in DKK1-treated mice, suggesting an inhibition of osteoblastic differentiation. doi:10.1371/journal.pmed.0040249.g003

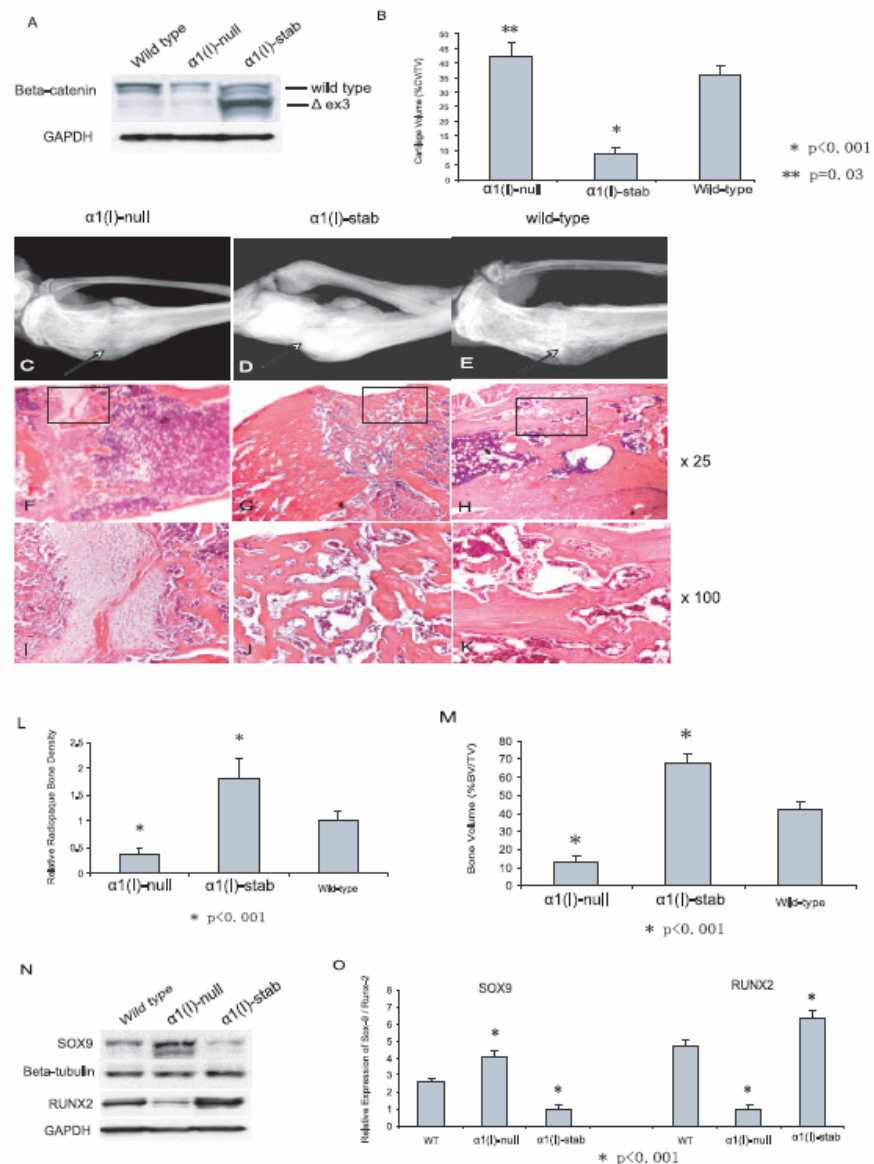


Figure 4. B-catenin Acts to Regulate Bone Formation in Mice Expressing Osteoblast-Specific β -Catenin Null or Stabilized Alleles

(A) Western blot analysis shows that β -catenin was regulated in the bone as expected in $\alpha 1(I)^{-Catnb^{null}}$ and $\alpha 1(I)^{-Catnb^{stab}}$ ($\alpha 1(I)^{-null}$ and $\alpha 1(I)^{-stab}$, respectively) mice.

(B) Histomorphometric analysis of calluses at 1 wk following fracture, as indicated by cartilage volume (CV) as a percentage of total callus tissue volume (TV).

(C-E) Radiographic analysis from $\alpha 1(I)^{-Catnb^{null}}$ mice (C), from $\alpha 1(I)^{-Catnb^{stab}}$ mice (D), and from wild-type littermate mice (E) at 3 wk after fracture. Arrows indicate fracture site.

(F-K) HE staining from $\alpha 1(I)^{-Catnb^{null}}$ mice (F and I), from $\alpha 1(I)^{-Catnb^{stab}}$ mice (G and J), and from wild-type mice (H and K) shows that bone healing was inhibited in $\alpha 1(I)^{-Catnb^{null}}$ mice at 3 wk after fracture, with remaining cartilage at the fracture site. $\alpha 1(I)^{-Catnb^{stab}}$ mice showed an accelerated fracture repair with abundant new bone tissue formed. Images (F, G, and H) are 25 $\times$ magnifications; (I, J, and K) are 100 $\times$ magnifications of the areas shown in the box in the lower-magnification images.

(L) Relative radiopaque bone density at the fracture site shows decreased bone density in $\alpha 1(I)^{-Catnb^{null}}$ mice and increased bone density in $\alpha 1(I)^{-Catnb^{stab}}$ mice.

(M) Assay of bone volume (BV) as a percentage of total callus tissue volume (TV) shows that $\alpha 1(I)^{-Catnb^{null}}$ mice displayed inhibition of fracture healing, whereas osteoblast-specific β -catenin activation in $\alpha 1(I)^{-Catnb^{stab}}$ mice enhanced bone healing.

(N and O) Regulation of SOX9 at 1 wk and RUNX2 at 3 wk following fracture in $\alpha 1(I)^{-Catnb^{null}}$, $\alpha 1(I)^{-Catnb^{stab}}$, and wild-type mice.

doi:10.1371/journal.pmed.0040249.g004

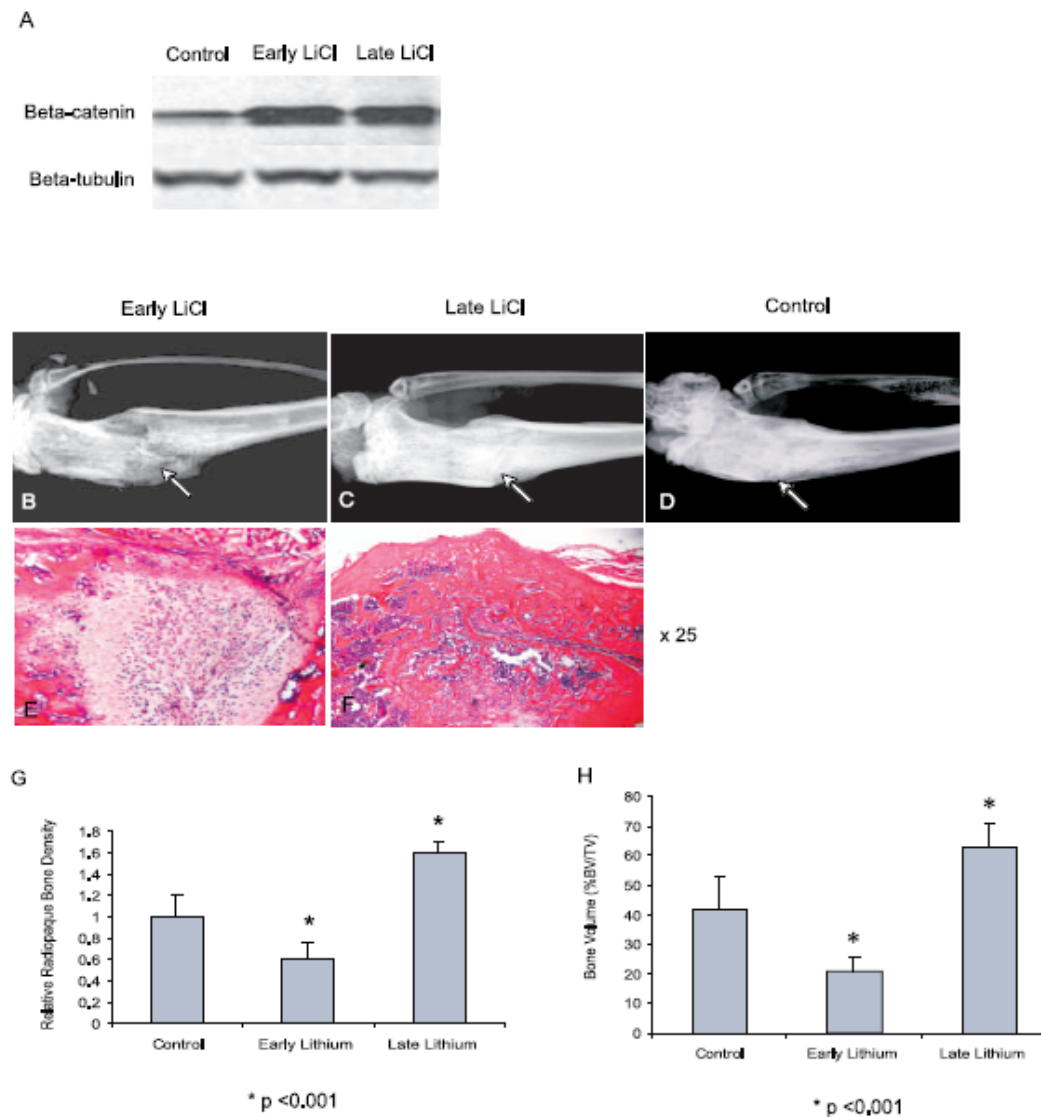


Figure 5. Lithium Treatment Regulates Bone Mass at the Fracture Site

Mice were treated with lithium starting either 2 wk before the fracture (early treatment) or 4 d after the fracture (late treatment).

