



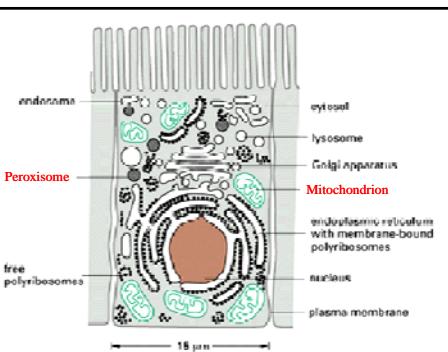
Universidad de Chile

Programa Académico de Bachillerato

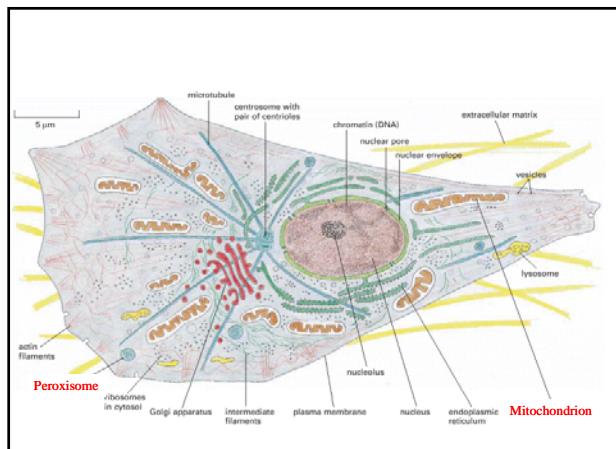
Introducción a la Biología Celular

2014

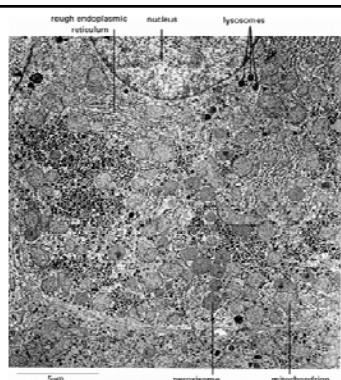
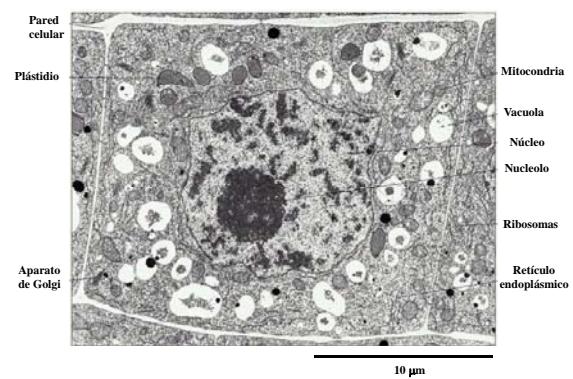
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The major intracellular compartments of an animal cell. The cytosol (gray), endoplasmic reticulum, Golgi apparatus, nucleus, mitochondrion, endosome, lysosome, and peroxisome are distinct compartments isolated from the rest of the cell by at least one selectively permeable membrane.



Todas las células tienen el mismo conjunto básico de organelos conformados por membranas



Electron micrograph of part of a liver cell seen in cross-section. Examples of most of the major intracellular compartments are indicated.



Kölliker (1850-1890) describió arreglos de gránulos en el sarcoplasma de músculo estriado, que fueron llamados posteriormente *sarcosomas* por Retzius (1890)

Fleming (1890) describió estructuras filamentosas en el citoplasma de muchos tipos celulares distintos.

Altman (1890) desarrolló una tinción específica para esas estructuras. Sugirió su autonomía (unidades vivas elementales) y notó su similitud con las bacterias (viven de manera independiente o en colonias en el citoplasma de la célula)

Benda (fines del siglo XIX) usa la palabra **mitocondria** que vino a reemplazar a los términos blefaroblastos, condriocitos, condriomitos, condrioplastos, condriosomas, condrioesferas, fila, cuerpos intersticiales, mitogel, cuerpos parabasales, esferoplastos, vermiculos.

Table 12-1. Relative Volumes Occupied by the Major Intracellular Compartments in a Liver Cell (Hepatocyte)

INTRACELLULAR COMPARTMENT	PERCENTAGE OF TOTAL CELL VOLUME
Cytosol	54
Mitochondria	22
Rough ER cisternae	9
Smooth ER cisternae plus	6
Golgi cisternae	
Nucleus	6
Peroxisomes	1
Lysosomes	1
Endosomes	1

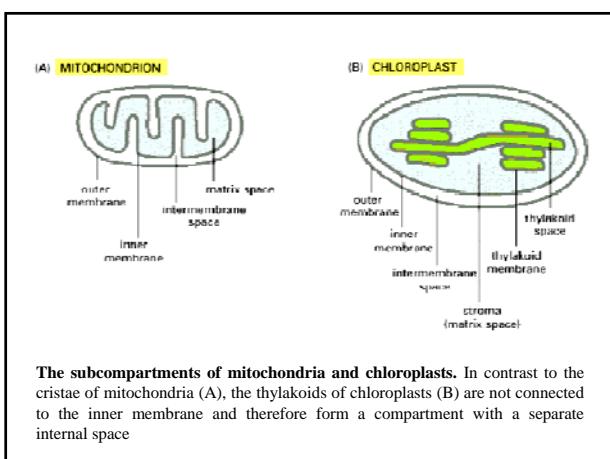
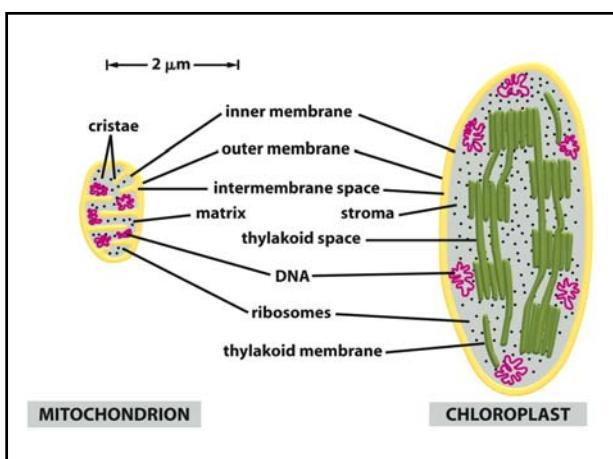
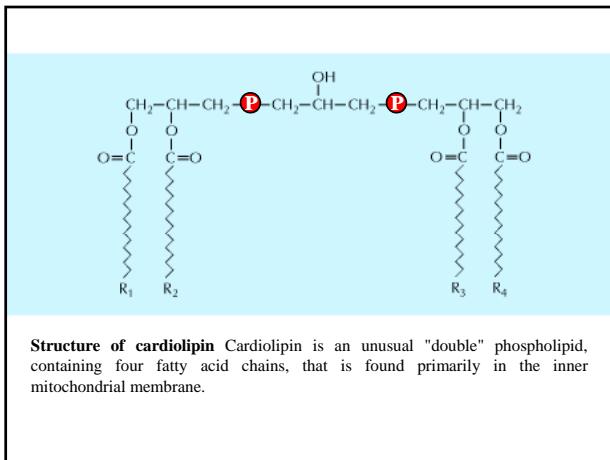
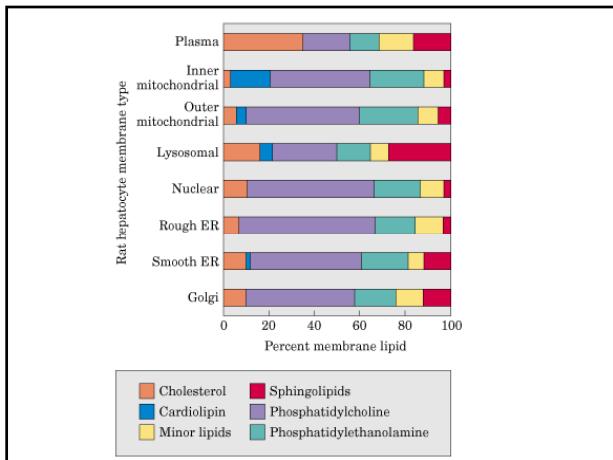
Table 12-2. Relative Amounts of Membrane Types in Two Kinds of Eucaryotic Cells