(A) β -catenin increased 3 wk after fracture in mice receiving either early or late lithium treatment.

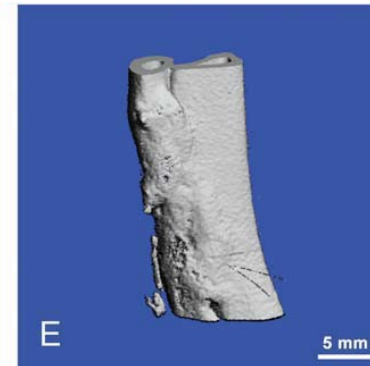
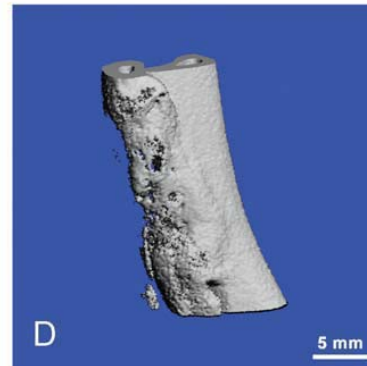
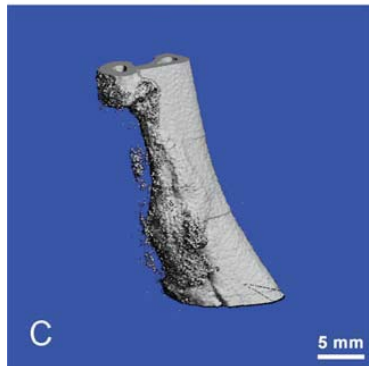
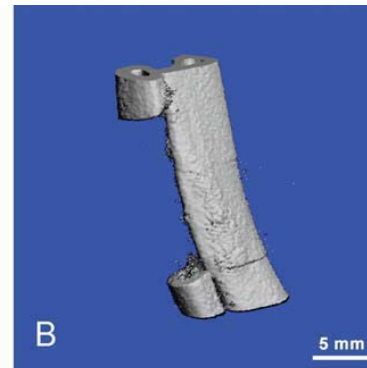
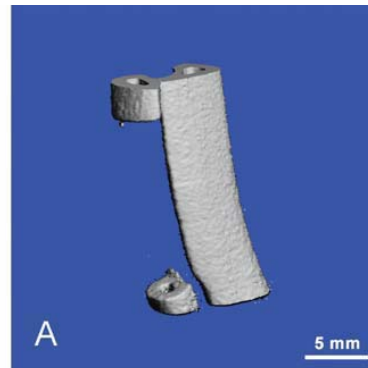
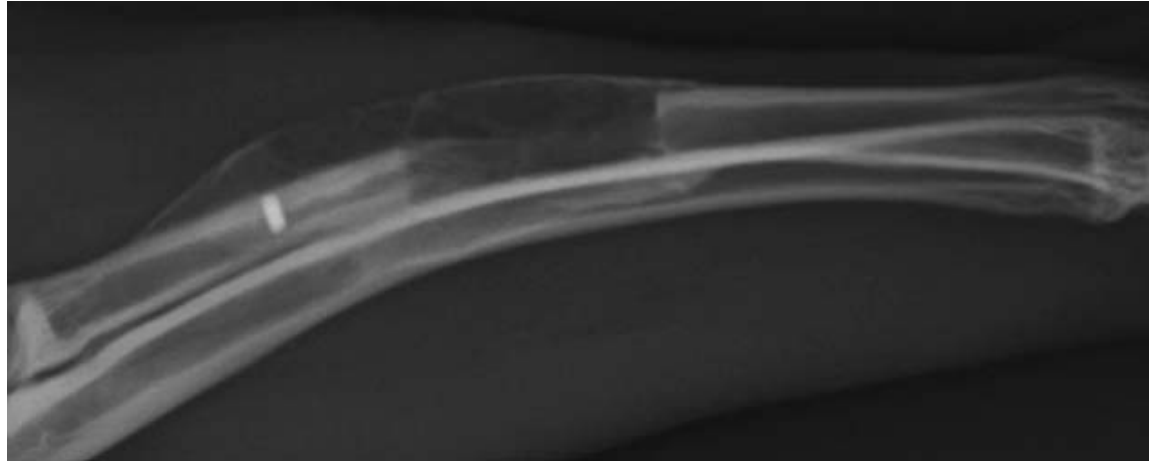
(B–D) Radiographic analysis from mice receiving early lithium treatment (B), late lithium treatment (C), or NaCl as a control (D), show that lithium enhanced fracture healing only when given late. Arrows indicate the fracture site.

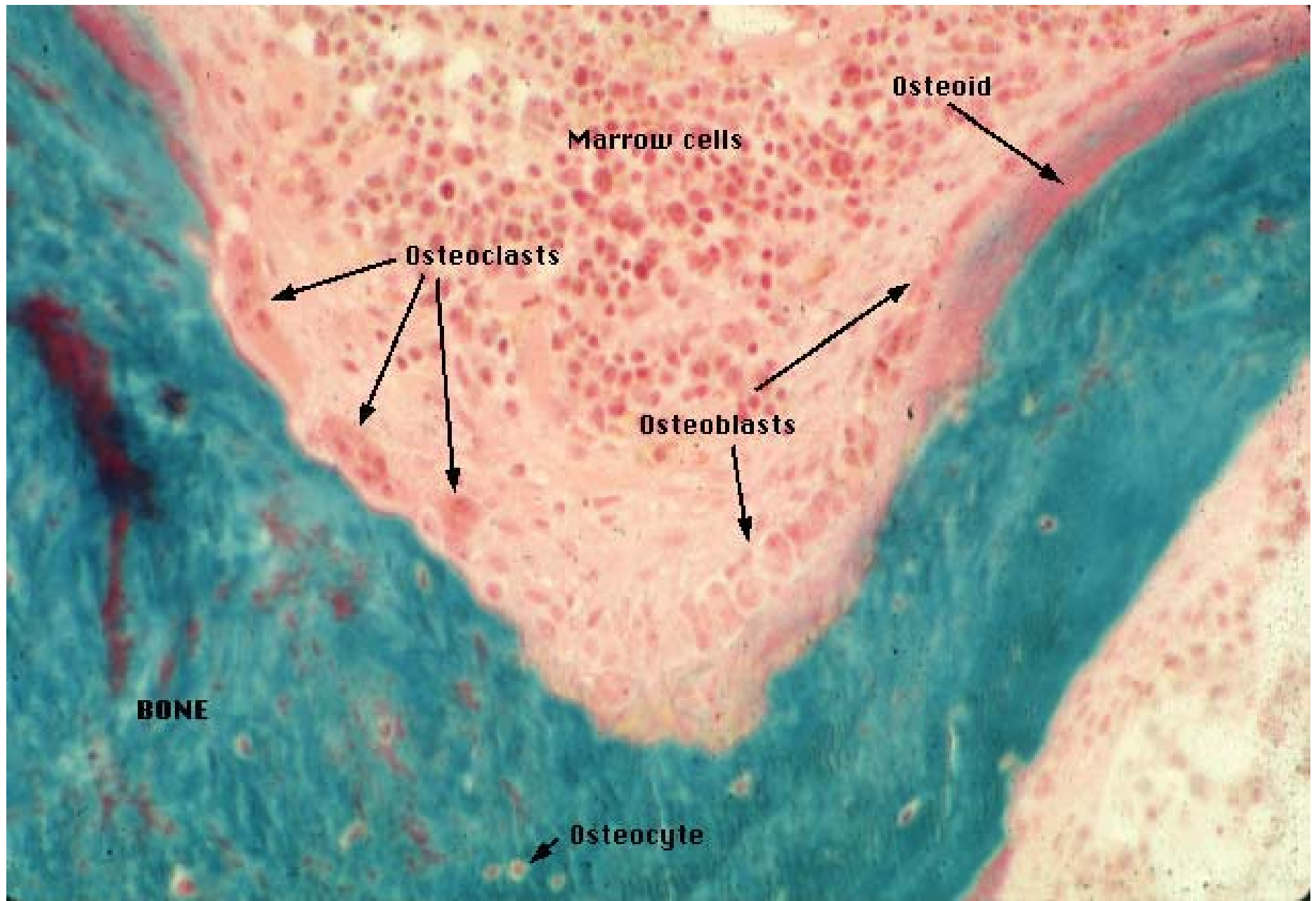
(E and F) HE staining from mice receiving early or late lithium treatments, respectively (25 $\times$). There is immature mesenchymal tissue at the fracture site in (E), and large amounts of bone in (F).

(G) Relative radiopaque bone density results show that early and late treatment have opposite effects on fracture healing.

(H) Histomorphometric analysis on bone volume (BV) as a percentage of total callus tissue volume (TV), showing that pharmacologic activation of β -catenin after healing cells become committed to the osteoblast lineage could improve healing, while treatment at earlier time points inhibited fracture repair.

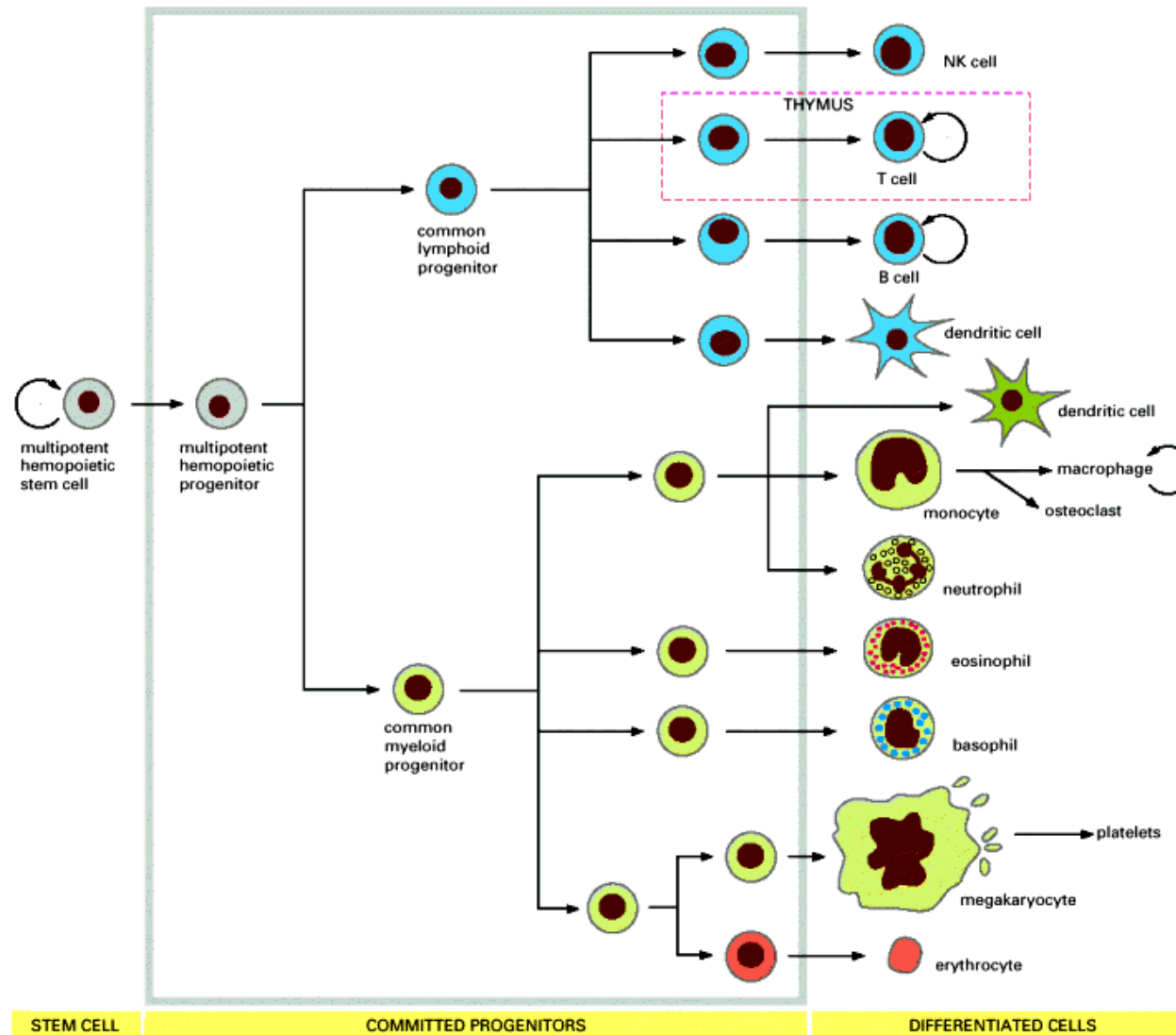
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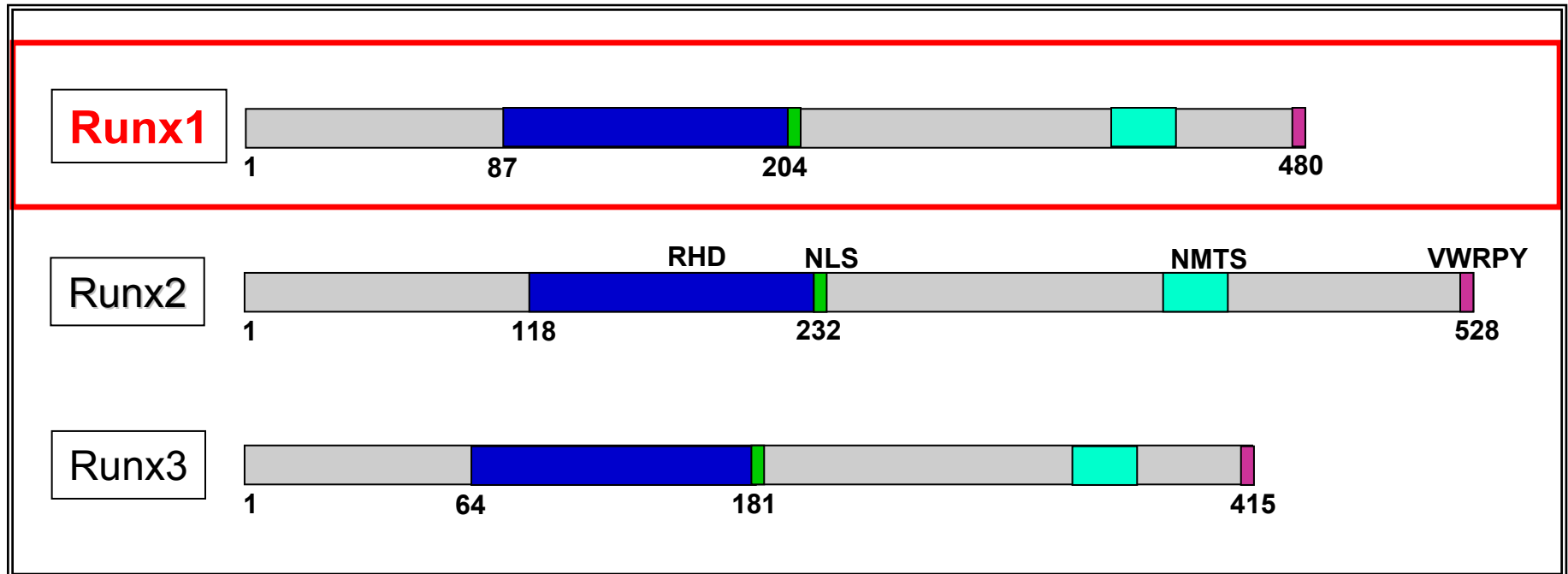
MECANISMOS MOLECULARES DE LA DIFERENCIACIÓN CELULAR

MODELO DE FORMACIÓN DE LAS CÉLULAS DE LA SANGRE



EL FACTOR DE TRANSCRIPCIÓN RUNX1

DOMINIOS FUNCIONALES DE LAS PROTEINAS RUNX



RHD *Runt Homology Domain (DNA binding, Protein-Protein interaction)*

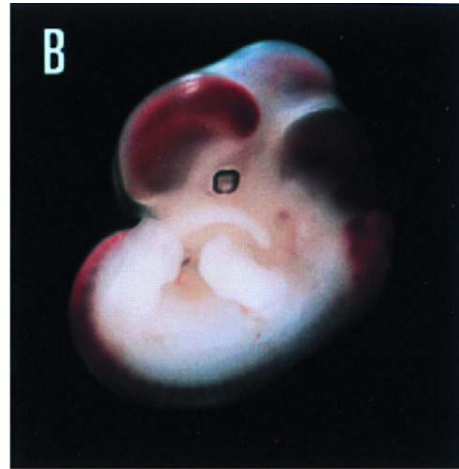
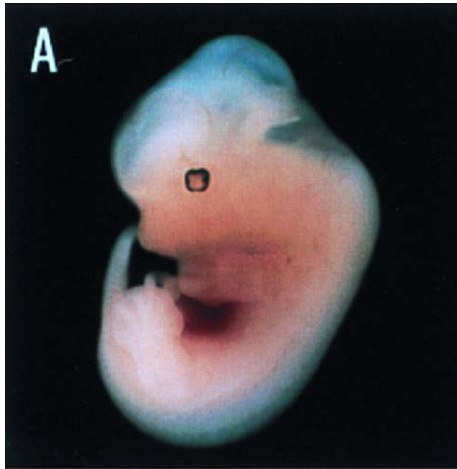
NSL *Nuclear Signaling Localization*