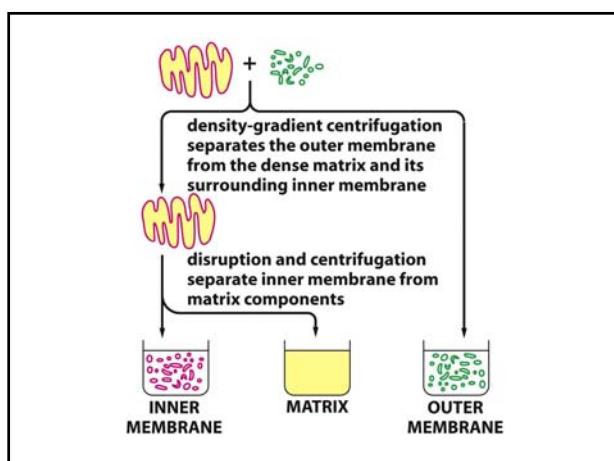
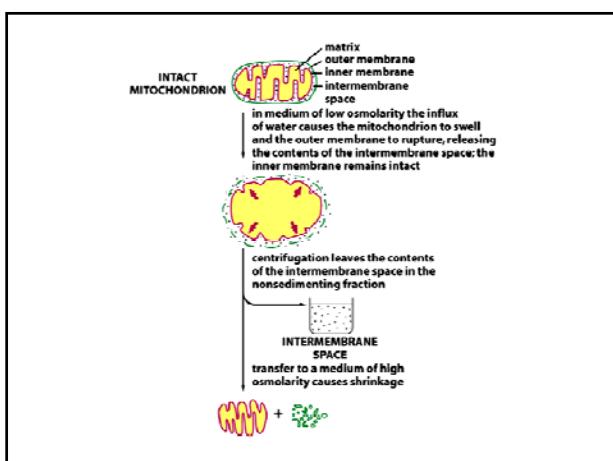
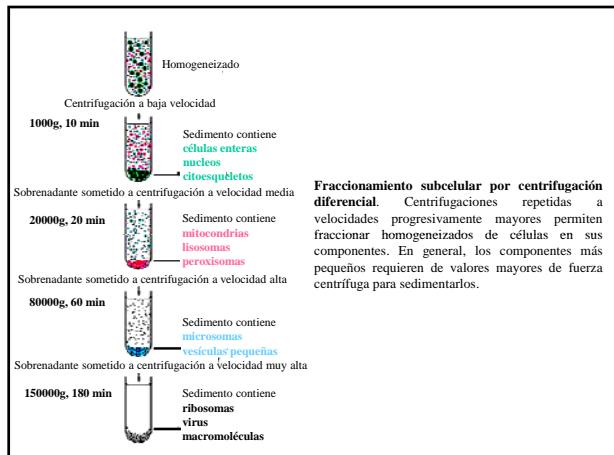
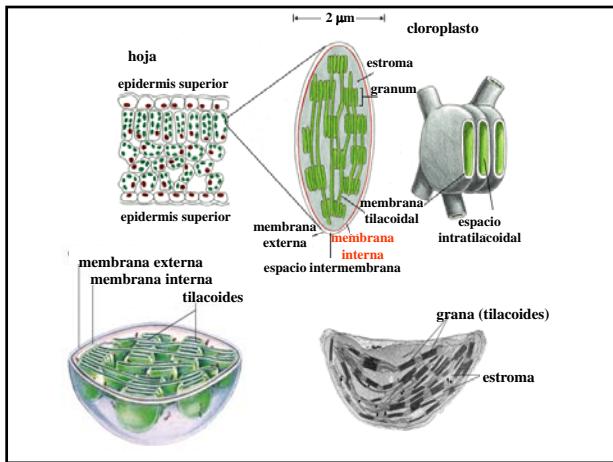
MEMBRANE TYPE	PERCENTAGE OF TOTAL CELL MEMBRANE	
	LIVER HEPATOCYTE*	PANCREATIC EXOCRINE CELL*
Plasma membrane	2	5
Rough ER membrane	35	60
Smooth ER membrane	16	<1
Golgi apparatus membrane	7	10
Mitochondria		
Outer membrane	7	4
Inner membrane	32	17
Nucleus		
Inner membrane	0.2	0.7
Secretory vesicle membrane	not determined	3
Lysosome membrane	0.4	not determined
Peroxisome membrane	0.4	not determined
Endosome membrane	0.4	not determined

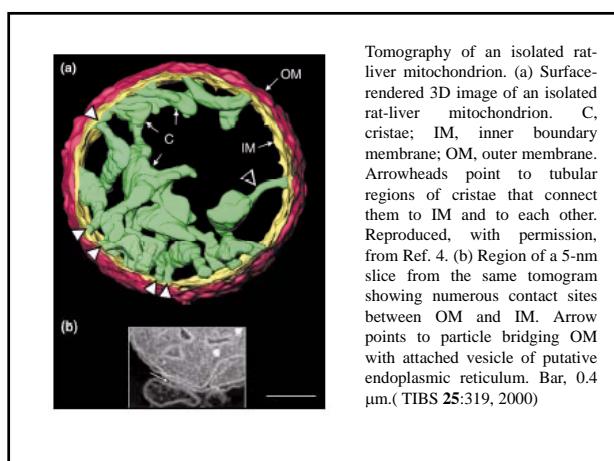
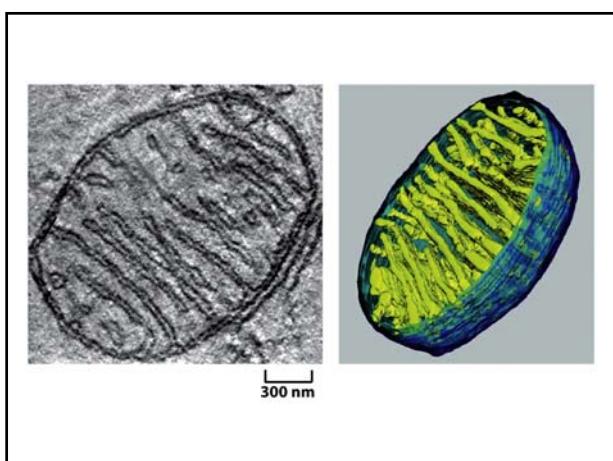
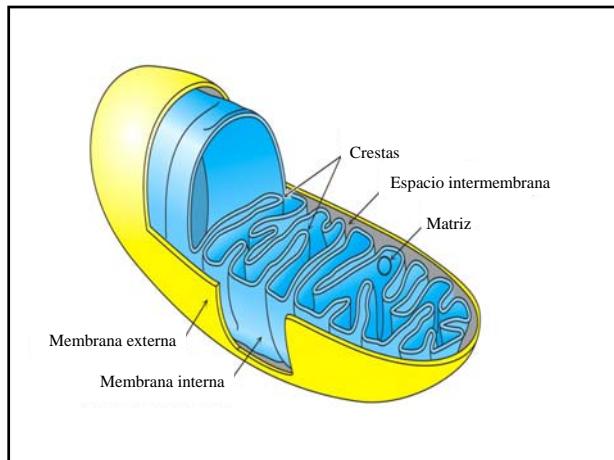
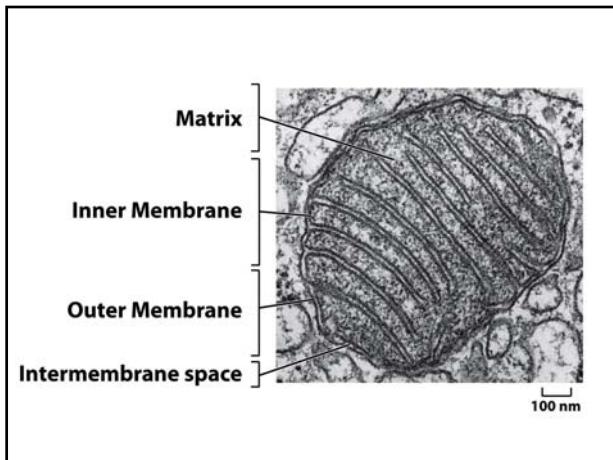
Approximate Lipid Compositions of Different Cell Membranes

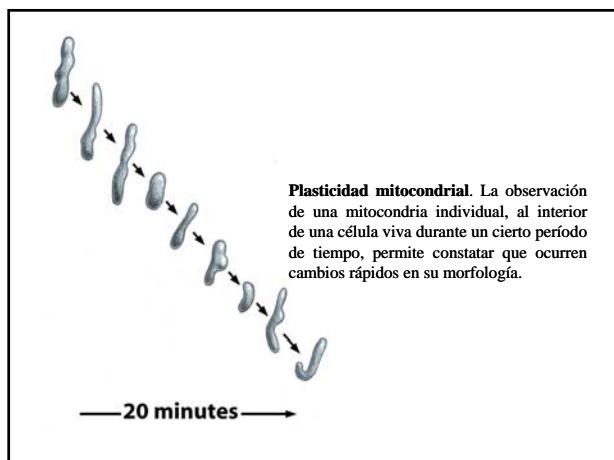
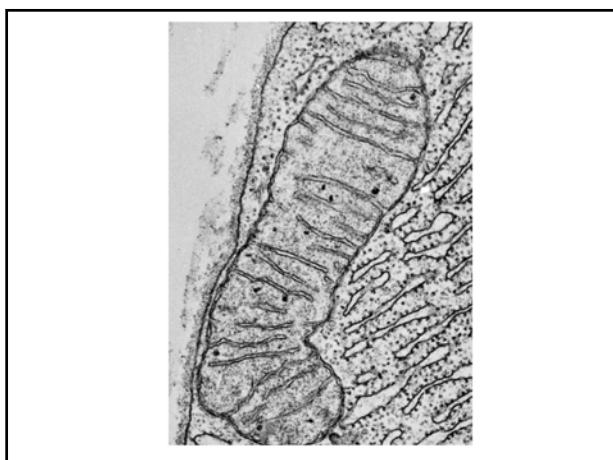
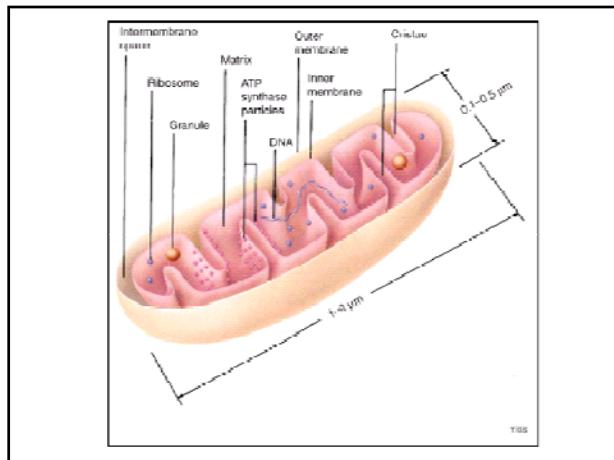
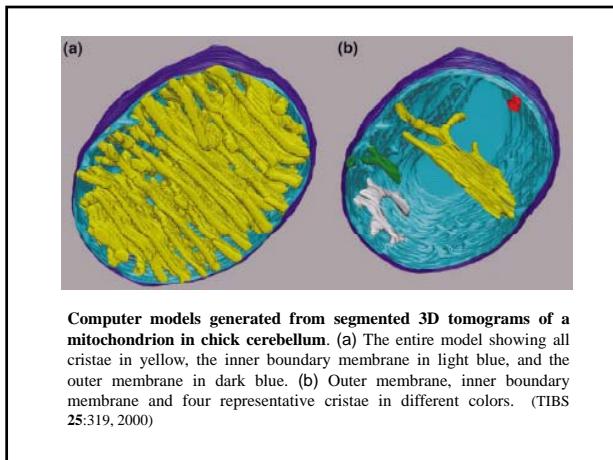
LIPID	PERCENTAGE OF TOTAL LIPID BY WEIGHT					
	LIVER CELL*	RBC*	MYELIN	MIT**	ER	
Cholesterol	17	23	22	3	6	0
Phosphatidylethanolamine	7	18	15	25	17	70
Phosphatidylserine	4	7	9	2	5	trace
Phosphatidylcholine	24	17	10	39	40	0
Sphingomyelin	19	18	8	0	5	0
Glycolipids	7	3	28	trace	trace	0
Others	22	13	8	21	27	30

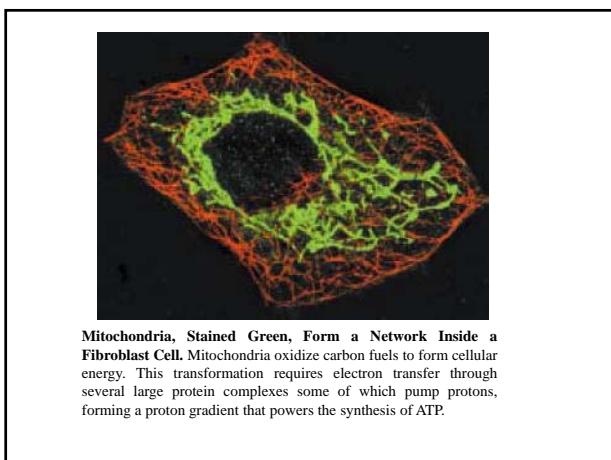
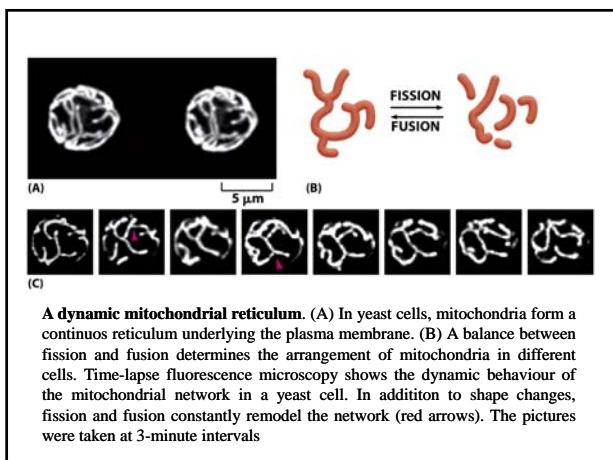
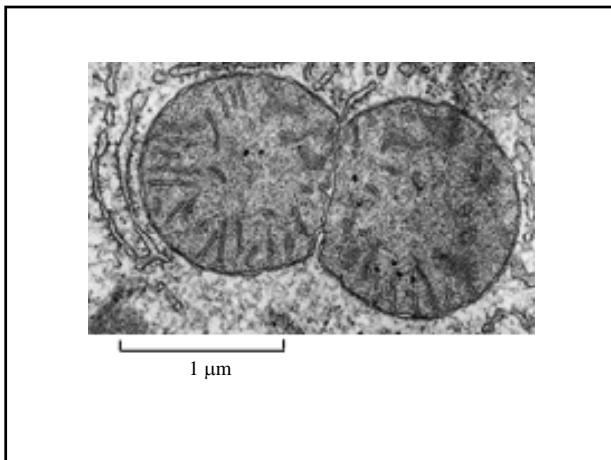
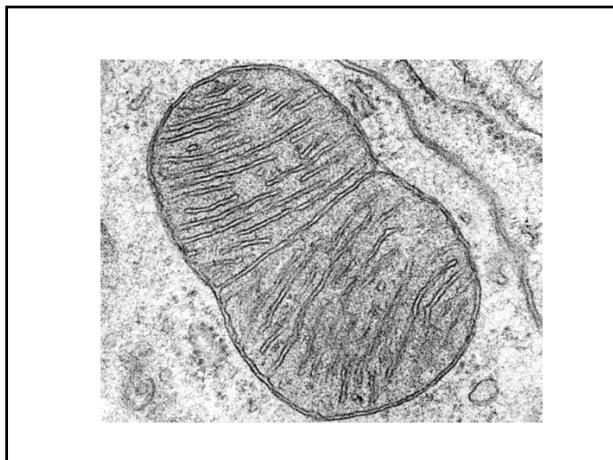
* Plasma membranes; ** Inner and Outer membranes

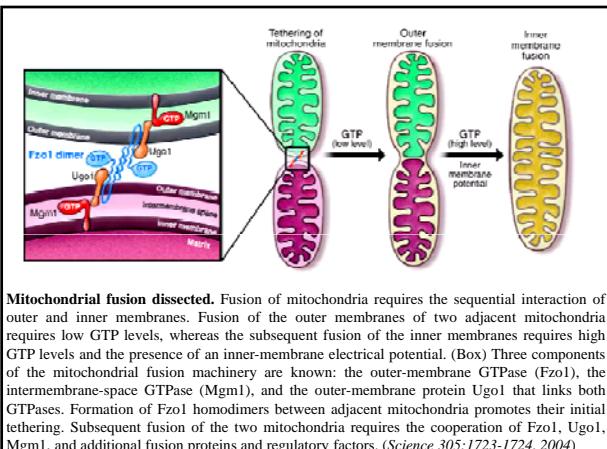










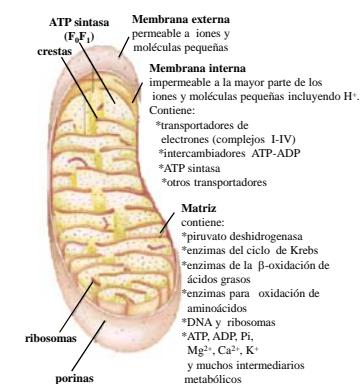


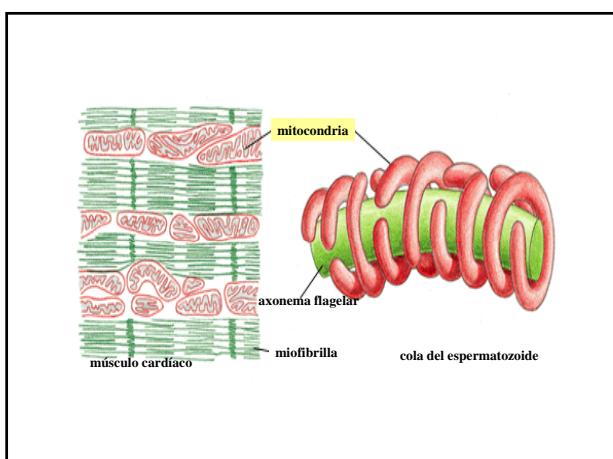
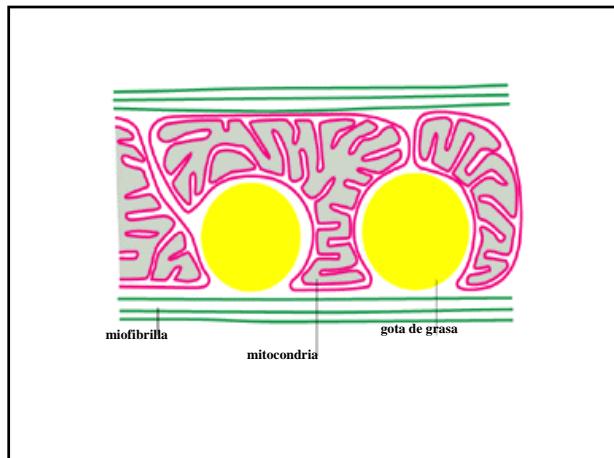
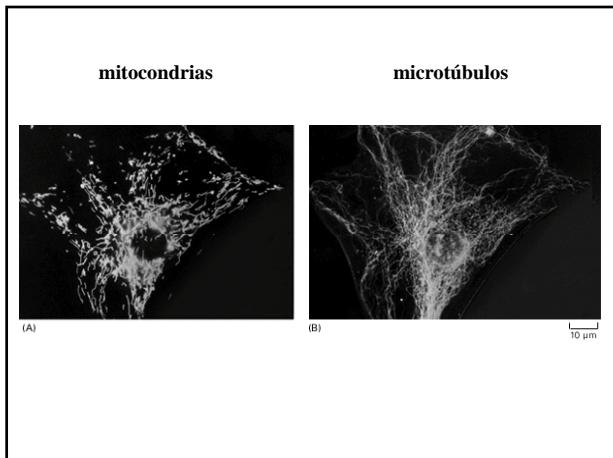
Mitochondria are dynamic organelles whose morphologies are controlled by fusion and fission. Mitochondrial fusion and fission are essential for normal mitochondrial function, implying that mitochondria do not function well as autonomous organelles.