NMTS *Nuclear Matriz Targeting Signal (PST-Rich Domain)*

VWRPY *Secuencia conservada de aminoácidos (Groucho/TLE)*

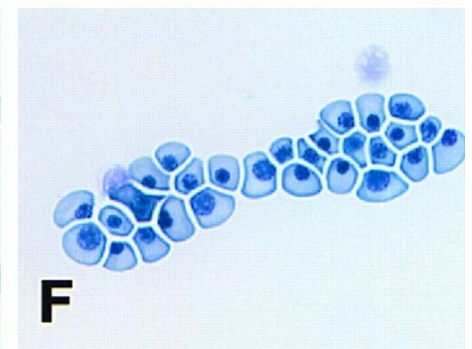
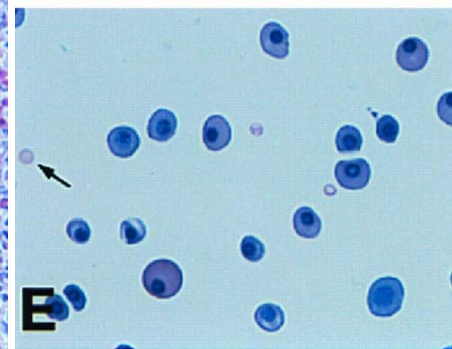
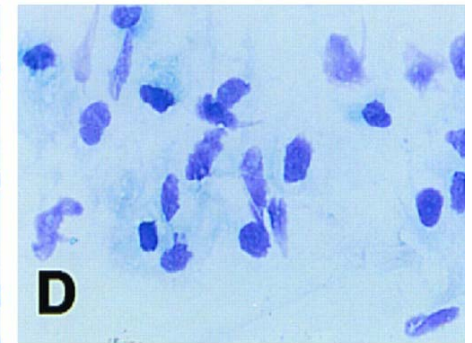
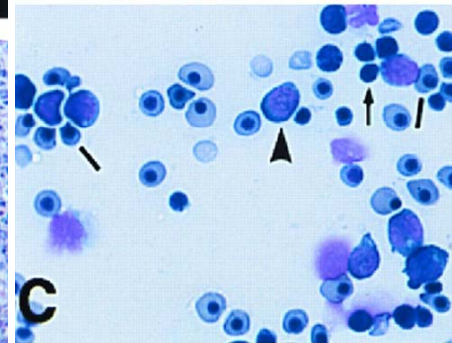
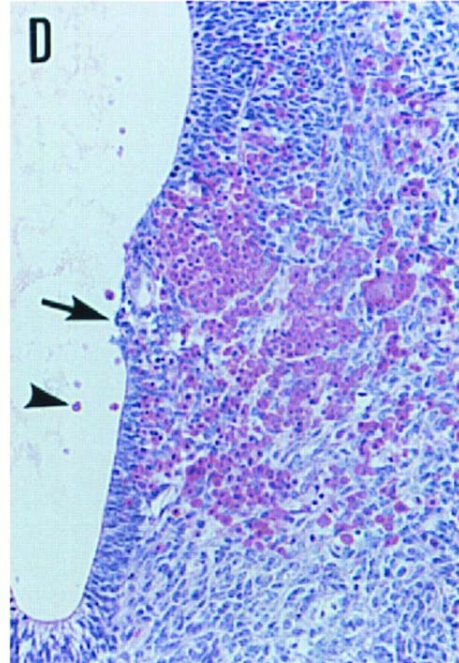
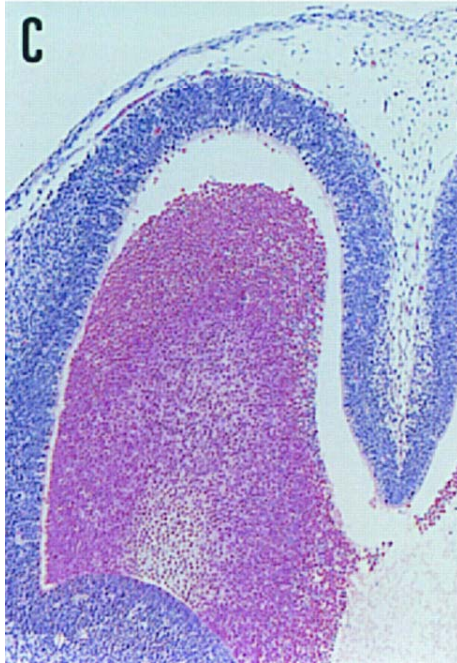
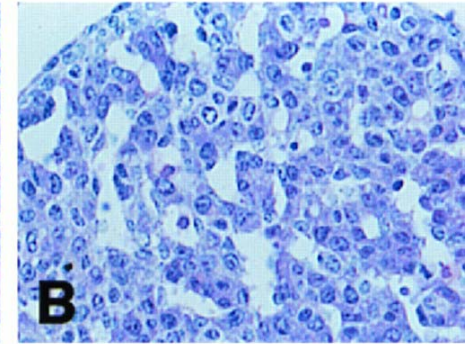
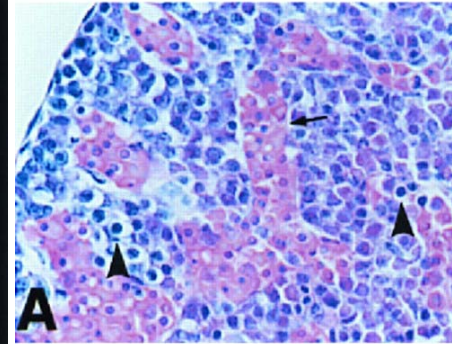
GENES REGULADOS POR RUNX1

CATEGORY	REFERENCE
<u>Cytokines</u> -- Granulocyte-macrophage colony stimulating factor (GM-CSF)..... -- Interleukin-3 (IL-3) -- Macrophage inflammatory protein 1-α (MIP-1α)	[Takahashi et al., 1995] [Cameron et al., 1994] [Bristow and Shore, 2003]
<u>Cell surface differentiation markers</u> - T cell receptor (TCR) α, β, δ, γ subunits..... - Macrophage colony stimulating factor-receptor 1 (M-CSF-R1)..... - Transforming growth factor β receptor-1 (TGFβ-R1)..... - Cell surface glycoprotein CD4, CD3ϵ, CD36..... - Cell surface glycoprotein CD-11a integrin..... - Immunoglobulin μ heavy chain (Igα)..... - Immunoglobulin μ heavy chain (IgH).....	[Hallberg et al., 1992; Prosser et al., 1992; Redondo et al., 1992] [Giese et al., 1995] [Zhang et al., 1994] [Ji et al., 1998] [Taniuchi et al., 2002] [Puig-Kroger et al., 2003] [Pardali et al., 2000] [Pardali et al., 2000]
<u>B-cell specific genes</u> - Granzyme B..... - B cell specific tyrosine kinase..... - $\lambda 5$	[Babichuk and Bleackley, 1997] [Libermann et al., 1999] [Martensson et al., 2001]
<u>Myeloid specific genes</u> - Mouse mast cell protease-6 (MMCP-6)..... - Myeloperoxidase MPO)..... - Neutrophil elastase - P14(ARF).....	[Ogihara et al., 1999] [Britos-Bray and Friedman, 1997] [Nuchprayoon et al., 1994] [Linggi et al., 2002]