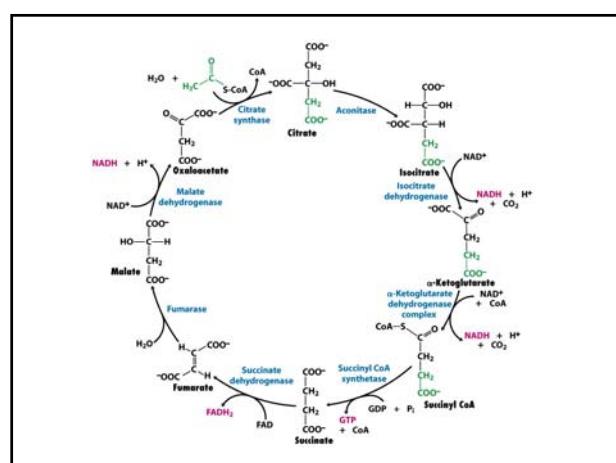
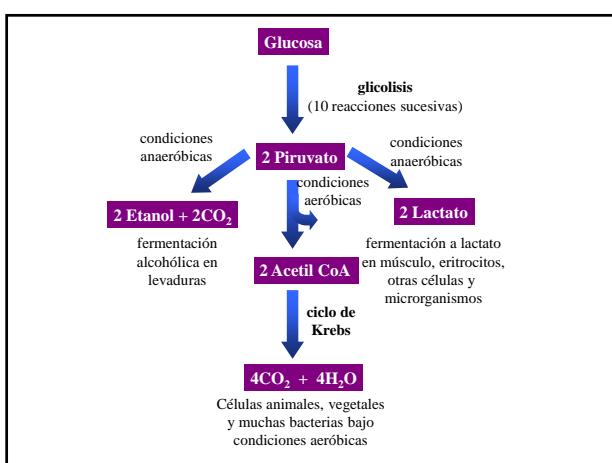
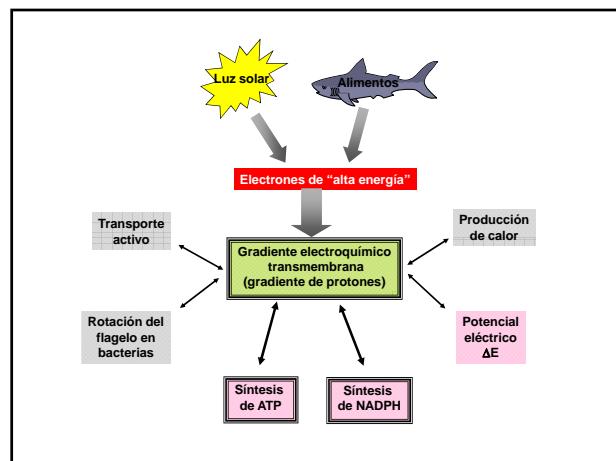
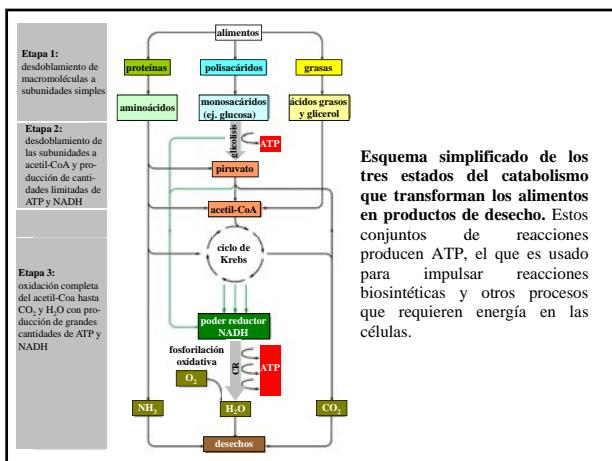
Chan D.C. 2006 *Annu. Rev. Cell. Dev. Biol.* 22:79-99

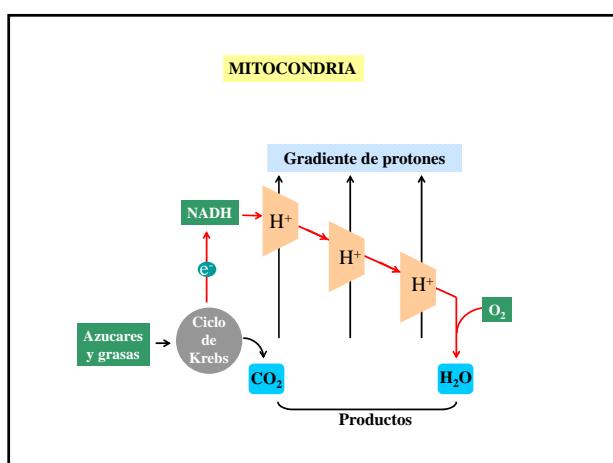
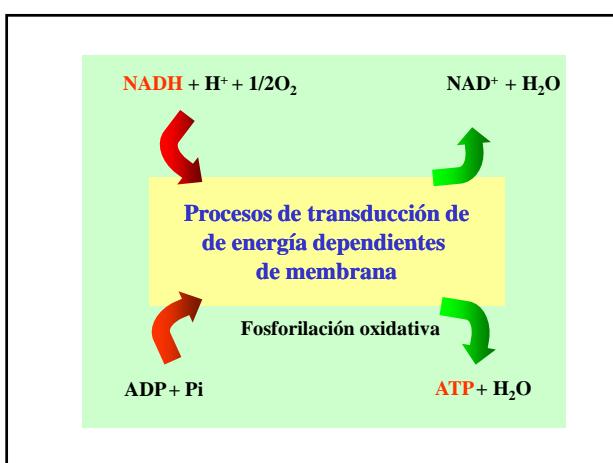
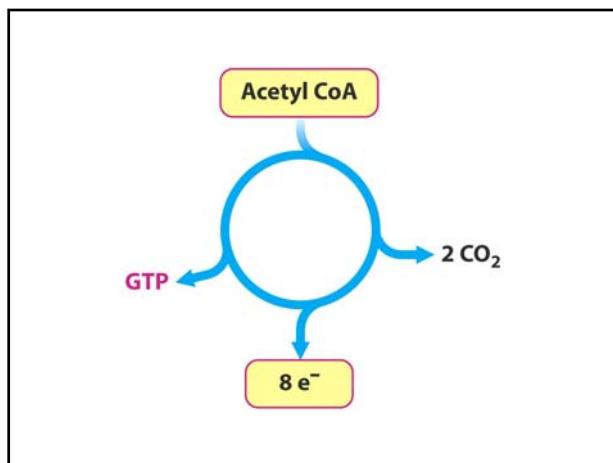
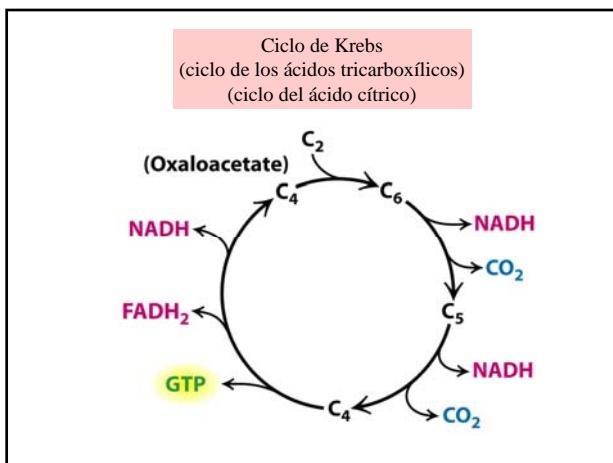
Mitochondrial dynamics plays important roles in vertebrate development and programmed cell death. Mutations in the mitochondrial fusion machinery lead to two human neurodegenerative disorders, Charcot-Marie-Tooth subtype 2A and autosomal dominant optic atrophy.

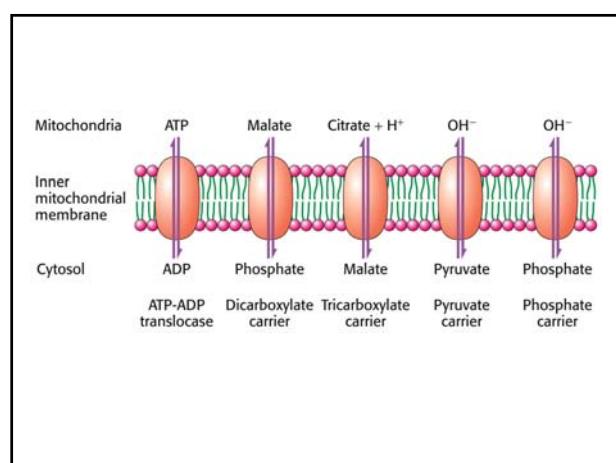
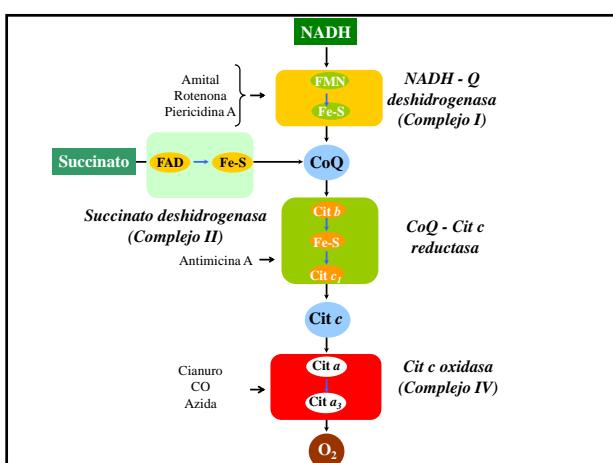
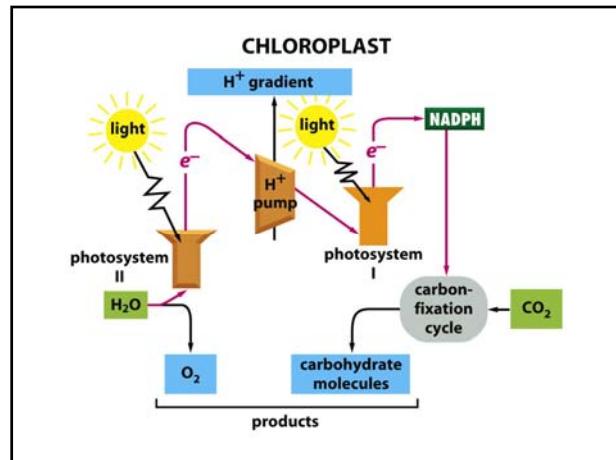
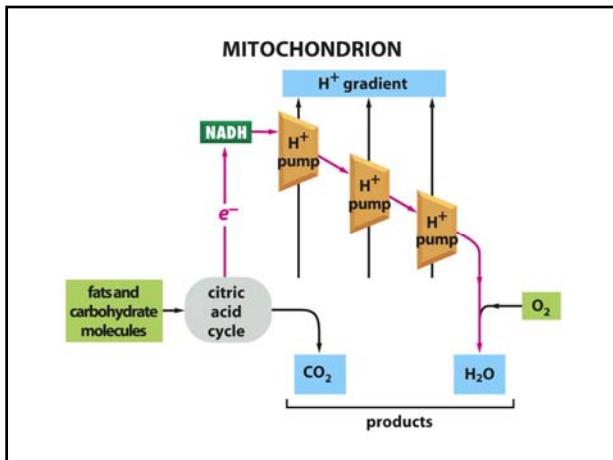
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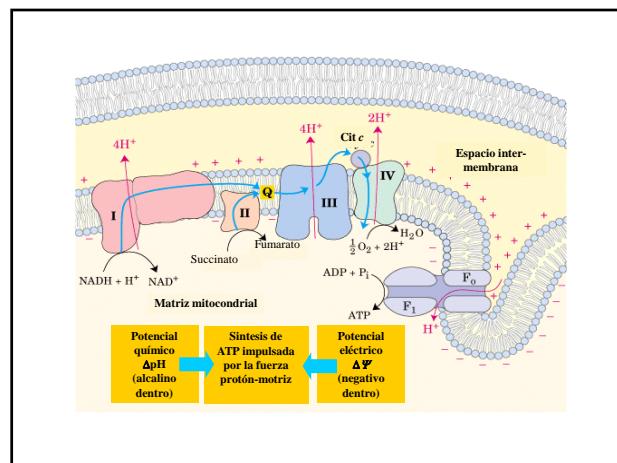
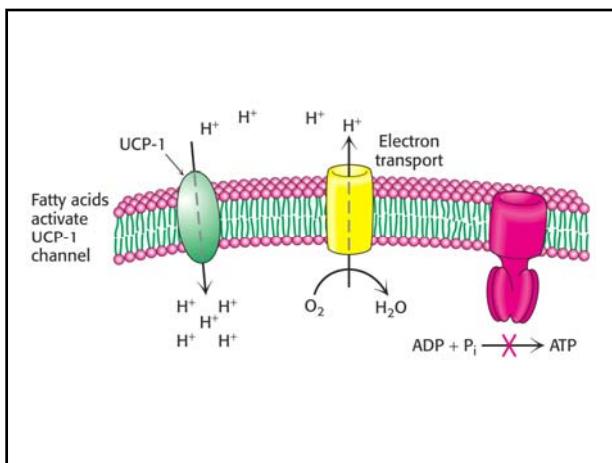












Coupling of Phosphorylation to Electron and Hydrogen Transfer by a Chemiosmotic Type of Mechanism

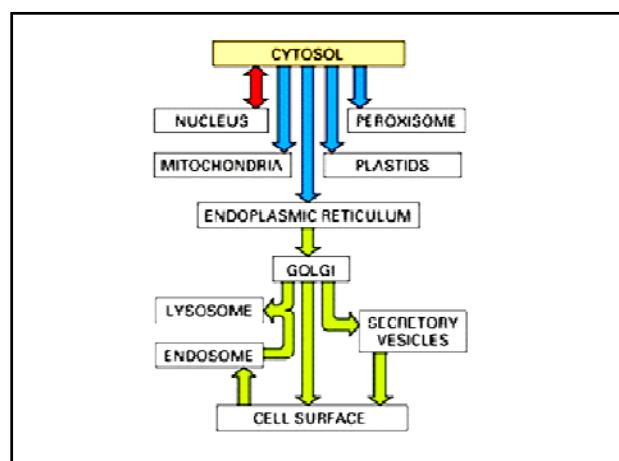
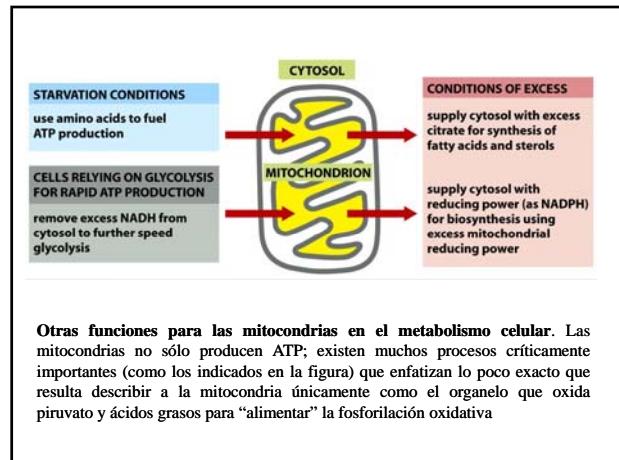
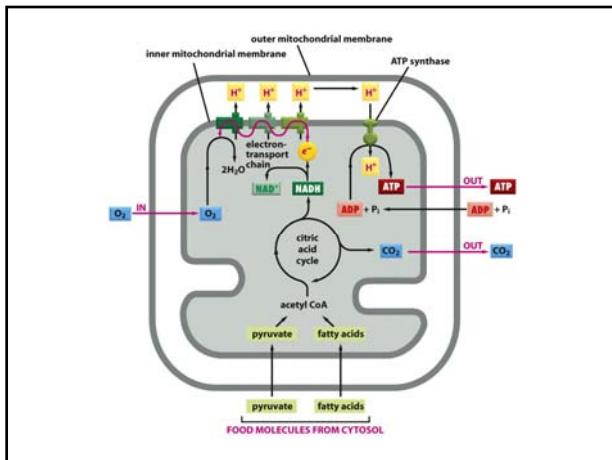
Peter Mitchell, University of Edinburgh, Edinburgh, Scotland

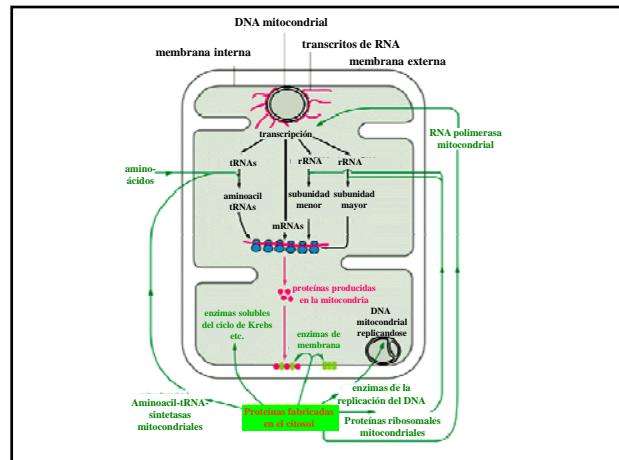
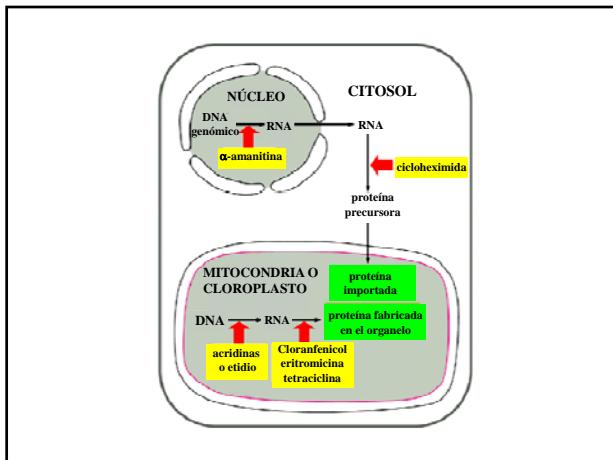
In the exact sciences, cause and effect are no more than events linked in sequence. Biochemists now generally accept the idea that metabolism is the cause of membrane transport. The underlying thesis of the hypothesis put forward here is that if the processes that we call metabolism and transport represent events in a sequence, not only can metabolism be the cause of transport, but also transport can be the cause of metabolism.

Nature, 1961, Volume 191, pages 144-148

Mitchell's Nobel Prize Lecture, in 1978, began as follows:

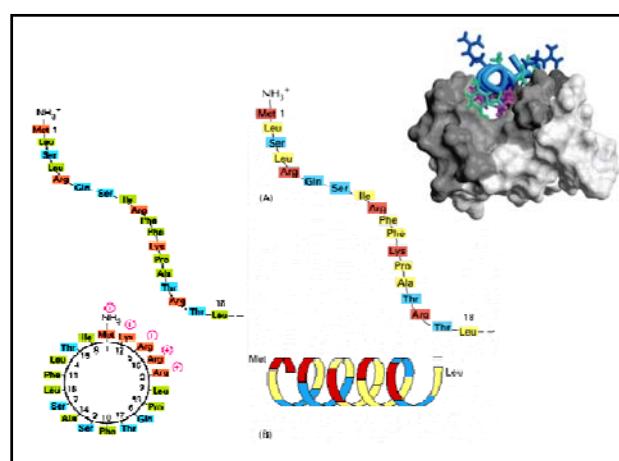
Although I had hoped that the chemiosmotic rationale of vectorial metabolism and biological energy transfer might one day come to be generally accepted, it would have been presumptuous of me to expect it to happen. Was it not Max Planck who remarked that a new scientific idea does not triumph by convincing its opponents, but rather because its opponents eventually die? The fact that what began as the chemiosmotic hypothesis has now been acclaimed as the chemiosmotic theory . . . has therefore both astonished and delighted me, particularly because those who were formerly my most capable opponents are still in the prime of their scientific lives.