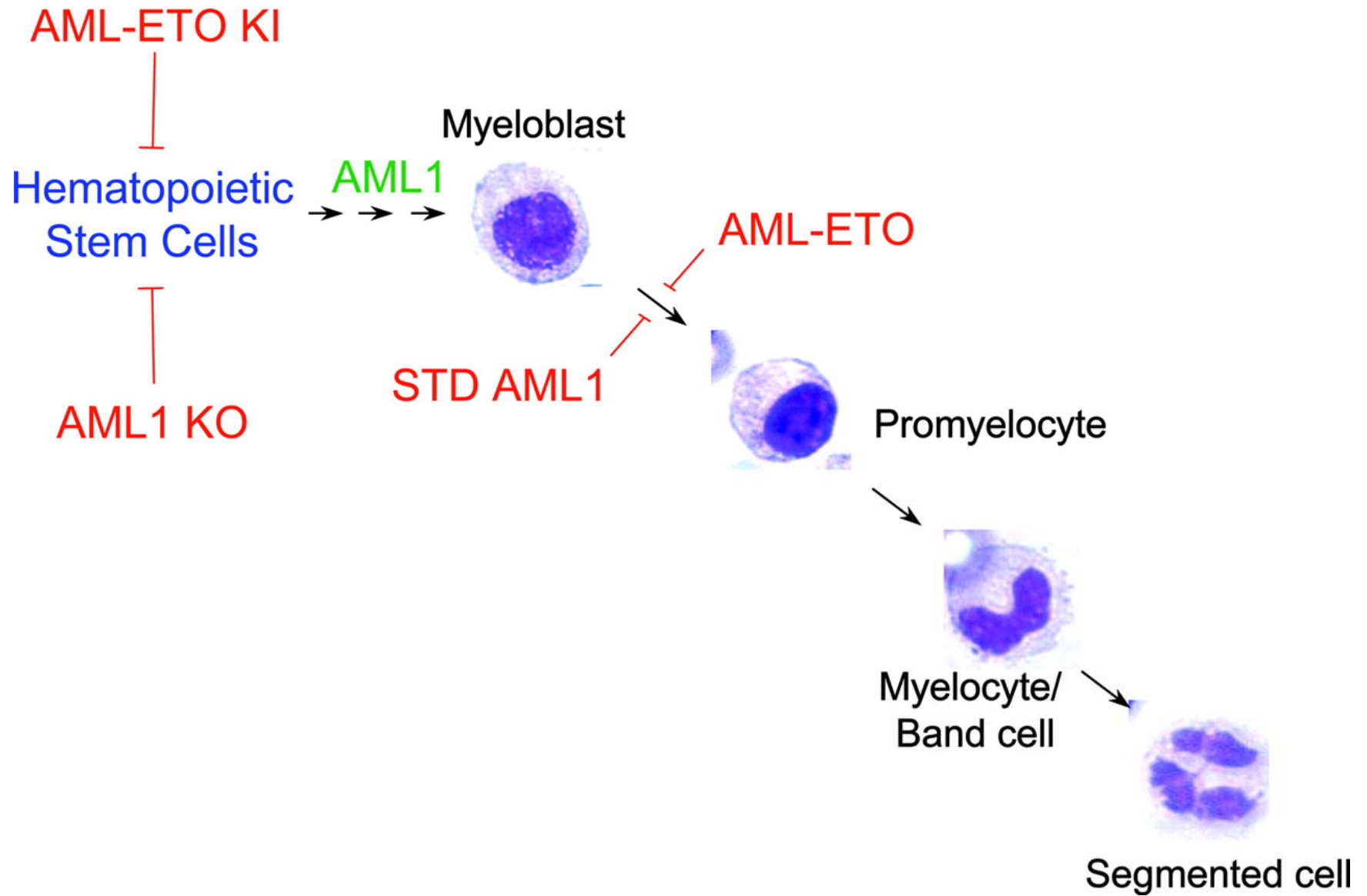


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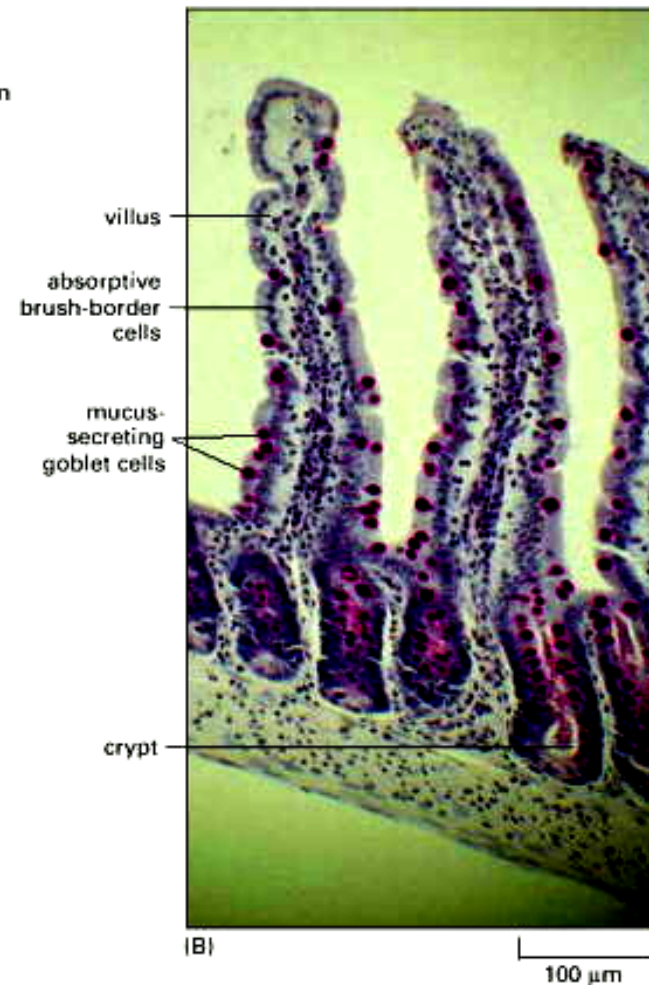
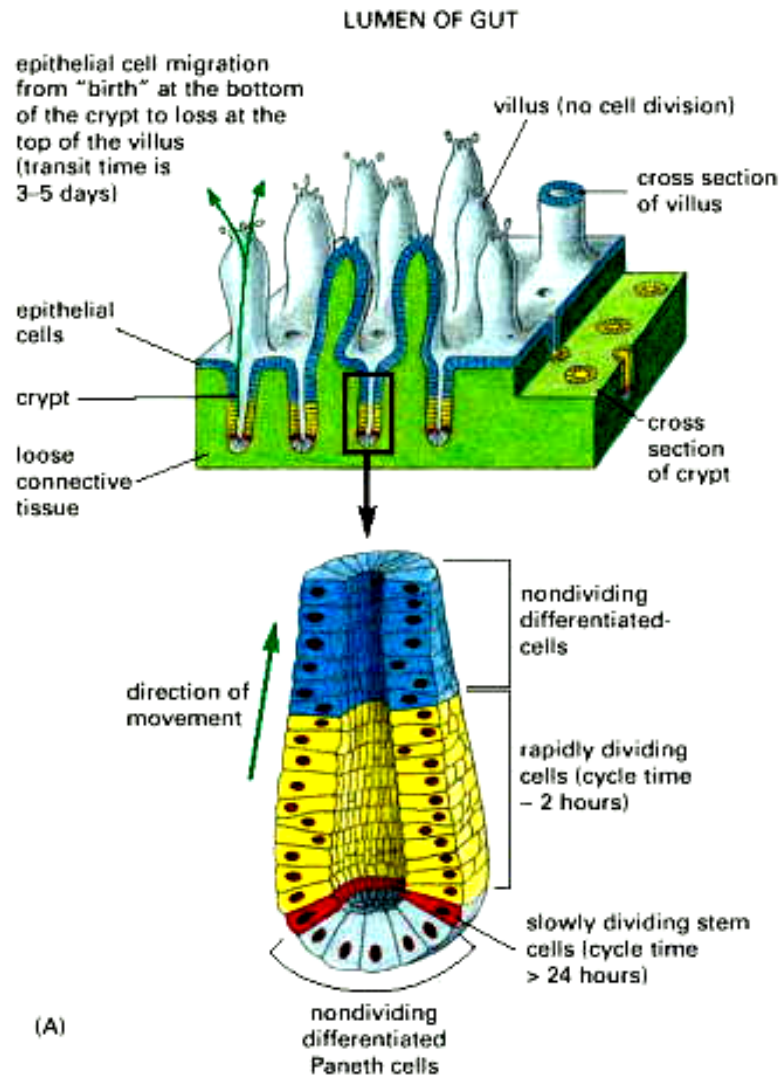