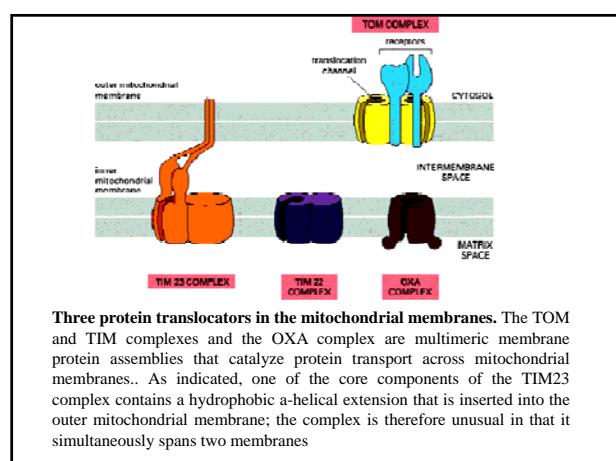
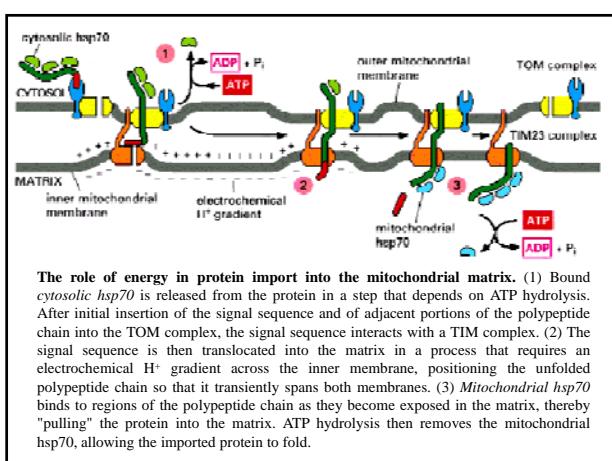
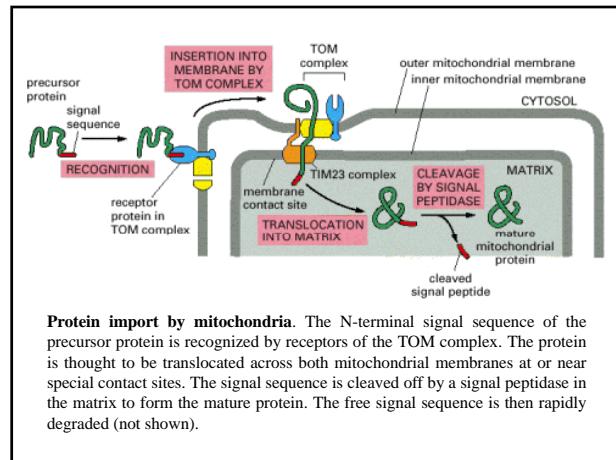
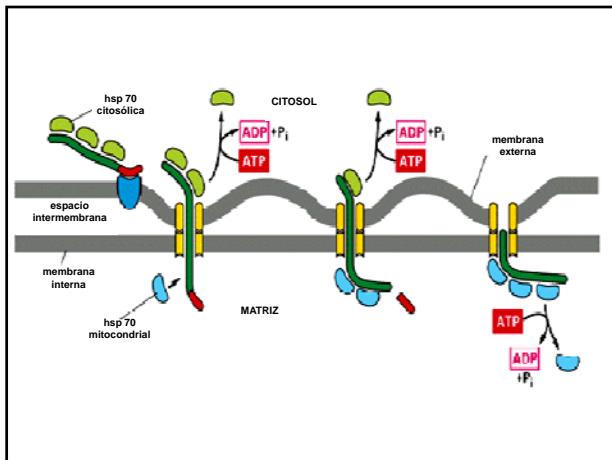


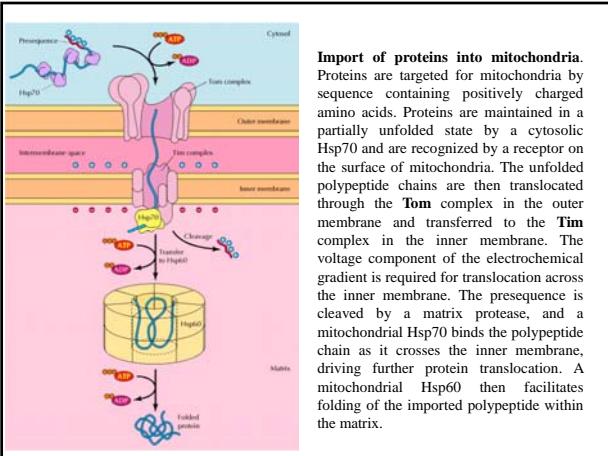


Function of Signal Peptide	Example of Signal Peptide
Import into ER	-H ₃ N-Met-Met-Ser-Phe-Val-Ser- Leu-Leu-Leu-Val Gly-Ile-Leu-Pho-Trp-Ala -Thr-Glu-Ala-Glu- Gln-Leu-Thr Lys Cys Glu Val-Phe-Gln
Requin in lumen of ER	-Lys-Asp-Glu-Leu-COO-
Import into mitochondria	-H ₃ N-Met-Leu-Ser-Leu-Arg-Gln-Ser-His-Arg-Phe- Phe-Lys-Pro-Ala-Thr-Arg-Thr-Leu-Cys-Ser- Ser-Arg-Tyr-Leu-Leu-
Import into nucleus	-Pro-Pro-Lys-Lys-Lys-Arg-Lys-Val-
Import into peroxisomes	-Ser-Lys-Leu-
Attach to membranes via the covalent linkage of a myristic acid to the amino terminus	-H ₃ N-Gly-Ser-Ser-Lys-Ser-Lys-Pro-Lys-

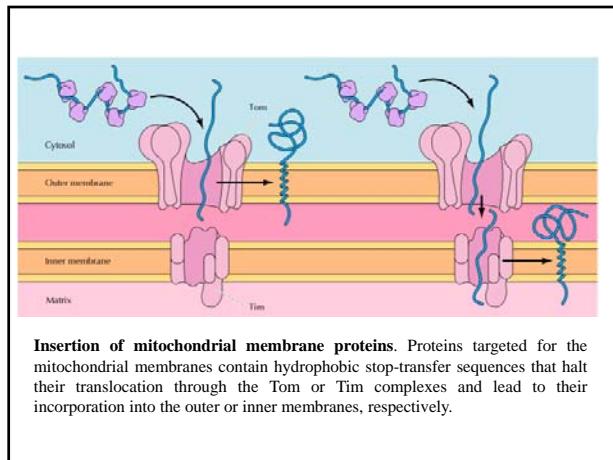
Notes:
 Positively charged amino acids are shown in red and negatively charged amino acids in green.
 An extended block of hydrophobic amino acids is enclosed in a yellow box.
 NH₃⁺ indicates the amino terminus of a protein; COOH indicates the carboxyl terminus.



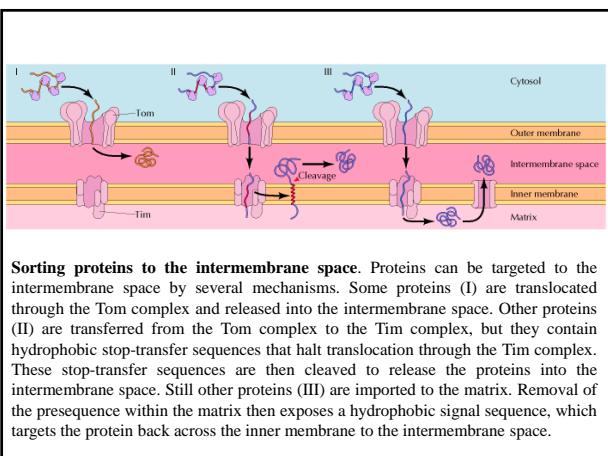




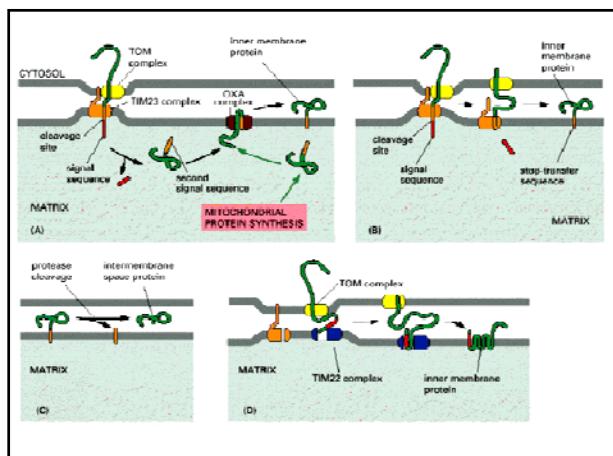
Import of proteins into mitochondria. Proteins are targeted for mitochondria by sequence containing positively charged amino acids. Proteins are maintained in a partially unfolded state by a cytosolic Hsp70 and are recognized by a receptor on the surface of mitochondria. The unfolded polypeptide chains are then translocated through the Tom complex in the outer membrane and transferred to the Tim complex in the inner membrane. The voltage component of the electrochemical gradient is required for translocation across the inner membrane. The presequence is cleaved by a matrix protease, and a mitochondrial Hsp60 binds the polypeptide chain as it crosses the inner membrane, driving further protein translocation. A mitochondrial Hsp60 then facilitates folding of the imported polypeptide within the matrix.

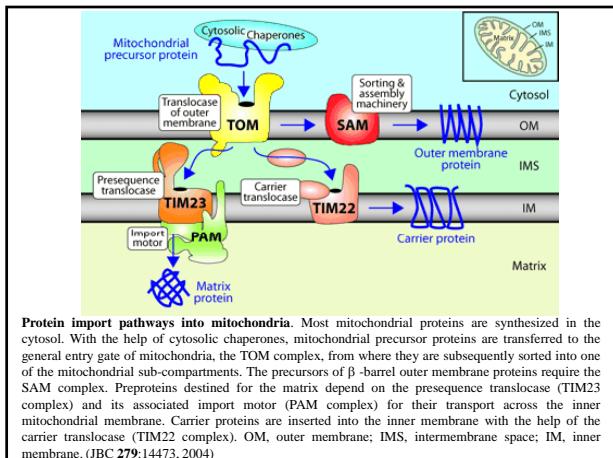


Insertion of mitochondrial membrane proteins. Proteins targeted for the mitochondrial membranes contain hydrophobic stop-transfer sequences that halt their translocation through the Tom or Tim complexes and lead to their incorporation into the outer or inner membranes, respectively.



Sorting proteins to the intermembrane space. Proteins can be targeted to the intermembrane space by several mechanisms. Some proteins (I) are translocated through the Tom complex and released into the intermembrane space. Other proteins (II) are transferred from the Tom complex to the Tim complex, but they contain hydrophobic stop-transfer sequences that halt translocation through the Tim complex. These stop-transfer sequences are then cleaved to release the proteins into the intermembrane space. Still other proteins (III) are imported to the matrix. Removal of the presequence within the matrix then exposes a hydrophobic signal sequence, which targets the protein back across the inner membrane to the intermembrane space.





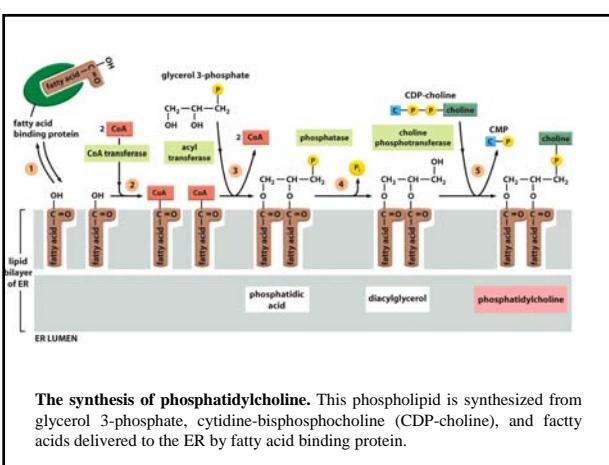
La translocación de proteínas a través de las membranas mitocondriales es mediada por complejos proteicos compuestos por múltiples subunidades que funcionan como translocadores de proteínas

El complejo TOM (Translocase of Outer Membrane) funciona a través de la membrana externa y el complejo TIM (Translocase of Inner Membrane) funciona a través de la membrana interna. Algunos componentes de estos complejos actúan como receptores de las proteínas a importar, y otros forman el canal de translocación

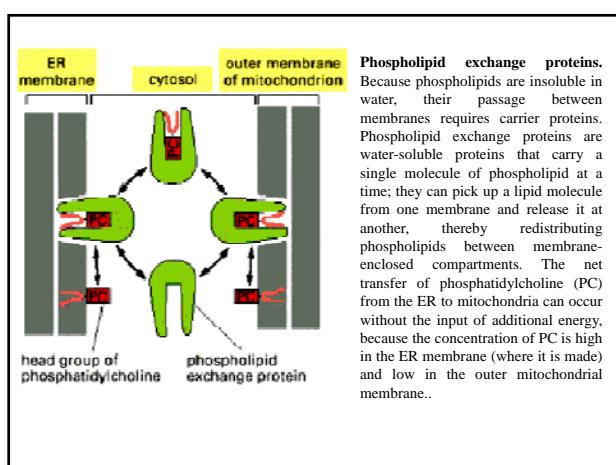
El complejo TOM se requiere para la importación de todas las proteínas codificadas en el genoma nuclear y ayuda a insertar proteínas transmembrana en la membrana externa. El complejo TIM23 transporta proteínas a la matriz mitocondrial y ayuda a insertar proteínas en la membrana interna

El complejo TIM22 media la inserción de una subclase de proteínas de la membrana interna que incluye una proteína que transporta ATP, ADP y fosfato

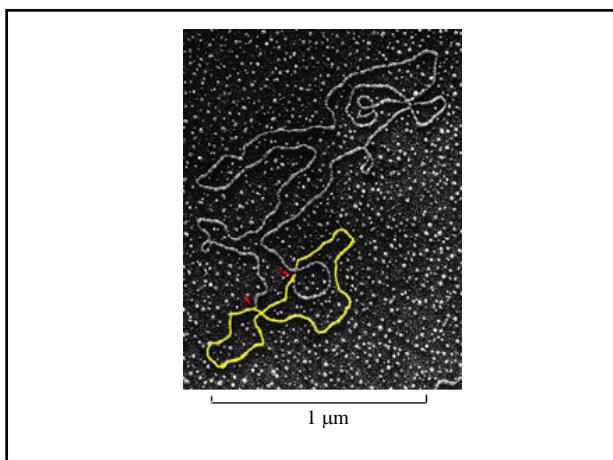
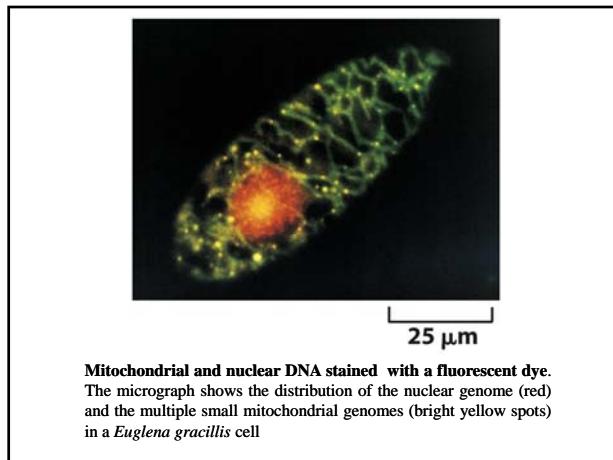
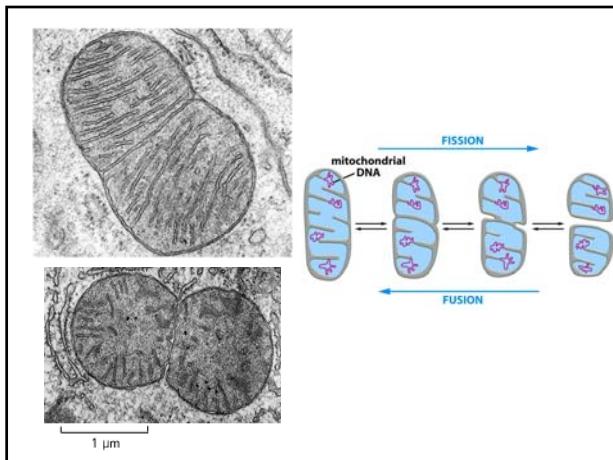
El complejo OXA, ubicado en la membrana interna, media la inserción de proteínas sintetizadas dentro de la mitocondria en la membrana interna. Además ayuda a insertar proteínas que han sido transportadas a la matriz previamente por los complejos TOM y TIM



The synthesis of phosphatidylcholine. This phospholipid is synthesized from glycerol 3-phosphate, cytidine-bisphosphocholine (CDP-choline), and fatty acids delivered to the ER by fatty acid binding protein.



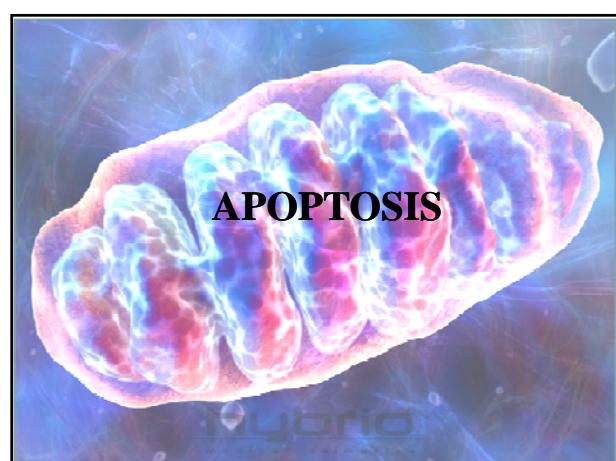
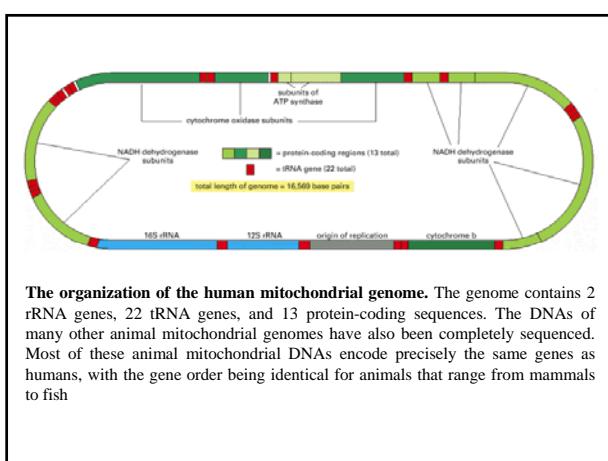
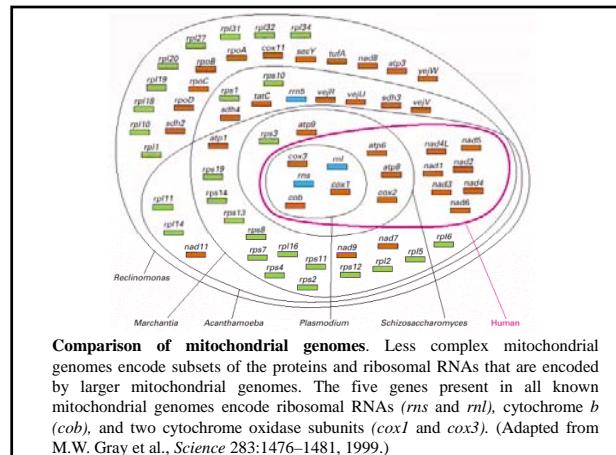
Phospholipid exchange proteins. Because phospholipids are insoluble in water, their passage between membranes requires carrier proteins. Phospholipid exchange proteins are water-soluble proteins that carry a single molecule of phospholipid at a time; they can pick up a lipid molecule from one membrane and release it at another, thereby redistributing phospholipids between membrane-enclosed compartments. The net transfer of phosphatidylcholine (PC) from the ER to mitochondria can occur without the input of additional energy, because the concentration of PC is high in the ER membrane (where it is made) and low in the outer mitochondrial membrane..