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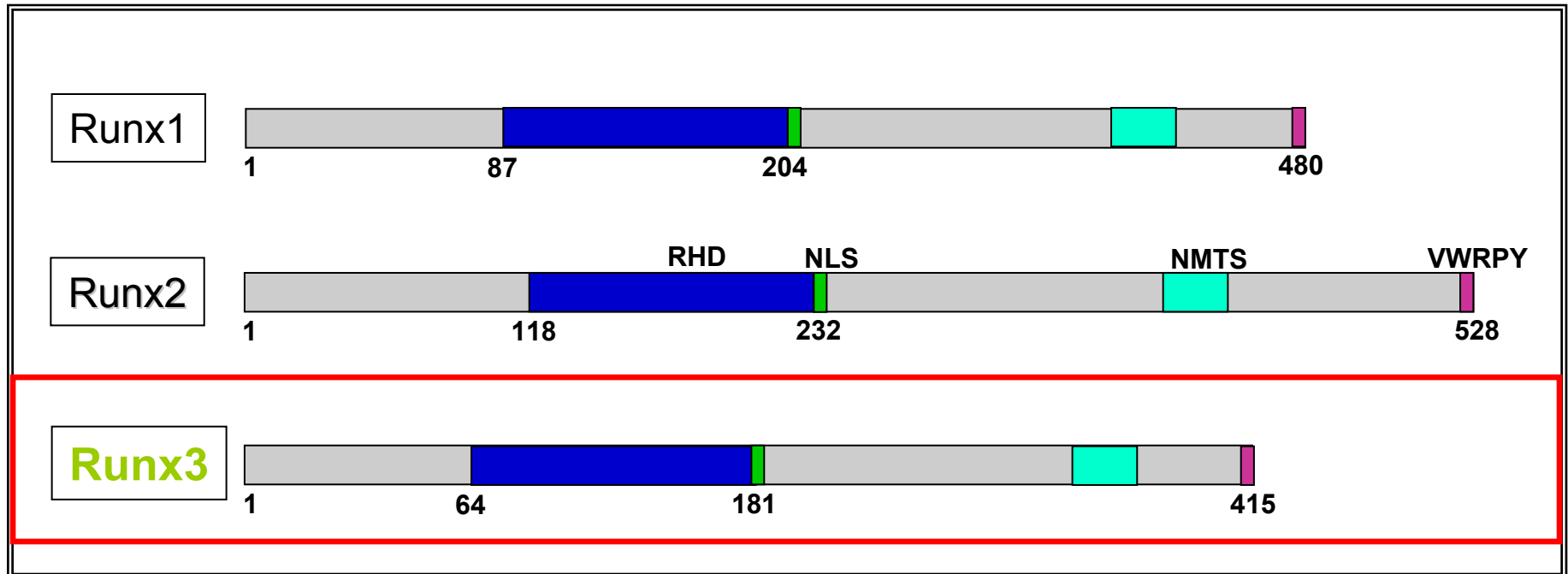
MECANISMOS MOLECULARES DE LA DIFERENCIACIÓN CELULAR

MODELO DE FORMACIÓN DEL SISTEMA GASTRO-INTESTINAL



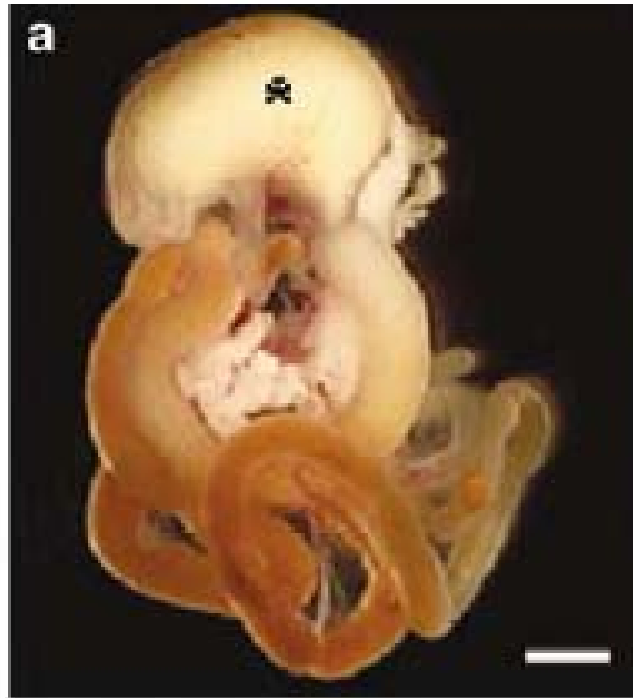
EL FACTOR DE TRANSCRIPCIÓN RUNX3

DOMINIOS FUNCIONALES DE LAS PROTEINAS RUNX

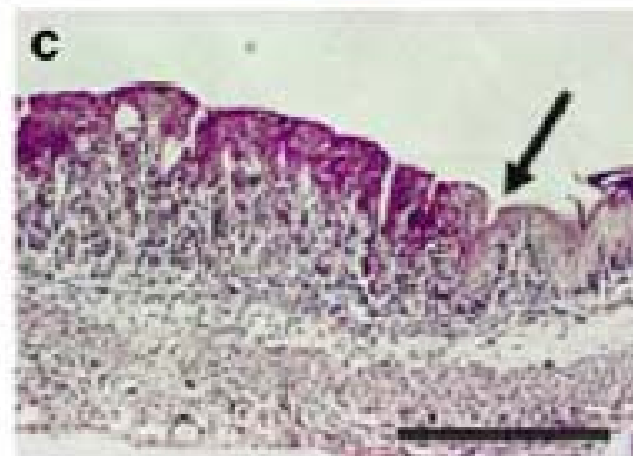
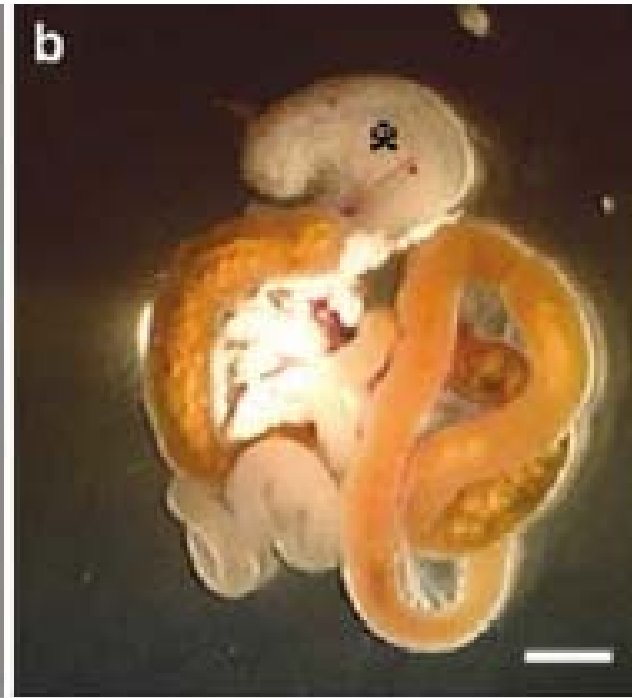


RHD	<i>Runt Homology Domain (DNA binding, Protein-Protein interaction)</i>
NSL	<i>Nuclear Signaling Localization</i>
NMTS	<i>Nuclear Matriz Targeting Signal (PST-Rich Domain)</i>
VWRPY	<i>Secuencia conservada de aminoácidos (Groucho/TLE)</i>

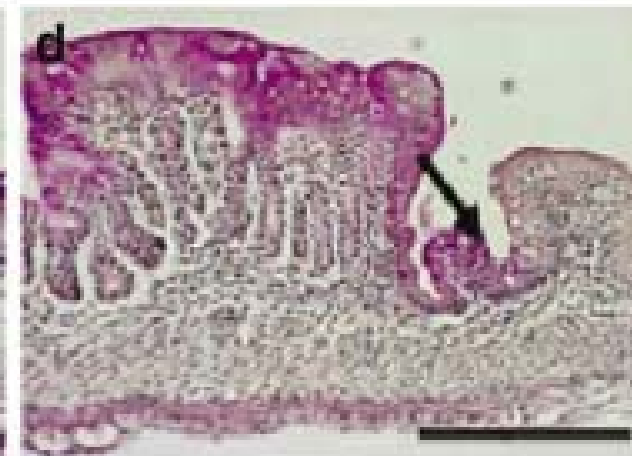
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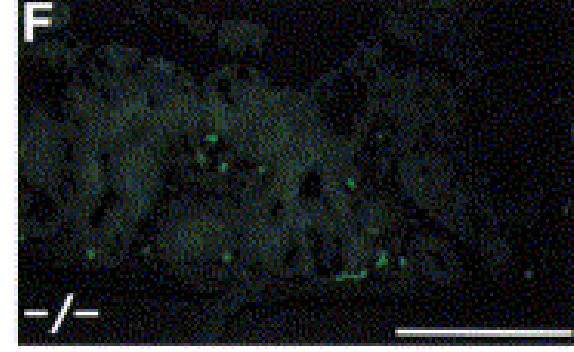
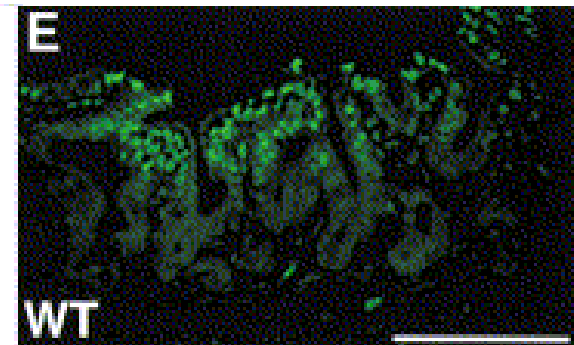
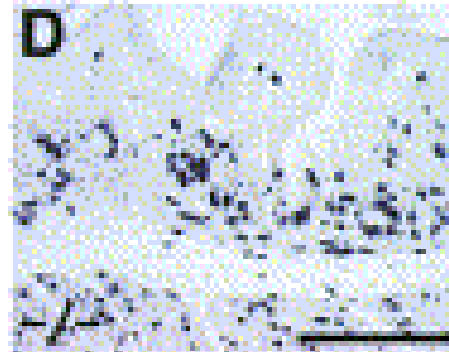
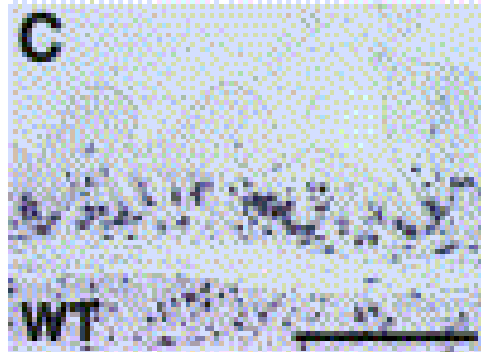
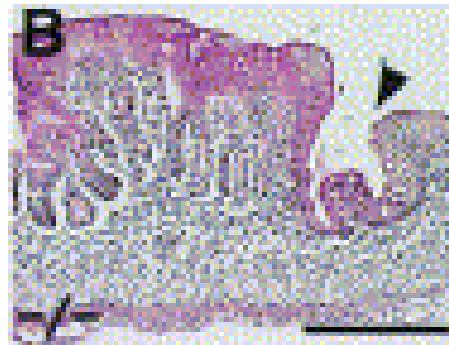
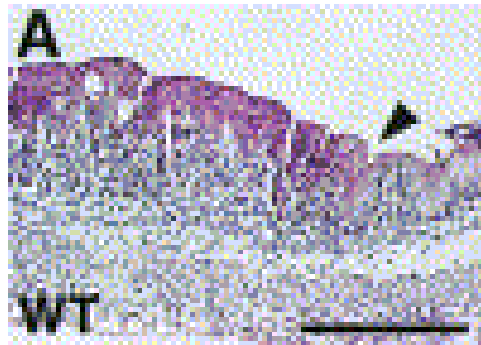
RUNX3 ^{-/-}