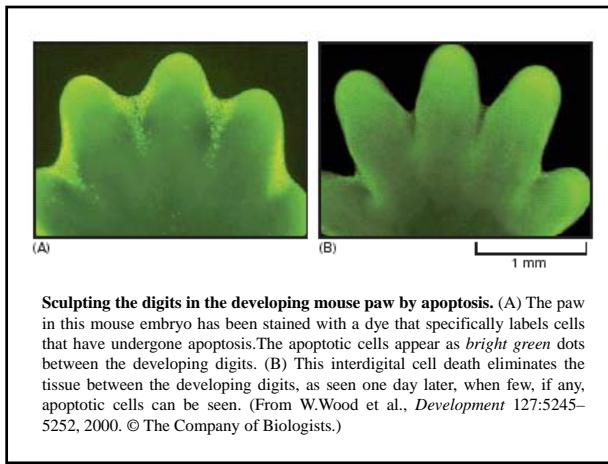


Tamaño del genoma de los organelos		Tamaño (kbp)
DNA de cloroplastos		
Plantas superiores	120-200	
<i>Chlamydomonas</i> (alga verde)	180	
DNA de mitocondrias		
Animales (gusanos planos,insectos, mamíferos)	16-19	
Plantas superiores	150-2500	
Hongos		
<i>Schizosaccharomyces pombe</i>	17	
<i>Aspergillus nidulans</i>	32	
<i>Neurospora crassa</i>	60	
<i>Saccharomyces cerevisiae</i>	78	
<i>Chlamydomonas</i> (alga verde)*	16	
Protozoos		
<i>Trypanosoma brucei</i>	22	
<i>Paramecium</i> *	40	
* moléculas lineales		

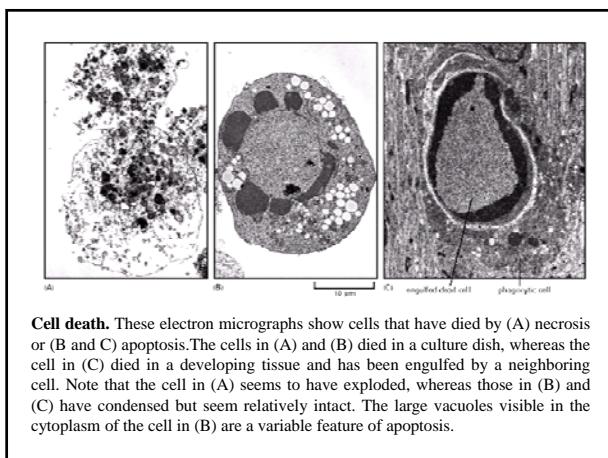
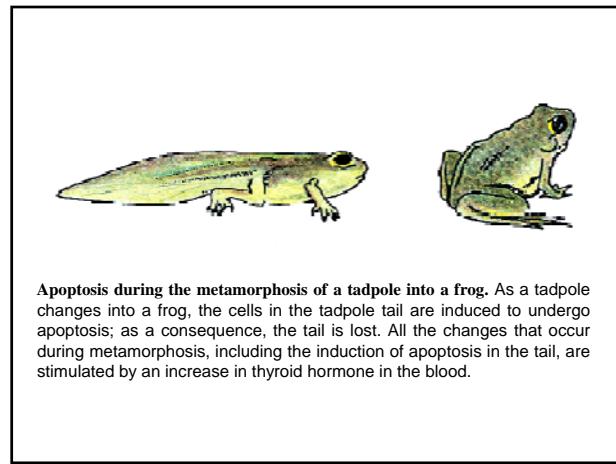
Cantidades relativas de DNA de organelos en algunos tejidos y tipos celulares				
Organismo	Tejido o tipo celular	Moléculas de DNA por organelo	Número de organelos por célula	DNA del organelo como % del DNA total
DNA de mitocondrias				
Rata	hígado	5-10	1000	1
Levadura*	vegetativa	2-50	1-50	15
<i>Xenopus laevis</i>	ocito	5-10	10^7	99
DNA de cloroplastos				
<i>Chlamydomonas</i>	vegetativa	80	1	7
Maíz	hojas	20-40	20-40	15

* la gran variación en el número y tamaño se debe a fragmentación y fusión mitocondrial

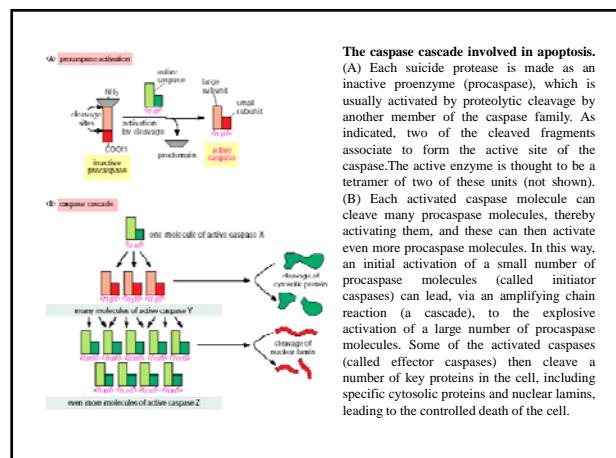




Sculpting the digits in the developing mouse paw by apoptosis. (A) The paw in this mouse embryo has been stained with a dye that specifically labels cells that have undergone apoptosis. The apoptotic cells appear as bright green dots between the developing digits. (B) This interdigital cell death eliminates the tissue between the developing digits, as seen one day later, when few, if any, apoptotic cells can be seen. (From W.Wood et al., *Development* 127:5245–5252, 2000. © The Company of Biologists.)

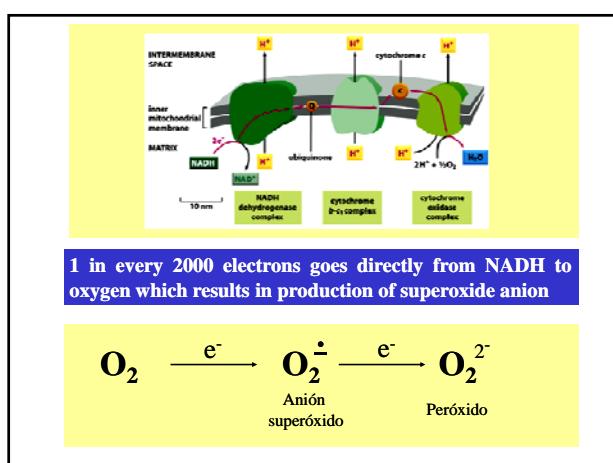
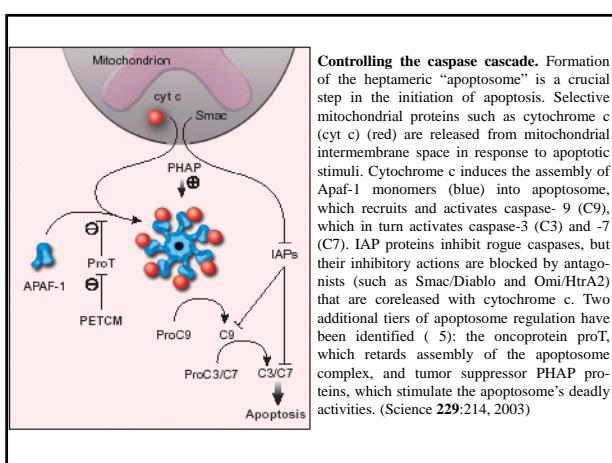
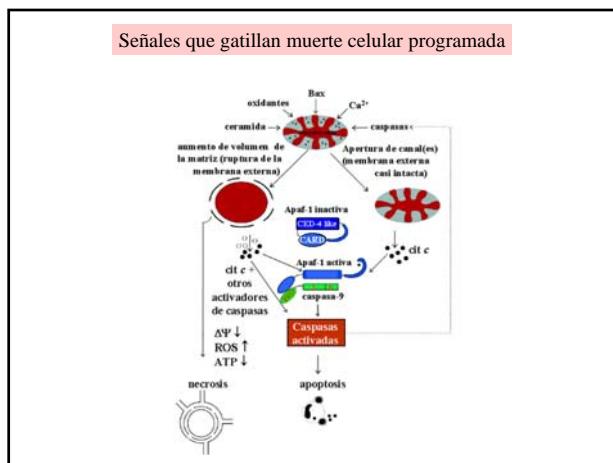
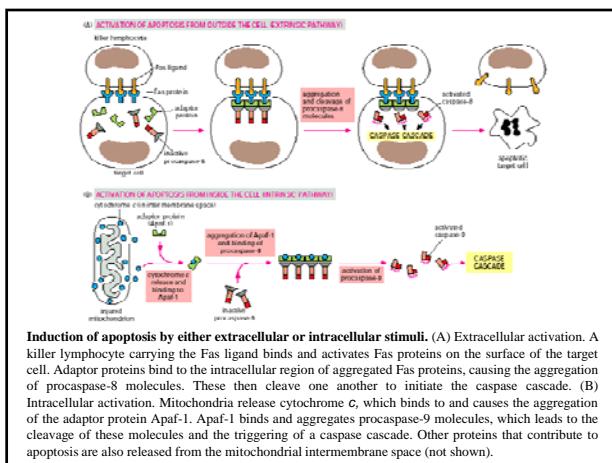


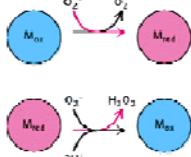
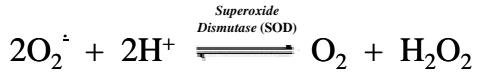
Cell death. These electron micrographs show cells that have died by (A) necrosis or (B and C) apoptosis. The cells in (A) and (B) died in a culture dish, whereas the cell in (C) died in a developing tissue and has been engulfed by a neighboring cell. Note that the cell in (A) seems to have exploded, whereas those in (B) and (C) have condensed but seem relatively intact. The large vacuoles visible in the cytoplasm of the cell in (B) are a variable feature of apoptosis.



The caspase cascade involved in apoptosis.

(A) Each suicide protease is made as an inactive proenzyme (procaspase), which is usually activated by proteolytic cleavage by another member of the caspase family. As indicated, two of the cleaved fragments associate to form the active site of the caspase. The active enzyme is thought to be a tetramer of two of these units (not shown). (B) Each activated caspase molecule can cleave many procaspase molecules, thereby activating them, and these can then activate even more procaspase molecules. In this way, an initial activation of a small number of procaspase molecules (called initiator caspases) can lead, via an amplifying chain reaction (a cascade), to the explosive activation of a large number of procaspase molecules. Some of the activated caspases (called effector caspases) then cleave a number of key proteins in the cell, including specific cytosolic proteins and nuclear lamins, leading to the controlled death of the cell.





Superoxide Dismutase Mechanism. The oxidized form of superoxide dismutase (Mox) reacts with one superoxide ion to form O₂ and generate the reduced form of the enzyme (Mred). The reduced form then reacts with a second superoxide and two protons to form hydrogen peroxide and regenerate the oxidized form of the enzyme.

Dismutation A reaction in which a single reactant is converted into two different products.