WT



RUNX3 ^{-/-}



MARCADOR DE PROLIFERACION CELULAR

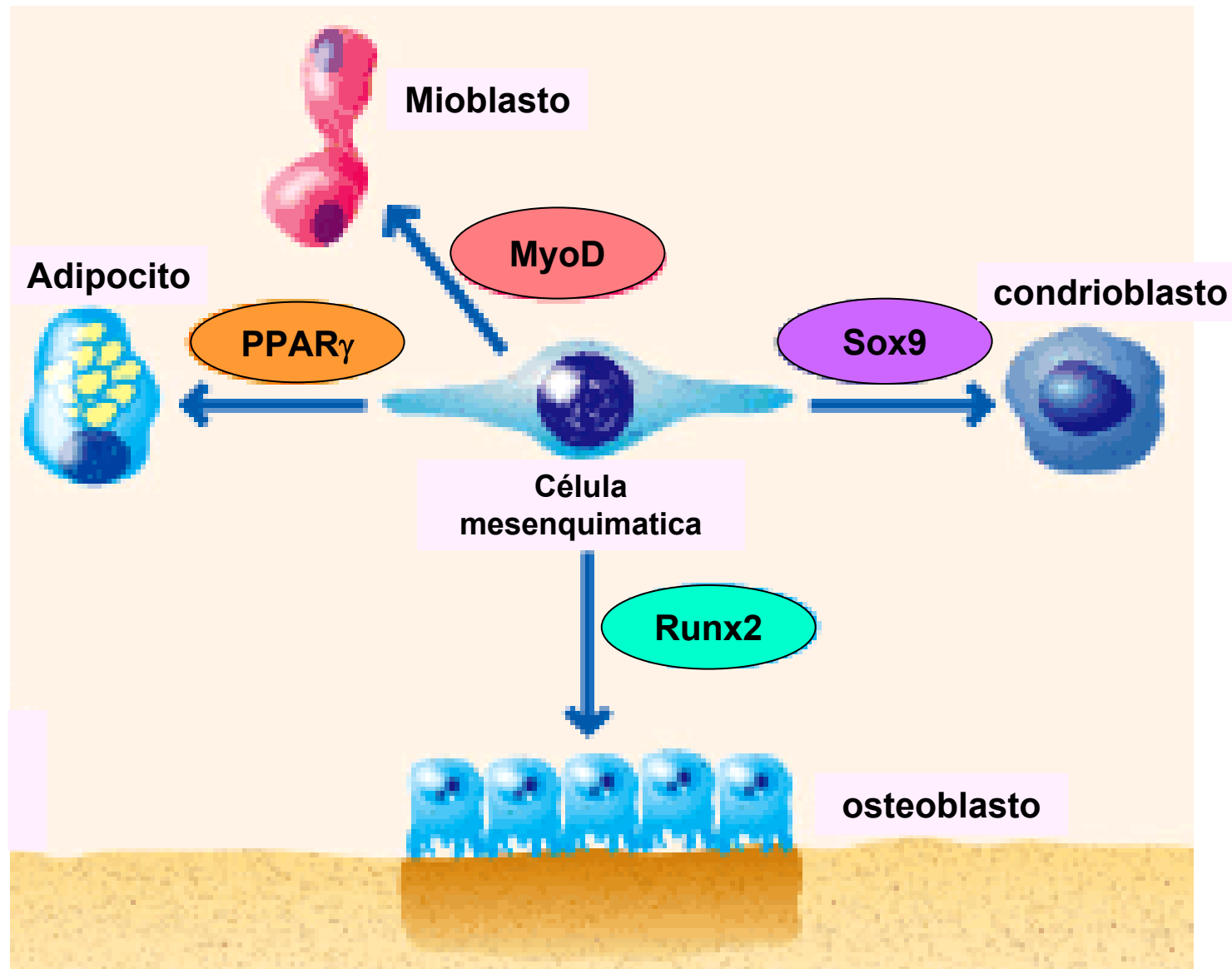
MARCADOR DE APOPTOSIS

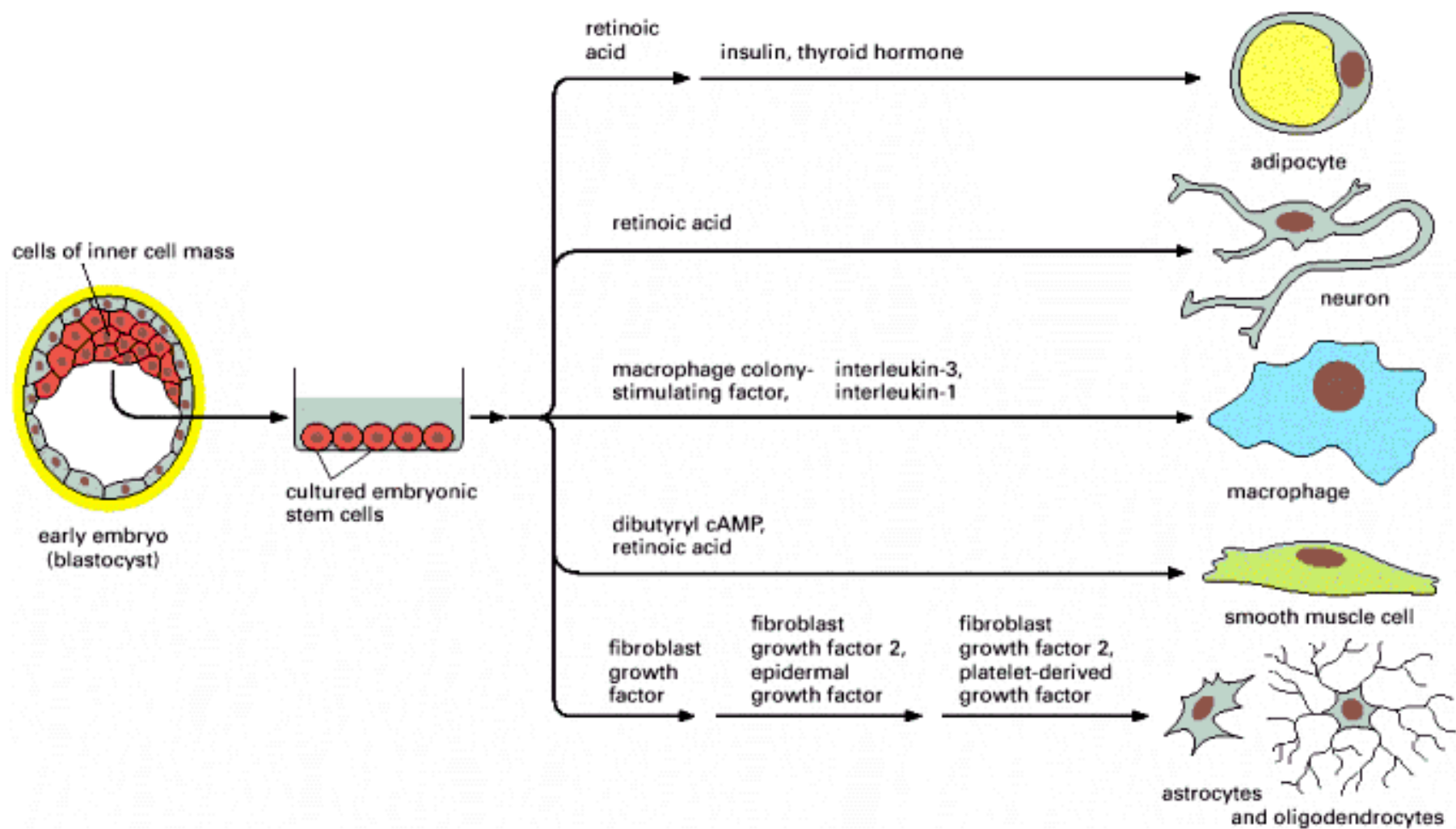
FAMILIA DE PROTEINAS RUNX

Factores de Transcripción Familia Runx	Función esencial propuesta	Efecto observado en ratones con inactivación homocigótica	Patología asociada a su deficiencia en humanos
Runx1	Hematopoyesis	Muerte fetal por falla en hematopoyesis (1)	Leucemia (2) (mieloide)
Runx3	Neurogénesis Timogénesis Desarrollo del tracto gastrointestinal	Desarrollo anormal del epitelio gástrico y de la raíz ganglionar dorsal (3)	Cáncer Gástrico (4)
Runx2	Osteogénesis Condrogénesis	Ratones de termino, pero inviiables debido a la ausencia de huesos (5)	Displasia Cleidocraneal (6)

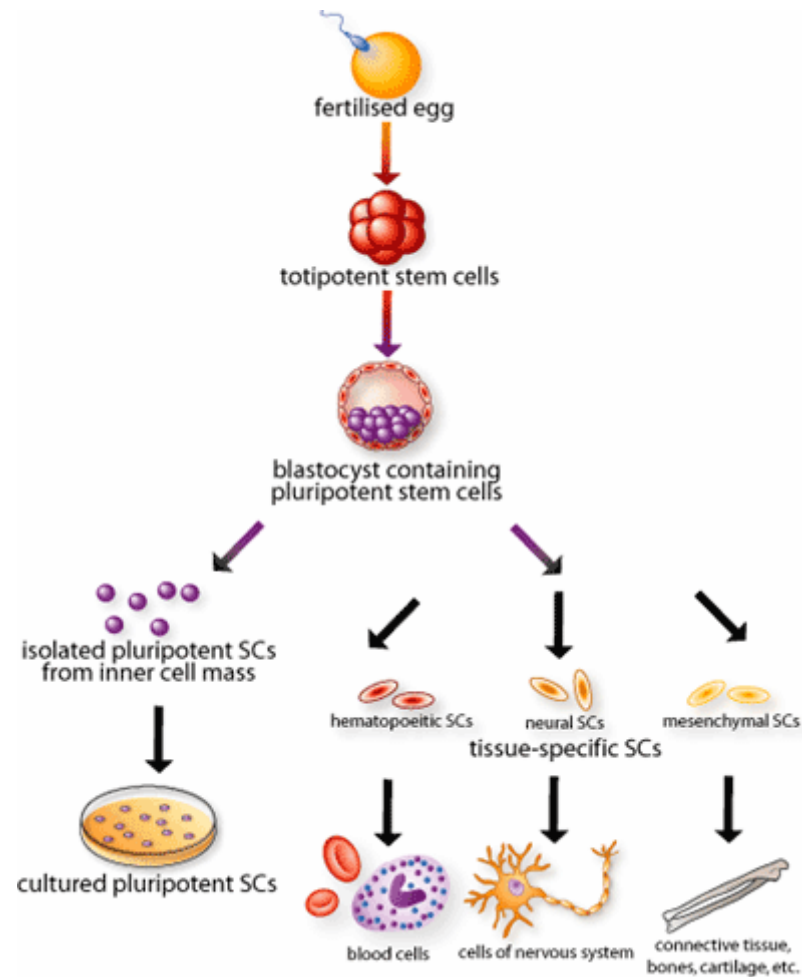
Ref.: (1) Wang y col., 1996; (2) Okuda y col., 1996; (3) Levanon y col., 2002; (4) Li y col., 2002;
(5) Komori y col., 1997; (6) Otto y col., 1997.

**STEM CELL
TERAPIA CELULAR
BASADA EN CELULAS
CON POTENCIAL DE DIFERENCIACIÓN**

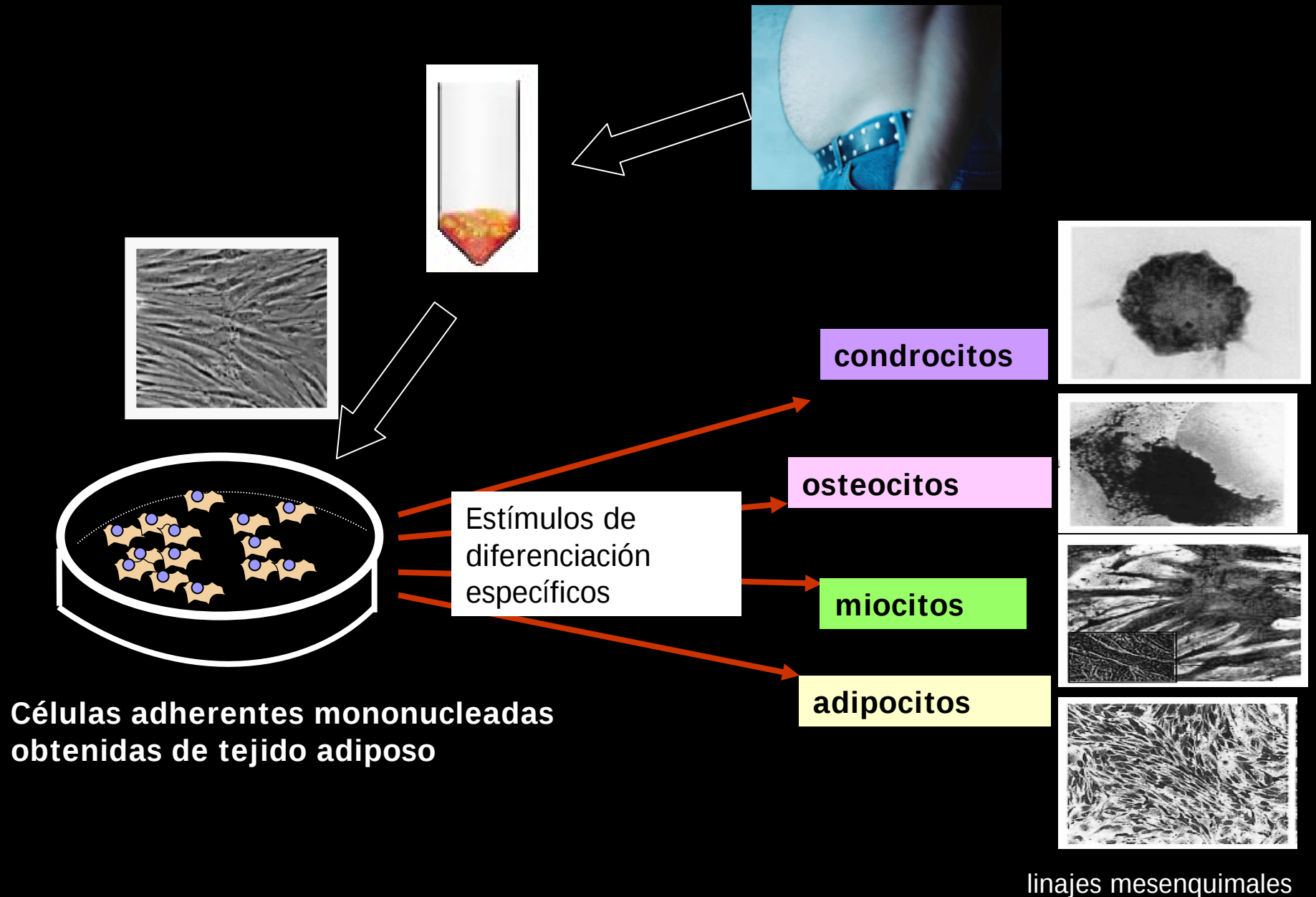




Aislamiento de stem cell embrionarias y diferenciación celular para terapia de reemplazo de células dañadas en tejidos específicos



Experimento de Zuk et al. Tissue Engineering 7 : 211-228, 2001



Aislamiento y diferenciación de células troncales de tejidos en adultos y terapia celular

